

SCANNED, # 8

1 2 FEB 2002

FORM PTO-1390 (Modified)
(REV 5-93)

U.S. DEPARTMENT OF COMMERCE PATENT AND TRADEMARK OFFICE

**TRANSMITTAL LETTER TO THE UNITED STATES
DESIGNATED/ELECTED OFFICE (DO/EO/US)
CONCERNING A FILING UNDER 35 U.S.C. 371**

ATTORNEY'S DOCKET NUMBER

065691-0270

INTERNATIONAL APPLICATION NO.
PCT FR00/02319

INTERNATIONAL FILING DATE
08/11/2000

U.S. APPLICATION NO. (if known, see 37 C.F.R. 1.52)
Unassigned

107049372

PRIORITY DATE CLAIMED
08/12/1999

TITLE OF INVENTION

Odorant-Binding Human Proteins Fixing Hydrophobic Ligands: Polypeptides and Polynucleotides Coding for said polypeptides and Uses Thereof

APPLICANT(S) FOR DO/EO/US

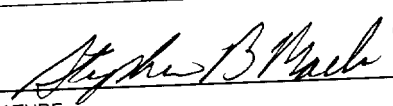

Gilles PITIOT, Eric LACAZETTE and Francoise GACHON

Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:

1. ☒ This is a FIRST submission of items concerning a filing under 35 U.S.C. 371.
 2. ☐ This is a SECOND or SUBSEQUENT submission of items concerning a filing under 35 U.S.C. 371.
 3. ☐ This express request to begin national examination procedures (35 U.S.C. 371(f)) at any time rather than delay examination until the expiration of the applicable time limit set in 35 U.S.C. 371(b) and PCT Articles 22 and 39(1).
 4. ☒ A proper Demand for International Preliminary Examination was made by the 19th month from the earliest claimed priority date.
 5. ☒ A copy of the International Application as filed (35 U.S.C. 371(c)(2))
 - ☐ is transmitted herewith (required only if not transmitted by the International Bureau).
 - ☒ has been transmitted by the International Bureau.
 - ☐ is not required, as the application was filed in the United States Receiving Office (RO/US)
 6. ☐ A translation of the International Application into English (35 U.S.C. 371(c)(2)).
 7. ☒ Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371(c)(3))
 - ☐ are transmitted herewith (required only if not transmitted by the International Bureau).
 - ☐ have been transmitted by the International Bureau.
 - ☐ have not been made; however, the time limit for making such amendments has NOT expired.
 - ☒ have not been made and will not be made.
 8. ☐ A translation of the amendments to the claims under PCT Article 19 (35 U.S.C. 371(c)(3)).
 9. ☐ An oath or declaration of the inventor(s) (35 U.S.C. 371(c)(4)).
 10. ☐ A translation of the annexes to the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371(c)(5)).
 11. ☐ Applicant claims small entity status under 37 CFR 1.27.
- Items 12. to 17. below concern other document(s) or information included:
12. ☐ An Information Disclosure Statement under 37 CFR 1.97 and 1.98.
 13. ☐ An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included.
 14. ☐ A FIRST preliminary amendment.
 - ☐ A SECOND or SUBSEQUENT preliminary amendment.
 15. ☐ A substitute specification.
 16. ☐ A change of power of attorney and/or address letter.
 17. ☒ Other items or information: Application Data Sheet, Paper Copy of Sequence Listing, and French Language Patent Application Specification

SCANNED, # 8

JC13 Rec'd PCT/PTO 12 FEB 2002

U.S. APPLICATION NO. (If known, see 37 C.F.R. 1.50) Unassigned		INTERNATIONAL APPLICATION NO. PCT/FR00/02319		ATTORNEY'S DOCKET NUMBER 065691-0270	
18. <input checked="" type="checkbox"/> The following fees are submitted:				CALCULATIONS	
				PTO USE ONLY	
Basic National Fee (37 CFR 1.492(a)(1)-(5): Search Report has been prepared by the EPO or JPO.....\$890.00					
International preliminary examination fee paid to USPTO (37 CFR 1.482).....\$710.00					
No international preliminary examination fee paid to USPTO (37 CFR 1.482) but international search fee paid to USPTO (37 CFR 1.445(a)(2))\$740.00					
Neither international preliminary examination fee (37 CFR 1.482) nor International search fee (37 CFR 1.445(a)(2)) paid to USPTO \$1,040.00					
International preliminary examination fee paid to USPTO (37 CFR 1.482) and all claims satisfied provisions of PCT Article 33(2)-(4)\$100.00					
ENTER APPROPRIATE BASIC FEE AMOUNT =				\$890.00	
Surcharge of \$130.00 for furnishing the oath or declaration later than 20 Months from the earliest claimed priority date (37 CFR 1.492(e))					
Claims	Number Filed	Included in Basic Fee	Extra Claims	Rate	
Total Claims	51	20	31	\$18.00	\$558.00
Independent Claims	5	3	2	\$84.00	\$168.00
Multiple dependent claim(s) (if applicable)				\$280.00	\$280.00
TOTAL OF ABOVE CALCULATIONS =					\$1896.00
Reduction by 1/2 for filing by small entity, if applicable.					\$0.00
SUBTOTAL =					\$1896.00
Processing fee of \$130.00 for furnishing English translation later the 20 months from the earliest claimed priority date (37 CFR 1.492(f)).				+	
TOTAL NATIONAL FEE =					\$1896.00
Fee for recording the enclosed assignment (37 CFR 1.21(h)). The assignment must be accompanied by an appropriate cover sheet (37 CFR 3.28, 3.31). \$40.00 per property +				+	
TOTAL FEES ENCLOSED =					\$1896.00
				Amount to be: refunded \$	
				charged \$	
<p>a. <input checked="" type="checkbox"/> A check in the amount of \$1896.00 to cover the above fees is enclosed.</p> <p>b. <input type="checkbox"/> Please charge my Deposit Account No. <u>19-0741</u> in the amount of \$0.00 to the above fees. A duplicate copy of this sheet is enclosed.</p> <p>c. <input checked="" type="checkbox"/> The Commissioner is hereby authorized to charge any additional fees which may be required, or credit any overpayment to Deposit Account No. <u>19-0741</u>. A duplicate copy of this sheet is enclosed.</p>					
<p>NOTE: Where an appropriate time limit under 37 CFR 1.494 or 1.495 has not been met, a petition to revive (37 CFR 1.137(a) or (b)) must be filed and granted to restore the application to pending status.</p>					
SEND ALL CORRESPONDENCE TO:					
Foley & Lardner Customer Number: 22428			 SIGNATURE		
 22428			NAME STEPHEN B. MAEBIUS		
PATENT TRADEMARK OFFICE			REGISTRATION NUMBER 35,264		

PTO/PCT Rec'd

15 AUG 2002

#6

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Atty. Docket No: 065691/0270

In re patent application of

PITOT, GILLES et al.

Serial No. 10/049,372

Filed: February 12, 2002

For: HUMAN ODORANT-BINDING PROTEINS WHICH BIND HYDROPHOBIC LIGANDS:
POLYPEPTIDES AND POLYNUCLEOTIDES ENCODING SAID POLYPEPTIDES, AND USES
THEREOF

STATEMENT TO SUPPORT FILING AND SUBMISSION IN
ACCORDANCE WITH 37 C.F.R. §§ 1.821-1.825

Assistant Commissioner for Patents
Washington, D.C. 20231
Box SEQUENCE

Sir:

In connection with a Sequence Listing submitted concurrently herewith, the undersigned hereby states that:

1. the submission, filed herewith in accordance with 37 C.F.R. § 1.821(g), does not include new matter;

2. the content of the attached paper copy and the attached computer readable copy of the Sequence Listing, submitted in accordance with 37 C.F.R. § 1.821(c) and (e), respectively, are the same; and

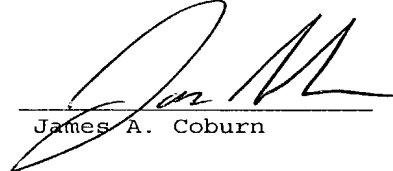
3. all statements made herein of their own knowledge are true and that all statements made on information and belief are believed to be true; and further, that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United

Serial No. 10/049,372

States Code and that such willful false statements may jeopardize the validity of the application or any patent resulting therefrom.

Respectfully submitted,

August 6, 2002
Date


James A. Coburn

HARBOR CONSULTING
Intellectual Property Services
1500A Lafayette Road
Suite 262
Portsmouth, N.H.
800-318-3021

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Application No. :

U.S. National Serial No. :

Filed :

PCT International Application No. : PCT/FR00/02319

VERIFICATION OF A TRANSLATION

I, the below named translator, hereby declare that:

My name and post office address are as stated below;

That I am knowledgeable in the French language in which the below identified international application was filed, and that, to the best of my knowledge and belief, the English translation of the amended sheets of the international application No. PCT/FR00/02319 is a true and complete translation of the amended sheets of the above identified international application as filed.

I hereby declare that all the statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the patent application issued thereon.

Date: February 28, 2002

Full name of the translator :


Elaine Patricia PARRISH

For and on behalf of RWS Group plc

Post Office Address :

Europa House, Marsham Way,
Gerrards Cross, Buckinghamshire,
England.

PTO/PCT Rec'd

15 AUG 2002

Atty. Dkt. No. 065691/0270

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Applicants: Gilles PITIOT et al.

Title: HUMAN ODORANT-BINDING PROTEINS WHICH BIND HYDROPHOBIC LIGANDS:
POLYPEPTIDES AND POLYNUCLEOTIDES ENCODING SAID POLYPEPTIDES,
AND USES THEREOF

Appl. No. 10/049,372

Filing Date: 08/15/2002

Examiner: Unassigned

Art Unit: Unassigned

AMENDMENT IN RESPONSE TO NOTICE UNDER 37 CFR §§1.821-825

Commissioner for Patents
Washington, D.C. 20231

Sir:

In response to the Notice to Comply With Requirements for Applications Containing Sequence Disclosures mailed April 16, 2002, please amend the application as follows:

In the Specification:

Please amend the specification as shown:

Please delete the paragraph on page 6, lines 28-30, and replace it with the following paragraph:

The polypeptides according to the invention are characterized in that they preferably comprise the Gly-Thr-Trp-Tyr (SEQ ID NO: 25) domain.

Please delete the paragraph on page 33, lines 6-15, and replace it with the following paragraph:

The upper lines represent the hOBPIIa sequences (SEQ ID NOS 17-19, 21-24) and the lower lines represent the hOBPIIb sequence (SEQ ID NO: 20) for which only the different nucleotides are represented; a dash indicates the absence of corresponding sequences. The shaded capitals are the exon sequences and the lower case letters are the intron sequences. The sizes indicated on the left are indicated in bp. The TATA box is in bold characters and the polyadenylation signal is underlined. The boxes indicate the splice acceptor sites for exons 5, 5b and 5c.

Please delete the paragraph on page 33, lines 35-39 thru page 34 lines 1-4, and replace it with the following paragraph:

FIGURE 5: Alignment of the protein sequences derived from the two human genes hOBPIIa and hOBPIIb (hOBPIIa α = OBP2aaHOMSA (SEQ ID NO: 2), hOBPIIb α = OBP2baHOMSA (SEQ ID NO: 10), hOBPIIb β = OBP2bbHOMSA (residues 15-179 of SEQ ID NO: 12), hOBPIIa γ = OBP2agHOMSA (SEQ ID NO: 6), hOBPIIa β = OBP2abHOMSA (SEQ ID NO: 4)), of human tear lipocalins (LCN1_HOMSA) (SEQ ID NO: 46), of rat OBPII (OBP2_RATNO) (SEQ ID NO: 47), of bovine lactoglobulin BLG (LACB_BOSTA) (SEQ ID NO: 48), of mouse MUP (MUP6_MUSMU) (SEQ ID NO: 49), of human RBP (RBP_HOMSA) (SEQ ID NO: 50), of bovine OBP (OBP_BOSTA) (SEQ ID NO: 51), of rat MUP (MUP_RATNO) (SEQ ID NO: 52) and of porcine OBP (OBP_SUSSC) (SEQ ID NO: 53).

Please delete the paragraph on page 34, lines 6-16 and replace it with the following paragraph:

The residues in the dark grey boxes are identical and those in the light gray boxes are similar. The elements of secondary structure predicted with the DSC program are underlined and the amino acid residues are in italics. The β -sheets and the α -helices are

numbered for hOPBIIa and b. The anticipated signal peptide cleavage site is indicated with an arrow (AAA↓LS) at position 15. Nonaligned sequences of very divergent forms of spliced genes, hOPBa6 (OPB2adHOMSA) (SEQ ID NO: 8) and hOBPby (OBP2bgHOMSA) (SEQ ID NO: 14) have been added at the bottom of the alignment after the analysis.

Please delete the paragraph on page 39, lines 30-38 thru page 40, lines 1-2, and replace it with the following paragraph:

A Lambda gt11 human testes cDNA library (Clontech) (10^7 p.f.u.) was amplified by 30 polymerase chain reaction (PCR) cycles (94°C 45 sec, 54°C 45 sec, 72°C 1 min 30 sec) with the primer oliEST58 CCTGCAGGTACATGAGCTTCC (SEQ ID NO: 26) and 5' or 3' screening amplimers for inserts located on the arms of the lambda gt11 vectors. A nested PCR was then carried out with oliEST26 CGCTGTATTTGCCAGGCTCC (SEQ ID NO: 27) and oligonucleotides specific for the arm of the vector. The PCR products were subcloned into the pGEM-T (r) vector, which made it possible to obtain the 5' end of the hOBPII gene cDNAs.

Please delete the paragraph on page 40, lines 20-39 thru page 41, lines 1-5, and replace it with the following paragraph:

Tissue samples were collected from 45 to 55 year old Caucasian individuals, in agreement with the French regulations in force. The total RNA is extracted according to a single-step method using the RNA NOW[®] reagent, according to the manufacturer's (Biogentex) instructions. 5 µg of total RNA were reversed-transcribed in a final volume of 20 µl using 0.5 ng of oligonucleotide GACTCGAGTCGACATCGATTTTTTTTTTTTTTTT (SEQ ID NO: 28) with the Superscript[®] preamplification system (Gibco BRL). Three µl of these reactions were then used for the following PCRs. The expression of the specific mRNAs was determined by PCR using: the primers TL: CCTCTCCCAGCCCCAGCAAG (SEQ ID NO: 29) and AP: GACTCGAGTCGACATCG (SEQ

ID NO: 30) for the LCN1-type genes (LCN1, LCN1b, LCN1c) and, for the hOBPII-type genes, the primers DE: CGCCCAGTGACCTGCCGAGGTC (SEQ ID NO: 31), and FI: CTTTATTTGGAGTCAGGTGGGTG (SEQ ID NO: 32). As a quality control for the RNA's the primers G3PDH1: CTCTGCCCCCTCTGCTGATG (SEQ ID NO: 33) and G3PDH2: CCTGCTTCACCACCTTCTTG (SEQ ID NO: 34) for the G3PDH gene were used; the G3PDH gene is considered to be constitutively expressed in all cell types. 32 PCR cycles (94°C 45 sec, 54°C 45 sec, 72°C 2 min 30 sec) were performed and the amplification products were separated on 1% agarose gel. The DNA was transferred onto a Hybond N+® membrane.

Please delete the paragraph on page 41, lines 7-16, and replace it with the following paragraph:

For the detection of the expression of the various genes, several oligonucleotides specific for the respective genes were synthesized:

- oLCN1: GACTCAGACTCCGGAGATGA (SEQ ID NO: 35),
- oLCN1b: AACTCAGACACCAGAGATGA (SEQ ID NO: 36),
- oLCN1c: GACTCAGATCCCGGAGATGA (SEQ ID NO: 37),
- oI5: CCAGGAGGGACCACTACA (SEQ ID NO: 38) specific for the hOBPIIb gene,
- oI4: CCGGGACGGACGACTACG (SEQ ID NO: 39) specific for the hOBPIIa gene,
- G3PDH3: CTCATGACCACAGTCCATGC (SEQ ID NO: 40).

Please delete the paragraph on page 41, lines 30-39 thru page 42, lines 1-7, and replace it with the following paragraph:

The genotyping is carried out with PCR reactions using 100 ng of genomic DNA originating from 8 CEPH reference families, and using the oligonucleotides oli9 TGTTCGGGAACGCAGCTT (SEQ ID NO: 41) and

oli10a TGCCGCTGTCCCCACGTCGG (SEQ ID NO: 42). The thermocycler parameters consist of an initial cycle at 94°C for 10 min followed by 30 cycles at 94°C for 30 sec, 55°C for 30 sec and 70°C for 45 sec, and then a final elongation step of 10 min at 70°C. The PCR products are then analyzed on a 3% agarose gel. The information regarding the markers for chromosome 9 may be obtained at the following internet address (<http://galton.ucl.ac.uk>); the analyses were carried out using the linkage study tools previously described in Lacazette *et al.* (1997). The haplotypes are reconstructed manually according to the recombination events previously described in family 1362 (Attwood *et al.*, 1994).

Please delete the paragraph on page 52, lines 34-39 thru page 54, lines 1-11, and replace it with the following paragraph:

In the case of the hOBPIIb gene, in addition to the hOBPIIb α mRNA previously described, a 106 bp super-numerary exon (exon 3b) between the previous exons 3 and 4 was identified (figure 3). This longer mRNA (782 nucleotides) encodes a 165 amino acid protein hOBPIIb β . From a protein structure point of view, hOBPIIb β is identical to hOBPIIb α up to the 5th putative β -sheet and then differs due to a reading frame shift. The predictions from the computer programs indicate that the motif ALWEALAI DTRLK (SEQ ID NO: 43) is an α -helix which is just behind the fifth β -sheet. Two additional β -sheets may be present in the long C-terminal portion.

Please delete the paragraph on page 58, lines 36-39 thru page 59, lines 1-23, and replace it with the following paragraph:

A PCR on the plasmid DNA of an hOBPIIb α clone using the primers BIIa/b (5' GTC GGA TCC CTG TCC TTC ACC CTG GAG G 3') (SEQ ID NO: 44), a sense oligonucleotide beginning 45 bases after the protein-initiating ATG, and XIIb (5' GTC CTC GAG GTG TTC GGG AAC GCA GCT TC 3') (SEQ ID NO: 45), an antisense oligonucleotide preceding the stop

codon of the hOBPIIb α protein, made it possible to amplify all of the DNA encoding the secreted hOBPIIb α protein. The BanH I and Xho I enzymatic restriction sites located at the ends of the two oligonucleotides (bases underlined) were used for directional cloning into a plasmid expression vector pGEX-6Pl, followed by transformation by electroporation (1800 V, 200 Ω , 25 μ F) into bacterial strain BL21. Synthesis of the recombinant protein is obtained by adding IPTG, at 5 mM final for 3 h, to 250 ml of culture of the strain in LB medium containing 100 μ g/ml of ampicillin pre-incubated at 37°C for 2 h. The centrifuged cultures are taken up in 25 ml of TENG buffer. The lysate is sonicated and recentrifuged. The fusion protein is then purified using 4 ml of beads bearing covalently attached insoluble glutathione (Sigma) per 25 ml of supernatant. After incubation for 4 h, they are washed with 3 volumes of 1M NaCl and then with 10 volumes of 1 X PBS. The elution is obtained by bringing the beads into contact with a glutathione solution (10 mM reduced glutathione, 50 mM Tris-HCl, pH 8.0) for 10 min.

REMARKS

Applicants believe that the present application is now in condition for allowance. Favorable reconsideration of the application as amended is respectfully requested.

The Examiner is invited to contact the undersigned by telephone if it is felt that a telephone interview would advance the prosecution of the present application.

Respectfully submitted,

Date August 14, 2002

By 

FOLEY & LARDNER
Customer Number: 22428

Stephen Maebius
Attorney for Applicants
Registration No. 35,264



22428

PATENT TRADEMARK OFFICE

Telephone: (202) 672-5407
Facsimile: (202) 672-5399

Should additional fees be necessary in connection with the filing of this paper, or if a petition for extension of time is required for timely acceptance of same, the Commissioner is hereby authorized to charge Deposit Account No.19-0741 for any such fees; and applicant(s) hereby petition for any needed extension of time.

MARKED UP VERSION ATTACHED TO AMENDMENT IN

SERIAL NO. 10/049,372

Marked up version of the paragraph on page 6, lines 28-30 is below:

The polypeptides according to the invention are characterized in that they preferably comprise the Gly-Thr-Trp-Tyr (SEQ ID NO: 25) domain.

Marked up version of the paragraph on page 33, lines 6-15, is below:

The upper lines represent the hOBPIIa sequences (SEQ ID NOS 17-19, 21-24) and the lower lines represent the hOBPIIb sequence (SEQ ID NO: 20) for which only the different nucleotides are represented; a dash indicates the absence of corresponding sequences. The shaded capitals are the exon sequences and the lower case letters are the intron sequences. The sizes indicated on the left are indicated in bp. The TATA box is in bold characters and the polyadenylation signal is underlined. The boxes indicate the splice acceptor sites for exons 5, 5b and 5c.

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RNA NOW[®] reagent, according to the manufacturer's (Biogentex) instructions. 5 µg of total RNA were reversed-transcribed in a final volume of 20 µl using 0.5 ng of oligonucleotide GACTCGAGTCGACATCGATTTTTTTTTTTTTTTTTT (SEQ ID NO: 28) with the Superscript[®] preamplification system (Gibco BRL). Three µl of these reactions were then used for the following PCRs. The expression of the specific mRNAs was determined by PCR using: the primers TL: CCTCTCCCAGCCCCAGCAAG (SEQ ID NO: 29) and AP: GACTCGAGTCGACATCG (SEQ ID NO: 30) for the LCN1-type genes (LCN1, LCN1b, LCN1c) and, for the hOBPII-type genes, the primers DE: CGCCCAGTGACCTGCCGAGGTC (SEQ ID NO: 31), and FI: CTTTATTTGGAGTCAGGTGGGTG (SEQ ID NO: 32). As a quality control for the RNA's the primers G3PDH1: CTCTGCCCCCTCTGCTGATG (SEQ ID NO: 33) and G3PDH2: CCTGCTTCACCACCTTCTTG (SEQ ID NO: 34) for the G3PDH gene were used; the G3PDH gene is considered to be constitutively expressed in all cell types. 32 PCR cycles (94°C 45 sec, 54°C 45 sec, 72°C 2 min 30 sec) were performed and the amplification products were separated on 1% agarose gel. The DNA was transferred onto a Hybond N+[®] membrane.

Marked up version of the paragraph on page 41, lines 6-16, is below:

For the detection of the expression of the various genes, several oligonucleotides specific for the respective genes were synthesized:

- olLCN1 : GACTCAGACTCCGGAGATGA (SEQ ID NO: 35),
- olLCN1b: AACTCAGACACCAGAGATGA (SEQ ID NO: 36),
- olLCN1c: GACTCAGATCCCGGAGATGA (SEQ ID NO: 37),
- ol5: CCAGGAGGGACCACTACA (SEQ ID NO: 38) specific for the hOBPIIb gene,
- ol4: CCGGGACGGACGACTACG (SEQ ID NO: 39) specific for the hOBPIIa gene,

- G3PDH3: CTCATGACCACAGTCCATGC (SEQ ID NO: 40).

Marked up version of the paragraph on page 41, lines 30-39 thru page 42, lines 1-7, is below:

The genotyping is carried out with PCR reactions using 100 ng of genomic DNA originating from 8 CEPH reference families, and using the oligonucleotides oli9 TGTTCGGGAACGCAGCTT (SEQ ID NO: 41) and oli10a TGCCGCTGTCCCCACGTCGG (SEQ ID NO: 42). The thermocycler parameters consist of an initial cycle at 94°C for 10 min followed by 30 cycles at 94°C for 30 sec, 55°C for 30 sec and 70°C for 45 sec, and then a final elongation step of 10 min at 70°C. The PCR products are then analyzed on a 3% agarose gel. The information regarding the markers for chromosome 9 may be obtained at the following internet address (<http://galton.ucl.ac.uk>); the analyses were carried out using the linkage study tools previously described in Lacazette *et al.* (1997). The haplotypes are reconstructed manually according to the recombination events previously described in family 1362 (Attwood *et al.*, 1994).

Marked up version of the paragraph on page 52, lines 34-39 thru page 54, lines 1-11, is below:

In the case of the hOBPIIb gene, in addition to the hOBPIIb α mRNA previously described, a 106 bp super-numerary exon (exon 3b) between the previous exons 3 and 4 was identified (figure 3). This longer mRNA (782 nucleotides) encodes a 165 amino acid protein hOBPIIb β . From a protein structure point of view, hOBPIIb β is identical to hOBPIIb α up to the 5th putative β -sheet and then differs due to a reading frame shift. The predictions from the computer programs indicate that the motif ALWEALAI DTRLK (SEQ ID NO: 43) is an α -helix which is just behind the fifth β -sheet. Two additional β -sheets may be present in the long C-terminal portion.

Marked up version of the paragraph on page 58, lines 36-39 thru page 59, lines 1-23, is below:

A PCR on the plasmid DNA of an hOBPIIb α clone using the primers BIIa/b (5' GTC GGA TCC CTG TCC TTC ACC CTG GAG G 3') (**SEQ ID NO: 44**), a sense oligonucleotide beginning 45 bases after the protein-initiating ATG, and XIIb (5' GTC CTC GAG GTG TTC GGG AAC GCA GCT TC 3') (**SEQ ID NO: 45**), an antisense oligonucleotide preceding the stop codon of the hOBPIIb α protein, made it possible to amplify all of the DNA encoding the secreted hOBPIIb α protein. The BanH I and Xho I enzymatic restriction sites located at the ends of the two oligonucleotides (bases underlined) were used for directional cloning into a plasmid expression vector pGEX-6Pl, followed by transformation by electroporation (1800 V, 200 Ω , 25 μ F) into bacterial strain BL21. Synthesis of the recombinant protein is obtained by adding IPTG, at 5 mM final for 3 h, to 250 ml of culture of the strain in LB medium containing 100 μ g/ml of ampicillin pre-incubated at 37°C for 2 h. The centrifuged cultures are taken up in 25 ml of TENG buffer. The lysate is sonicated and recentrifuged. The fusion protein is then purified using 4 ml of beads bearing covalently attached insoluble glutathione (Sigma) per 25 ml of supernatant. After incubation for 4 h, they are washed with 3 volumes of 1M NaCl and then with 10 volumes of 1 X PBS. The elution is obtained by bringing the beads into contact with a glutathione solution (10 mM reduced glutathione, 50 mM Tris-HCl, pH 8.0) for 10 min.

CLAIMS

1. An isolated polypeptide comprising an amino acid sequence having at least 90% identity with the amino acid sequences SEQ ID No. 2, SEQ ID No. 4, SEQ ID No. 6, SEQ ID No. 8, SEQ ID No. 12 or SEQ ID No. 14.
2. An isolated polypeptide, characterized in that it comprises a polypeptide chosen from:
- a) a polypeptide of sequence SEQ ID No. 2, SEQ ID No. 4, SEQ ID No. 6, SEQ ID No. 8, SEQ ID No. 12 or SEQ ID No. 14;
 - b) a polypeptide which is a variant of a polypeptide of amino acid sequences defined in a);
 - c) a polypeptide homologous to the polypeptide defined in a) or b) and comprising at least 90% identity with said polypeptide of a);
 - d) a fragment of at least 15 consecutive amino acids of a polypeptide defined in a), b) or c);
 - e) a biologically active fragment of a polypeptide defined in a), b) or c).
3. An isolated polypeptide selected from a polypeptide corresponding to the sequence SEQ ID No. 2 and named OBPII_{αα}, to the sequence SEQ ID No. 4 and named OBPII_{αβ}, to the sequence SEQ ID No. 6 and named OBPII_{αγ}, and to the sequence SEQ ID No. 12 and named OBPII_{ββ}.
4. The polypeptide as claimed in any one of claims 1 to 3, characterized in that it comprises at least the Gly-Thr-Trp-Tyr domain.

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10. The use of a polynucleotide as claimed in any one of claims 5 to 8, as a primer for amplifying or polymerizing nucleic acid sequences.

5

11. The use of a polynucleotide as claimed in claim 8, as a sense or antisense oligonucleotide for controlling the expression of the corresponding protein product.

10

12. A recombinant vector for cloning a polynucleotide as claimed in one of claims 5 to 8 and/or for expressing a polypeptide as claimed in one of claims 1 to 4, characterized in that it contains a polynucleotide as claimed in any one of claims 5 to 8.

15

13. The vector as claimed in claim 12, characterized in that it comprises the elements which allow the expression, optionally the secretion, of said polypeptide in a host cell.

20

14. The vector as claimed in any one of claims 12 to 13, characterized in that the elements which allow the expression of said polypeptide are chosen from:

25

a) the isolated polynucleotide of sequence SEQ ID No. 15 and SEQ ID No. 16;

30

b) a polynucleotide, the sequence of which is complementary to the sequence of the polynucleotide defined in a);

35

c) a polynucleotide, the sequence of which comprises at least 80% identity with a polynucleotide defined in a) or in b);

d) a polynucleotide which hybridizes, under conditions of high stringency, with a sequence of the polynucleotide defined in a), b) or c).

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15. The vector as claimed in claims 13 and 14, for expression in eukaryotic cells, selected from viral DNA and naked DNA.

5

16. A host cell, characterized in that it is transformed with a vector as claimed in one of claims 12 to 15.

10 17. A process for preparing a recombinant polypeptide, characterized in that a host cell as claimed in claim 16 is cultured under conditions which allow the expression and, optionally, the secretion of said recombinant polypeptide, and in that said recombinant
15 polypeptide is recovered.

18. A recombinant polypeptide which can be obtained using a process as claimed in claim 17.

20 19. The use of a polypeptide chosen from a polypeptide as claimed in any one of claims 1 to 4 and 18, or a fragment thereof, as a protein which binds to a hydrophobic ligand, preferably an odorous molecule.

25 20. The use of a polypeptide as claimed in any one of claims 1 to 4 and 18, or of the polypeptide hOBPII_{ba} of sequence SEQ ID No. 10, a polypeptide which is a variant of said polypeptide hOBPII_{ba}, a polypeptide homologous to said polypeptide hOBPII_{ba} comprising at
30 least 90% identity with said polypeptide hOBPII_{ba} or a fragment thereof, as a competitive inhibitor, as an agonist or as an antagonist of the cellular receptors for lipocalins.

35 21. A monoclonal or polyclonal antibody, and fragments thereof, characterized in that it is specifically against an isolated polypeptide as claimed in one of claims 1 to 4 and 18.

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22. The use of an antibody as claimed in claim 21, for demonstrating the presence of a polypeptide as claimed in one of claims 1 to 4 and 18 in a biological sample.

5

23. A process for detecting an antibody against hOBPII in human serum from an allergic and/or asthmatic patient, using an hOBPII polypeptide.

10

24. A process for controlling the volatilization of an odorant, characterized in that it comprises a step of binding of said odorant with a polypeptide as claimed in any one of claims 1 to 4 and 18, or with the polypeptide hOBPII_{ba} of sequence SEQ ID No. 10, a polypeptide which is a variant of said polypeptide hOBPII_{ba} or a polypeptide homologous to said polypeptide hOBPII_{ba} comprising at least 90% identity with said polypeptide hOBPII_{ba}.

15

20

25. The process as claimed in claim 24, characterized in that the polypeptide is bound to a solid support.

26. The process as claimed in claim 24, characterized in that the polypeptide is in a liquid composition.

25

27. The process as claimed in claim 26, characterized in that said composition is a fragranced composition for the skin.

30

28. A process for screening a molecule, preferably odorants or flavors, which comprises passing the molecule over a substrate which comprises a polypeptide as claimed in any one of claims 1 to 4 and 18, or the polypeptide hOBPII_{ba} of sequence SEQ ID No. 10, a polypeptide which is a variant of said polypeptide hOBPII_{ba} or a polypeptide homologous to said polypeptide hOBPII_{ba} comprising at least 90% identity with said polypeptide hOBPII_{ba}, bound to said substrate, said polypeptide

35

35. The polypeptide as claimed in claim 34, characterized in that said polypeptide is expressed in the form of a protein from fusion with a protein which
5 allows specific cellular addressing.

36. The polypeptide as claimed in claim 35, characterized in that said protein which allows specific cellular addressing is chosen from the group composed
10 of interleukins, of cytokines, of lymphokines, of interferons, of growth factors, of hormones and of antibodies.

37. The polypeptide as claimed in claim 34, characterized in that said polypeptide is combined with a
15 molecule which allows specific cellular addressing.

38. The polypeptide as claimed in claim 37, characterized in that said molecule which allows specific
20 cellular addressing is chosen from the group composed of steroids, of interleukins, of cytokines, of lymphokines, of interferons, of growth factors, of hormones and of antibodies.

39. The polypeptide as claimed in any one of claims 1 to 4 and 18, or the polypeptide hOBPII_{ba} of sequence SEQ ID No. 10, a polypeptide which is a variant of said polypeptide hOBPII_{ba} or a polypeptide homologous to said polypeptide hOBPII_{ba} comprising at least 90% identity
30 with said polypeptide hOBPII_{ba}, as a pharmaceutical compound transporter.

40. A pharmaceutical composition comprising a pharmaceutical compound bound at least to a polypeptide as
35 claimed in one of claims 34 to 39, and a pharmaceutically acceptable vehicle.

41. The pharmaceutical composition as claimed in
AMENDED SHEET

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claim 40, characterized in that the pharmaceutical compound is chosen from the group of anticancer agents.

42. The pharmaceutical composition as claimed in claim 41, characterized in that said anticancer agent is a radioactive isotope chosen from the group: iodine¹³¹, yttrium⁹⁰, gold¹⁹⁹, palladium¹⁰⁰, copper⁶⁷, bismuth²¹⁷ and antimony²¹¹.

43. The pharmaceutical composition as claimed in any one of claims 40 to 42, characterized in that said polypeptide as claimed in any one of claims 34 to 39 constitutes a delayed form of delivery of said pharmaceutical compound in the body.

44. A pharmaceutical composition comprising an expression vector as claimed in claim 12 or 13, and a pharmaceutically acceptable vehicle.

45. The pharmaceutical composition as claimed in any one of claims 40 to 44, for treating cancer preferably chosen from breast cancer, uterine cancer, prostate cancer, liver cancer and pulmonary epithelial cell carcinoma.

46. The use of a polypeptide chosen from the polypeptide hOBPII_{ba} of sequence SEQ ID No. 10, a polypeptide which is a variant of said polypeptide hOBPII_{ba} and a polypeptide homologous to said polypeptide hOBPII_{ba} comprising at least 90% identity with said polypeptide hOBPII_{ba}, for preparing a medicinal product intended for the treatment of uterine cancer, prostate cancer, liver cancer and pulmonary epithelial cell cancer.

47. The use of the polypeptide as claimed in any one of claims 1 to 4 and 18, or the polypeptide hOBPII_{ba} of sequence SEQ ID No. 10, or a polypeptide which is a

variant of said polypeptide hOBPII_{ba} or a polypeptide homologous to said polypeptide hOBPII_{ba} comprising at least 90% identity with said polypeptide hOBPII_{ba}, for preparing a medicinal product intended for the transport of a compound across the placental barrier.

48. The use as claimed in claim 47, characterized in that said transport is carried out from the gestating mother to the fetus, and in that said compound is chosen from hormones, essential fatty acids, lipophilic medicinal products and vitamins.

49. The use as claimed in claim 47, characterized in that said transport is carried out from the fetus to the mother and is intended for the detoxification of the fetus.

50. The polypeptide as claimed in claims 1 to 4 and 18, or the polypeptide hOBPII_{ba} of sequence SEQ ID No. 10, a polypeptide which is a variant of said polypeptide hOBPII_{ba} or a polypeptide homologous to said polypeptide hOBPII_{ba} comprising at least 90% identity with said polypeptide hOBPII_{ba}, as a pregnancy marker.

51. The polypeptide as claimed in claims 1 to 4 and 18, or the polypeptide hOBPII_{ba} of sequence SEQ ID No. 10, a polypeptide which is a variant of said polypeptide hOBPII_{ba} or a polypeptide homologous to said polypeptide hOBPII_{ba} comprising at least 90% identity with said polypeptide hOBPII_{ba}, as a marker for a fetoplacental pathological condition

SEQUENCE LISTING

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LACAZETTE, ERIC
GACHON, FRANCOISE

<120> HUMAN ODORANT-BINDING PROTEINS WHICH BIND HYDROPHOBIC
LIGANDS: POLYPEPTIDES AND POLYNUCLEOTIDES ENCODING SAID
POLYPEPTIDES, AND USES THEREOF

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gtttggaaag ccactgggga ggacaggagc ggggacacgg cgtcagggct gcagcagtgg 11760
ggccaccgca gggctcccg ctcagggggtc tcagagggat cctccaagct cccccatttt 11820
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<210> 17
<211> 253
<212> DNA
<213> Homo sapiens

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tgctcccagg aactggcca gggggctata aagaacatct cgagaggagc cagcacagcc 120
ttgttcagac gccagtgac ctgccgaggt cggcagcaca gagctctgga gatgaagacc 180
ctgttcctgg gtgtcacgct cggcctggcc gctgccctgt ccttcacctt ggaggaggag 240
gatgtgagct ggg 253

```

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<210> 18
<211> 154
<212> DNA
<213> Homo sapiens

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cttgaagcc acgttcacct tcatgtgagt gttg 154

```

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agcctggcaa atacagcgcc tgtgagcccc t 91

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ggggag 126

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22

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ggtgaggggc c 131

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<211> 188
<212> DNA
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aggatcgg 188

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<211> 44
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<210> 24
<211> 130
<212> DNA
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ccaaataaag 130

<210> 25
<211> 4
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: Synthetic peptide

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23

<210> 26
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 <212> DNA
 <213> Artificial Sequence

<220>
 <223> Description of Artificial Sequence: Primer

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<210> 27
 <211> 20
 <212> DNA
 <213> Artificial Sequence

<220>
 <223> Description of Artificial Sequence: Primer

<400> 27
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<210> 28
 <211> 35
 <212> DNA
 <213> Artificial Sequence

<220>
 <223> Description of Artificial Sequence: Synthetic
 oligonucleotide

<400> 28
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<210> 29
 <211> 20
 <212> DNA
 <213> Artificial Sequence

<220>
 <223> Description of Artificial Sequence: Primer

<400> 29
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<210> 30
 <211> 17
 <212> DNA
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<220>
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<210> 36
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<210> 38
 <211> 18
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26

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20

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18

<210> 42
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20

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 1 5 10

<210> 44
 <211> 28

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1				5					10					15		
Gln	Ala	His	His	Leu	Leu	Ala	Ser	Asp	Glu	Glu	Ile	Gln	Asp	Val	Ser	
			20					25					30			
Gly	Thr	Trp	Tyr	Leu	Lys	Ala	Met	Thr	Val	Asp	Arg	Glu	Phe	Pro	Glu	
		35					40					45				
Met	Asn	Leu	Glu	Ser	Val	Thr	Pro	Met	Thr	Leu	Thr	Thr	Leu	Glu	Gly	
	50					55					60					
Gly	Asn	Leu	Glu	Ala	Lys	Val	Thr	Met	Leu	Ile	Ser	Gly	Arg	Cys	Gln	
65					70					75					80	
Glu	Val	Lys	Ala	Val	Leu	Glu	Lys	Thr	Asp	Glu	Pro	Gly	Lys	Tyr	Thr	
				85					90					95		
Ala	Asp	Gly	Gly	Lys	His	Val	Ala	Tyr	Ile	Ile	Arg	Ser	His	Val	Lys	
			100					105					110			
Asp	His	Tyr	Ile	Phe	Tyr	Cys	Glu	Gly	Glu	Leu	His	Gly	Lys	Pro	Val	
		115					120					125				
Arg	Gly	Val	Lys	Leu	Val	Gly	Arg	Asp	Pro	Lys	Asn	Asn	Leu	Glu	Ala	
	130					135					140					
Leu	Glu	Asp	Phe	Glu	Lys	Ala	Ala	Gly	Ala	Arg	Gly	Leu	Ser	Thr	Glu	
145					150					155					160	

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<400> 48
Met  Lys  Cys  Leu  Leu  Leu  Ala  Leu  Ala  Leu  Thr  Cys  Gly  Ala  Gln  Ala
  1              5              10              15

Leu  Ile  Val  Thr  Gln  Thr  Met  Lys  Gly  Leu  Asp  Ile  Gln  Lys  Val  Ala
      20              25              30

```

29

Gly Thr Trp Tyr Ser Leu Ala Met Ala Ala Ser Asp Ile Ser Leu Leu
 35 40 45
 Asp Ala Gln Ser Ala Pro Leu Arg Val Tyr Val Glu Glu Leu Lys Pro
 50 55 60
 Thr Pro Glu Gly Asp Leu Glu Ile Leu Leu Gln Lys Trp Glu Asn Gly
 65 70 75 80
 Glu Cys Ala Gln Lys Lys Ile Ile Ala Glu Lys Thr Lys Ile Pro Ala
 85 90 95
 Val Phe Lys Ile Asp Ala Leu Asn Glu Asn Lys Val Leu Val Leu Asp
 100 105 110
 Thr Asp Tyr Lys Lys Tyr Leu Leu Phe Cys Met Glu Asn Ser Ala Glu
 115 120 125
 Pro Glu Gln Ser Leu Ala Cys Gln Cys Leu Val Arg Thr Pro Glu Val
 130 135 140
 Asp Asp Glu Ala Leu Glu Lys Phe Asp Lys Ala Leu Lys Ala Leu Pro
 145 150 155 160
 Met His Ile Arg Leu Ser Phe Asn Pro Thr Gln Leu Glu Glu Gln Cys
 165 170 175

His Ile

<210> 49
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 <212> PRT
 <213> Murine sp.

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 His Ala Glu Glu Ala Ser Ser Thr Gly Arg Asn Phe Asn Val Glu Lys
 20 25 30
 Ile Asn Gly Glu Trp His Thr Ile Ile Leu Ala Ser Asp Lys Arg Glu
 35 40 45
 Lys Ile Glu Asp Asn Gly Asn Phe Arg Leu Phe Leu Glu Gln Ile His
 50 55 60
 Val Leu Glu Asn Ser Leu Val Leu Lys Phe His Thr Val Arg Asp Glu
 65 70 75 80
 Glu Cys Ser Glu Leu Ser Met Val Ala Asp Lys Thr Glu Lys Ala Gly
 85 90 95
 Glu Tyr Ser Val Thr Tyr Asp Gly Phe Asn Thr Phe Thr Ile Pro Lys
 100 105 110

30

Thr Asp Tyr Asp Asn Phe Leu Met Ala His Leu Ile Asn Glu Lys Asp
 115 120 125
 Gly Glu Thr Phe Gln Leu Met Gly Leu Tyr Gly Arg Glu Pro Asp Leu
 130 135 140
 Met Ser Asp Ile Lys Glu Arg Phe Ala Gln Leu Cys Glu Glu His Gly
 145 150 155 160
 Ile Leu Arg Glu Asn Ile Ile Asp Leu Ser Asn Ala Asn Arg Cys Leu
 165 170 175
 Gln Ala Arg Glu
 180

<210> 50
 <211> 199
 <212> PRT
 <213> Homo sapiens

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 1 5 10 15
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 20 25 30
 Lys Ala Arg Phe Ser Gly Thr Trp Tyr Ala Met Ala Lys Lys Asp Pro
 35 40 45
 Glu Gly Leu Phe Leu Gln Asp Asn Ile Val Ala Glu Phe Ser Val Asp
 50 55 60
 Glu Thr Gly Gln Met Ser Ala Thr Ala Lys Gly Arg Val Arg Leu Leu
 65 70 75 80
 Asn Asn Trp Asp Val Cys Ala Asp Met Val Gly Thr Phe Thr Asp Thr
 85 90 95
 Glu Asp Pro Ala Lys Phe Lys Met Lys Tyr Trp Gly Val Ala Ser Phe
 100 105 110
 Leu Gln Lys Gly Asn Asp Asp His Trp Ile Val Asp Thr Asp Tyr Asp
 115 120 125
 Thr Tyr Ala Val Gln Tyr Ser Cys Arg Leu Leu Asn Leu Asp Gly Thr
 130 135 140
 Cys Ala Asp Ser Tyr Ser Phe Val Phe Ser Arg Asp Pro Asn Gly Leu
 145 150 155 160
 Pro Pro Glu Ala Gln Lys Ile Val Arg Gln Arg Gln Glu Glu Leu Cys
 165 170 175
 Leu Ala Arg Gln Tyr Arg Leu Ile Val His Asn Gly Tyr Cys Asp Gly
 180 185 190


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<400> 52
Met Lys Leu Leu Leu Leu Leu Cys Leu Gly Leu Thr Leu Val Cys
  1                      5                      10                      15

Gly His Ala Glu Glu Ala Ser Ser Thr Arg Gly Asn Leu Asp Val Ala
  20                      25                      30

Lys Leu Asn Gly Asp Trp Phe Ser Ile Val Val Ala Ser Asn Lys Arg
  35                      40                      45

Glu Lys Ile Glu Glu Asn Gly Ser Met Arg Val Phe Met Gln His Ile
  50                      55                      60

```

Asp Val Leu Glu Asn Ser Leu Gly Phe Lys Phe Arg Ile Lys Glu Asn
65 70 75 80

Gly Glu Cys Arg Glu Leu Tyr Leu Val Ala Tyr Lys Thr Pro Glu Asp
85 90 95

Gly Glu Tyr Phe Val Glu Tyr Asp Gly Gly Asn Thr Phe Thr Ile Leu
100 105 110

Lys Thr Asp Tyr Asp Arg Tyr Val Met Phe His Leu Ile Asn Phe Lys
115 120 125

Asn Gly Glu Thr Phe Gln Leu Met Val Leu Tyr Gly Arg Thr Lys Asp
130 135 140

Leu Ser Ser Asp Ile Lys Glu Lys Phe Ala Lys Leu Cys Glu Ala His
145 150 155 160

Gly Ile Thr Arg Asp Asn Ile Ile Asp Leu Thr Lys Thr Asp Arg Cys
165 170 175

Leu Gln Ala Arg Gly
180

<210> 53
<211> 157
<212> PRT
<213> Sus scrofa

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1 5 10 15

Ile Thr Ser Tyr Ile Gly Ser Ser Asp Leu Glu Lys Ile Gly Glu Asn
20 25 30

Ala Pro Phe Gln Val Phe Met Arg Ser Ile Glu Phe Asp Asp Lys Glu
35 40 45

Ser Lys Val Tyr Leu Asn Phe Phe Ser Lys Glu Asn Gly Ile Cys Glu
50 55 60

Glu Phe Ser Leu Ile Gly Thr Lys Gln Glu Gly Asn Thr Tyr Asp Val
65 70 75 80

Asn Tyr Ala Gly Asn Asn Lys Phe Val Val Ser Tyr Ala Ser Glu Thr
85 90 95

Ala Leu Ile Ile Ser Asn Ile Asn Val Asp Glu Glu Gly Asp Lys Thr
100 105 110

Ile Met Thr Gly Leu Leu Gly Lys Gly Thr Asp Ile Glu Asp Gln Asp
115 120 125

[illegible]

33

Leu Glu Lys Phe Lys Glu Val Thr Arg Glu Asn Gly Ile Pro Glu Glu
130 135 140

Asn Ile Val Asn Ile Ile Glu Arg Asp Asp Cys Pro Ala
145 150 155

PTO/PCT Rec'd

15 AUG 2002

10/049372#3

WO 01/12806

PCT/FR00/02319

HUMAN ODORANT-BINDING PROTEINS WHICH BIND HYDROPHOBIC
LIGANDS: POLYPEPTIDES AND POLYNUCLEOTIDES ENCODING
SAID POLYPEPTIDES, AND APPLICATIONS THEREOF

- 5 The present invention relates to the demonstration of novel human odorant-binding proteins, hereafter named "OBPs", and also to the applications thereof, both therapeutically and non-therapeutically.
- 10 The present invention is based on the identification of a family of lipocalin genes, composed of three genes and two pseudogenes, on human chromosome 9q34; the three genes correspond to the LCN1 gene which has already been described and to two novel genes which are
- 15 the subject of the present invention and are called hOBPIIa and hOBPIIb. In addition, the invention is based on the attribution of novel functions to already known human lipocalins, through the demonstration of new areas of expression.
- 20 Although a certain number of OBPs have already been demonstrated (Pelosi et al., 1996), such as the rat OBP for example (see in particular patent EP-0 335 654), the OBPs according to the present invention are human
- 25 OBPs which have a very large number of advantages, as will emerge from the remainder of the text, compared to the murine proteins.
- 30 These OBP proteins of the lipocalin family have, as far as some of them are concerned, been mentioned indirectly in patent WO 99/07740, but their OBP function has never been described to date; the same is also true of the proteins LCN1, retinol-binding protein (RBP) and apolipoprotein D (ApoD), as will be more fully explained
- 35 hereinafter.

Historically, the lipocalin family (Pervaiz and Brew, 1987), has been defined based on the human retinol-

binding protein (RBP) and on 3 other proteins: bovine β -lactoglobulin, rat $\alpha 2\mu$ -globulin and human $\alpha 1$ -microglobulin. From these reference proteins, and using sequence homologies, the lipocalin family has increased

5 in size so as to now incorporate a large number of proteins, more than about 100, both in eukaryotes and in prokaryotes (Flower et al., 1995, 1996). This family consists of small proteins (160-190 amino acids) which contain a hydrophobic pocket and are generally secreted

10 (Bocskei et al., 1992; Senoo et al., 1990, Zeng et al., 1996; Miller, 1998), although, in certain cases, they are proteins which remain associated with the membrane (Nagata et al., 1991). In vertebrates, the sequence identities between the various lipocalins are in the

15 region of 20%; however, the sequence identities are greater for the orthologous proteins, and also for the recent paralogous genes described (Igarashi et al., 1992; Dewald et al., 1992).

20 The present invention relates to the isolated OBPII polypeptides encoded by the two new isolated human genes, OBPIIa and OBPIIb, located at the 9q34 locus; the invention also relates to the corresponding polynucleotide sequences, the corresponding mRNAs and also

25 the regulatory promoter sequences which determine the expression profile in the various tissues and in particular in the secretory tissues. These two genes encode at least seven different polypeptides given the existence of alternative splicing of the transcripts.

30 The OBPIIa gene encodes at least 4 different polypeptides, named OBPII $_{a\alpha}$, OBPII $_{a\beta}$, OBPII $_{a\gamma}$ and OBPII $_{a\delta}$, and the OBPIIb gene encodes three different polypeptides, named OBPII $_{b\alpha}$, OBPII $_{b\beta}$ and OBPII $_{b\delta}$. Even though one gene is mainly expressed, all the messenger forms are found

35 in the nasal structure.

The subject of the present invention is therefore an isolated polypeptide comprising an amino acid sequence having at least 90% identity with the amino acid

sequences SEQ ID No. 2, SEQ ID No. 4, SEQ ID No. 6, SEQ ID No. 8, SEQ ID No. 10, SEQ ID No. 12 or SEQ ID No. 14, and LCN1, ApoD and RBP.

- 5 It should be understood that the invention relates to the polypeptides obtained by purification from natural sources or obtained using recombinant techniques, as will be described subsequently; the invention also relates to the polypeptides obtained by chemical
10 synthesis, which may then comprise unnatural amino acids. In the present description, the term "polypeptide" will be used to denote equally a protein or a peptide.
- 15 The expression "percentage identity between the sequences" is intended to denote the percentage of amino acids which are identical between the sequences, obtained with the best possible sequence alignment, this percentage being purely statistical and the
20 differences between the polypeptides being distributed randomly and over the entire length.

The present invention also relates to an isolated polypeptide, characterized in that it comprises a polypeptide chosen from:

- 25 a) a polypeptide of sequence SEQ ID No. 2, SEQ ID No. 4, SEQ ID No. 6, SEQ ID No. 8, SEQ ID No. 10, SEQ ID No. 12 or SEQ ID No. 14;
- 30 b) a polypeptide which is a variant of a polypeptide of amino acid sequences defined in a);
- 35 c) a polypeptide homologous to the polypeptide defined in a) or b) and comprising at least 90% identity with said polypeptide of a);
- d) a fragment of at least 15 consecutive amino acids of a polypeptide defined in a), b) or c);

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e) a biologically active fragment of a polypeptide defined in a), b) or c).

5 The term "variant polypeptide" will be intended to mean all the mutated polypeptides which may exist naturally, in particular in humans, and which correspond in particular to truncations, substitutions, deletions and/or additions of amino acid residues. The variant
10 polypeptides according to the invention conserve at least one hydrophobic ligand-binding domain.

The term "homologous polypeptide" will be intended to denote the polypeptides exhibiting, compared to the
15 polypeptides of SEQ ID No. 2, SEQ ID No. 4, SEQ ID No. 6, SEQ ID No. 8, SEQ ID No. 10, SEQ ID No. 12 and SEQ ID No. 14, certain modifications, such as in particular a deletion, addition or substitution of at least one amino acid, a truncation, an extension and/or
20 a chimeric fusion. Among the homologous polypeptides, preference is given to those in which the amino acid sequence exhibits at least 90% identity, in preference 95%, preferably 97%, and even more preferably 99% identity with the amino acid sequences of the poly-
25 peptides according to the invention. In the case of a substitution, one or more amino acids, which may or may not be consecutive, are replaced with "equivalent amino acids". The expression "equivalent amino acid" is herein aimed at denoting any amino acid which can be
30 substituted for one of the amino acids of the basic structure, without however modifying the essential functional properties or characteristics of the corresponding polypeptides, their biological activities, such as for example the *in vivo* induction of antibodies capable of recognizing the polypeptide, the amino acid
35 sequence of which is included in the amino acid sequence SEQ ID No. 2, SEQ ID No. 4, SEQ ID No. 6, SEQ ID No. 8, SEQ ID No. 10, SEQ ID No. 12 or SEQ ID No. 14, or a fragment thereof. These equivalent amino

- 5 -

acids may be determined either based on their structural homology with the amino acids for which they substitute, or based on the results of cross biological assays which the various polypeptides may produce. By way of example, mention will be made of the possibilities of substitutions which can be made without their being, as a result of this, an extensive modification of the biological activities of the correspondingly modified polypeptides; for example, replacing leucine with valine or isoleucine, aspartic acid with glutamic acid, glutamine with asparagine, arginine with lysine, etc., it naturally being possible to envision the reverse substitutions under the same conditions. Thus, it is possible to envision introducing certain modifications, such as in particular a deletion, addition or substitution of at least one amino acid, into the alpha-helices of the protein without destroying the calyx formed by the structure composed of the beta-sheets; similarly, it is possible to introduce equivalent amino acids which make it possible to conserve the hydrophobic nature of the beta-sheets. It may also be advantageous to introduce modifications into the sequence of the polypeptides of the invention in order to generate homologous polypeptides lacking protease sites.

The term "biologically active fragment" will be intended to denote, in particular, a fragment of amino acid sequence of a polypeptide according to the invention, exhibiting at least one of the functional properties or characteristics of the polypeptides according to the invention, in particular in that: (i) it is capable of being recognized by an antibody specific for a polypeptide according to the invention or by antibodies produced by patients during an immune reaction; (ii) it has at least one of the domains or regions as defined below; (iii) it is capable of binding a hydrophobic ligand and in particular odorous molecules, preferably pheromones; (iv) it is capable of specifically binding a receptor.

The term "polypeptide fragment" is intended to denote a polypeptide comprising a minimum of 15 amino acids, in preference 18 amino acids, preferably 25 amino acids and even more preferably 50 amino acids. The polypeptide fragments according to the invention, obtained by cleavage of said polypeptide with a proteolytic enzyme, or with a chemical reagent, or by placing said polypeptide in a very acid environment, are also part of the invention. When the intention is to use a sequence of 15, 18, 25 or 50 amino acids, this, of course, preferably involves the portions corresponding to functional epitopes of the above polypeptides, which may be integrated into a polypeptide with a longer structure, for example in the form of a fusion protein; this will depend on the intended uses of these polypeptides.

According to a preferred embodiment, the present invention relates to an isolated polypeptide selected from a polypeptide corresponding to the sequence SEQ ID No. 2 and named OBPII_{aα}, to the sequence SEQ ID No. 4 and named OBPII_{aβ}, to the sequence SEQ ID No. 6 and named OBPII_{aγ}, to the sequence SEQ ID No. 10 and named OBPII_{bα}, and to the sequence SEQ ID No. 12 and named OBPII_{bβ}.

The polypeptides according to the invention are characterized in that they preferably comprise the Gly-Thr-Trp-Tyr domain.

The invention also relates to the isolated polynucleotide characterized in that it encodes a polypeptide as described above. The polynucleotide according to the invention is chosen from the polynucleotide of sequence SEQ ID No. 1, SEQ ID No. 3, SEQ ID No. 5, SEQ ID No. 7, SEQ ID No. 9, SEQ ID No. 11 or SEQ ID No. 13.

The invention also relates to an isolated polynucleotide, characterized in that it comprises a polynucleotide chosen from:

- 5 a) a polynucleotide of sequence SEQ ID No. 1, SEQ ID No. 3, SEQ ID No. 5, SEQ ID No. 7, SEQ ID No. 9, SEQ ID No. 11 or SEQ ID No. 13, or the sequence of which is that of the RNA corresponding to the sequence SEQ ID No. 1, SEQ ID No. 3, SEQ ID No. 5, SEQ ID No. 7,
10 SEQ ID No. 9, SEQ ID No. 11 or SEQ ID No. 13;
- b) a polynucleotide, the sequence of which is complementary to the sequence of a polynucleotide defined in a),
15
- c) a polynucleotide, the sequence of which comprises at least 80% identity, in preference 90%, preferably 95%, and even more preferably 97% identity with a polynucleotide defined in a) or b),
20
- d) a polynucleotide which hybridizes, under conditions of high stringency, with a sequence of a polynucleotide defined in a), b) or c),
- 25 e) a fragment of at least 15 consecutive nucleotides, in preference 21 consecutive nucleotides, and preferably 30 consecutive nucleotides, of a polynucleotide defined in a), b), c) or d).

30 In the present description, the terms "polynucleotide", "oligonucleotide", "polynucleotide sequence", "nucleotide sequence" or "nucleic acid" will be intended to denote a DNA fragment, both a double-stranded DNA, a single-stranded DNA and products for transcription of
35 said DNAs, and/or an RNA fragment, said synthetic or isolated natural fragments, possibly comprising unnatural nucleotides, denoting a precise chain of nucleotides, which may or may not be modified, making

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it possible to define a fragment or a region of a nucleic acid.

The expression "polynucleotide of complementary sequence" is intended to denote any DNA, the nucleotides of which are complementary to those of SEQ ID No. 1, SEQ ID No. 3, SEQ ID No. 5, SEQ ID No. 7, SEQ ID No. 9, SEQ ID No. 11 or SEQ ID No. 13, or a portion of SEQ ID No. 1, SEQ ID No. 3, SEQ ID No. 5, SEQ ID No. 7, SEQ ID No. 9, SEQ ID No. 11 or SEQ ID No. 13, and the orientation of which is reversed.

For the purpose of the present invention, the term "percentage identity" is intended to mean a percentage of bases which are identical between the polynucleotides, obtained after the best alignment, this percentage being purely statistical and the differences between the two polynucleotides being distributed randomly and over their entire length.

For the purpose of the present invention, the expression "hybridization under conditions of high stringency" means that the conditions of temperature and ionic strength are chosen such that they allow the hybridization between two complementary DNA fragments to be maintained. By way of illustration, conditions of high stringency for the hybridization step for the purposes of defining the polynucleotide fragments described above are advantageously as follows:

The DNA-DNA or DNA-RNA hybridization is carried out in two steps: (1) prehybridization at 42°C for 3 hours in phosphate buffer (20 mM, pH 7.5) containing 5 × SSC (1 × SSC corresponds to a solution of 0.15 M NaCl + 0.015 M sodium citrate), 50% of formamide, 7% of sodium dodecyl sulfate (SDS), 10 × Denhardt's, 5% of dextran sulfate and 100 µg/ml of salmon sperm DNA; (2) actual hybridization for 20 hours at a temperature which depends on the length of the probe (i.e.: 42°C for a

probe of length > 100 nucleotides), followed by 2 washes for 20 minutes at 20°C in 2 × SSC + 2% SDS, 1 wash for 20 minutes at 20°C in 0.1 × SSC + 0.1% SDS. The final wash is carried out in 0.1 × SSC + 0.1% SDS for 30 minutes at 60°C for a probe of length > 100 nucleotides. The high stringency hybridization conditions described above for a nucleotide of defined length will be adjusted by those skilled in the art for longer or shorter oligonucleotides, according to the teaching of Sambrook et al. (1989).

According to one embodiment of the invention, the polynucleotide according to the invention is characterized in that it is directly or indirectly labeled with a radioactive compound or a nonradioactive compound. The polynucleotide according to the invention is used as a primer for amplifying or polymerizing nucleic acid sequences; the invention also relates to the use of a polynucleotide according to the invention as a probe for detecting nucleic acid sequences. According to the invention, the polynucleotide fragments which may be used as a probe or as a primer in processes for detecting, identifying, titering or amplifying nucleic acid sequence will have a minimum length of 9 bases, preferably 18 bases, and even more preferably 36 bases. Finally, the invention relates to the use of a polynucleotide according to the invention as a sense or antisense oligonucleotide for controlling the expression of the corresponding protein product, in this case a polypeptide according to the invention.

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oligonucleotide; however, the sequences used are generally labeled so as to obtain sequences which can be used for many applications. The primers, probes and oligonucleotides according to the invention are labeled
5 with radioactive elements or with nonradioactive molecules; among the radioactive isotopes used, mention may be made of ^{32}P , ^{33}P , ^{35}S , ^3H or ^{125}I . The nonradioactive entities are selected from ligands, such as biotin, avidin, streptavidin or digoxigenin, haptens,
10 dyes and luminescent agents, such as radioluminescent, chemiluminescent, bioluminescent, fluorescent or phosphorescent agents.

The invention also comprises a method for detecting
15 and/or titering nucleic acid according to the invention, in a biological sample, characterized in that it comprises the following steps: (i) isolating the DNA from the biological sample to be analyzed, or obtaining a cDNA from the RNA of the biological sample; (ii)
20 specifically amplifying the DNA encoding the polypeptide according to the invention using primers; (iii) analyzing the amplification products.

The invention also comprises a kit for detecting and/or
25 titering a nucleic acid according to the invention, in a biological sample, characterized in that it comprises the following elements: (i) a pair of nucleic acid primers according to the invention, (ii) the reagents required to perform a DNA amplification reaction and,
30 optionally, (iii) a component for verifying the sequence of the amplified fragment, more particularly a probe according to the invention.

The invention also comprises a method for detecting
35 and/or titering nucleic acid according to the invention, in a biological sample, characterized in that it comprises the following steps: (i) bringing a probe according to the invention into contact with a biological sample; (ii) detecting and/or titering the

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hybrid formed between said probe and the DNA of the biological sample.

5 The invention also comprises a kit for detecting and/or
titering nucleic acid according to the invention, in a
biological sample, characterized in that it comprises
the following elements: (i) a probe according to the
invention, (ii) the reagents required to carry out a
10 hybridization reaction and, where appropriate, (iii) a
pair of primers according to the invention, and also
the reagents required for a DNA amplification reaction.

The invention in particular relates to the methods
according to the invention described above, for
15 detecting and diagnosing cells of cancerous origin, and
mainly breast, uterine, ovarian, prostate and lung
cancers.

The polynucleotides according to the invention may thus
20 be used as a primer and/or probe in processes using in
particular the PCR (polymerase chain reaction) technique
(Erlich, 1989; Innis et al., 1990, and Rolfs et al.,
1991). This technique requires choosing pairs of oligo-
nucleotide primers which border the fragment which must
25 be amplified. Reference may, for example, be made to
the technique described in American patent U.S. No.
4 683 202. The amplified fragments can be identified,
for example after agarose or polyacrylamide gel
electrophoresis or after a chromatography technique
30 such as gel filtration or ion-exchange chromatography.
The specificity of the amplification can be controlled
by molecular hybridization using, as a probe, the
nucleotide sequences polynucleotides of the invention,
plasmids containing these sequences or products of
35 amplification thereof. The amplified nucleotide
fragments may be used as reagents in hybridization
reactions in order to demonstrate the presence, in a
biological sample, of a target nucleic acid of sequence

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complementary to that of said amplified nucleotide fragments.

5 The invention is also directed toward the nucleotide fragments which can be obtained by amplification using primers according to the invention.

Other techniques for amplifying target nucleic acid may advantageously be used as an alternative to PCR (PCR-like), using pairs of primers of nucleotide sequences according to the invention. The term "PCR-like" will be intended to denote all methods using direct or indirect reproductions of nucleic acid sequences, or in which the labeling systems have been amplified; these techniques are of course known; in general, this involves amplifying the DNA with a polymerase; when the sample of origin is an RNA, a reverse transcription should be performed beforehand. There are currently a large number of processes for this amplification, such as for example the SDA (strand displacement amplification) technique (Walker et al., 1992), the TAS (transcription-based amplification system) technique described by Kwoh et al. in 1989, the 3SR (self-sustained sequence replication) technique described by Guatelli et al. in 1990, the NASBA (nucleic acid sequence based amplification) technique described by Kievitis et al. in 1991, the TMA (transcription mediated amplification) technique, the LCR (ligase chain reaction) technique described by Landegren et al. in 1988 and improved by Barany et al. in 1991, which uses a heat-stable ligase, the RCR (repair chain reaction) technique described by Segev in 1992, the CPR (cycling probe reaction) technique described by Duck et al. in 1990, and the Q-beta-replicase amplification technique described by Miele et al. in 1983 and improved in particular by Chu et al. in 1986 and Lizardi et al. in 1988, and then by Burg et al. (1996) and also by Stone et al. (1996).

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When the target polynucleotide to be detected is an RNA, for example an mRNA, use will advantageously be made, prior to carrying out an amplification reaction using the primers according to the invention or to
 5 carrying out a detection process using the probes of the invention, of an enzyme of the reverse transcriptase type in order to obtain a cDNA from the RNA contained in the biological sample. The cDNA obtained will then be used as a target for the primers or the probes used
 10 in the amplification or detection process according to the invention.

The nucleotide probes according to the invention hybridize specifically with a DNA or RNA molecule of a
 15 polynucleotide according to the invention, more particularly with the sequences SEQ ID No. 1, SEQ ID No. 3, SEQ ID No. 5, SEQ ID No. 7, SEQ ID No. 9, SEQ ID No. 11 and SEQ ID No. 13, under high stringency hybridization conditions as given in the form of a
 20 previous example.

The hybridization technique may be carried out in diverse ways (Matthews *et al.*, 1988). The most general method consists in immobilizing the nucleic acid
 25 extracted from the cells of various tissues or from cells in culture, on a support (such as nitrocellulose, nylon or polystyrene), and in incubating the immobilized target nucleic acid with the probe, under well-defined conditions. After hybridization, the
 30 excess probe is removed and the hybrid molecules formed are detected by the suitable method (measurement of the radioactivity, of the fluorescence or of the enzymatic activity linked to the probe).

35 According to another embodiment of the nucleic acid probes according to the invention, they may be used as a capture probe. In this case, a probe, termed "capture probe", is immobilized on a support and is used to capture, by specific hybridization, the target nucleic

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acid obtained from the biological sample to be tested, and the target nucleic acid is then detected using a second probe, termed "detection probe", labeled with an easily detectable element.

5

In a preferred embodiment, the invention comprises the use of a sense or antisense oligonucleotide for controlling the expression of the corresponding protein product. Among the advantageous nucleic acid fragments, 10 mention should in particular be made of the antisense oligonucleotides the structure of which ensures, by hybridization with the target sequence, inhibition of the expression of the corresponding product. Mention should also be made of the sense oligonucleotides 15 which, by interaction with proteins involved in regulating the expression of the corresponding product, will induce either inhibition or activation of this expression. The oligonucleotides according to the invention have a minimum length of 9 bases, preferably 20 of 18 bases, and even more preferably 36 bases.

The invention relates to a recombinant vector for cloning a polynucleotide according to the invention and/or for expressing a polynucleotide according to the 25 invention, characterized in that it contains a polynucleotide according to the invention as described above. The vector according to the invention is characterized in that it comprises the elements which allow the expression of said sequences in a host cell 30 and, optionally, the secretion of said sequences out of the host cell.

The term "expression vector" is intended to mean both autonomously replicating expression vectors of the 35 plasmid type and systems intended to ensure integration in the cells, but these expression vectors may also be expression vectors of the viral type or even, when the intention is to perform, for example, gene therapy, naked DNA. Among the viral vectors, preference is given

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to those derived from adenoviruses, from adenovirus-associated viruses (AAVs), from retroviruses, from lentiviruses, and preferably HIV derivatives, from poxviruses and from the herpesvirus, for expression in
5 a eukaryotic system. Among the nonviral vectors, preference is given to naked polynucleotides, such as naked DNA or naked RNA according to the technique developed by the company VICAL, yeast artificial chromosomes (YACs) for expression in yeast, mouse
10 artificial chromosomes (MACs) for expression in murine cells and, preferably, human artificial chromosomes (HACs) for expression in human cells.

According to a particular embodiment, the vector
15 according to the invention comprises elements for controlling the expression of the polypeptides; these control elements are preferably chosen from (i) the promoter sequence of the hOBPIIa gene according to the invention, which corresponds to the sequence SEQ ID
20 No. 15 and/or from the promoter sequence of the hOBPIIb gene according to the invention, which corresponds to the sequence SEQ ID No. 16; (ii) a polynucleotide, the sequence of which is complementary to the sequence SEQ ID No. 15 and SEQ ID No. 16; (iii) a polynucleotide,
25 the sequence of which comprises at least 80% identity with a polynucleotide defined in (i) or (ii); (iv) a polynucleotide which hybridizes, under conditions of high stringency, with the polynucleotide sequence defined in (i), (ii) or (iii).

30 The computer tools available to those skilled in the art allow them to easily identify the regulatory promoter boxes required and sufficient to control the gene expression, in particular the TATA, CCAAT and GC
35 boxes, and also the stimulatory (enhancer) or inhibitory (silencer) regulatory sequences which control, in CIS, the expression of the genes according to the invention; among these regulatory sequences, mention should be made of IRE, MRE and CRE.

It is also within the scope of the invention to use the elements defined above and chosen from the sequence SEQ ID No. 15 and SEQ ID No. 16, for controlling the expression of heterologous polypeptides other than those of the invention, and in particular for directing the expression of heterologous polypeptides in the cell types in which the polypeptides according to the invention are normally expressed.

The invention also comprises the host cells, in particular the eukaryotic and prokaryotic cells, characterized in that they are transformed with the vectors according to the invention. The host cells are preferably transformed under conditions which allow the expression of a recombinant polypeptide according to the invention. The host cell may be chosen from bacterial cells, but also from yeast cells and from plant and animal cells; the host cell is preferably a mammalian cell (Edwards and Aruffo, 1993), but also an insect cell in which it is possible to use processes which use baculoviruses for example (Luckow, 1993). These cells may be obtained by introducing into host cells a nucleotide sequence inserted into a vector as defined above, and then culturing said cells under conditions which allow the replication and/or expression of the transfected nucleotide sequence.

According to a particular embodiment, the invention also relates to a transgenic animal or plant which comprises host cells according to the invention.

The invention also relates to a method for preparing a polypeptide, characterized in that it uses a vector according to the invention. More particularly, the invention relates to a method for preparing a recombinant polypeptide, characterized in that transformed cells according to the invention are cultured under conditions which allow the expression of said

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The polypeptides obtained by chemical synthesis and possibly comprising unnatural amino acids, corresponding to said recombinant polypeptides, are also included in the invention. The peptides according to the invention
5 may also be prepared using techniques which are conventional in the domain of peptide synthesis. This synthesis may be performed in homogeneous solution or in solid phase.

10 The recombinant polypeptide purification processes used are known to those skilled in the art. The recombinant polypeptide may be purified from cell lysates and extracts, and/or from the culture medium supernatant, by methods used individually or in combination, such as
15 fractionation, chromatography methods, immunoaffinity techniques using specific monoclonal or polyclonal antibodies, etc. A preferred variant consists in producing a recombinant polypeptide fused to a "carrier" protein (chimeric protein). The advantage of
20 this system is that it allows stabilization and a decrease in the proteolysis of the recombinant product, an increase in solubility during the *in vitro* renaturation and/or simplification of the purification when the fusion partner has affinity for a specific
25 ligand.

The present invention relates to the use of a polypeptide chosen from an hOBPIIa and hOBPIIb polypeptide according to the invention, or a fragment thereof, and
30 from LCN1, retinol-binding protein (RBP) and apolipoprotein D (ApoD), as a protein which binds to a hydrophobic ligand, preferably an odorous molecule. In fact, besides the OBPII polypeptides mentioned above, the OBP function of the lipocalins LCN1, RBP and ApoD
35 has never been suggested. Specifically, while the presence of LCN1 in the nasal mucus has been described (Redl et al., 1992), it was, in fact, attributed another function, namely an antiprotease function, whereas the main function of the LCN1 protein purely

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polypeptide according to the invention combined with an adjuvant of the immune response, and then purifying the specific antibodies contained in the serum of the immunized animals on an affinity column to which the polypeptide having been used as the antigen has been attached beforehand. The polyclonal antibodies according to the invention may also be prepared by purification on an affinity column on which a polypeptide according to the invention has been immobilized beforehand.

10

The invention also relates to a monoclonal antibody specific for a polypeptide according to the invention and capable of inhibiting the interaction between said polypeptide and the cellular receptor to which said polypeptide specifically binds. According to another embodiment, the monoclonal antibody according to the invention is capable of inhibiting the interaction between said polypeptide and its hydrophobic ligands, in preference the odorous molecules, and preferably the pheromones, to which said polypeptide binds.

20

The antibodies of the invention may also be labeled in the same way as described previously for the nucleic acid probes of the invention, and preferably with labeling of the enzymatic, fluorescent or radioactive type. Such labeled antibodies may be used to detect these polypeptides in a biological sample. The biological sample preferably consists of a fluid, for example human biopsies, serum or blood. They thus constitute a means for analyzing the expression of the polypeptide according to the invention, for example by immunofluorescence and/or gold-labeling, and/or with enzymatic immunoconjugates.

30

More generally, the antibodies of the invention may advantageously be used in any situation in which the expression of a polypeptide according to the invention is to be observed, and more particularly in immunocytochemistry, in immunohistochemistry or in Western

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blotting experiments, or in ELISA and RIA techniques. It is thus within the scope of the invention to provide a method for detecting and/or titering a polypeptide according to the invention, in a biological sample, characterized in that it comprises the steps of bringing the biological sample into contact with antibodies according to the invention, and then revealing the antigen-antibody complex formed.

Also falling within the context of the invention is a kit for detecting and/or titering a polypeptide according to the invention, in a biological sample, characterized in that it comprises the following elements: (i) a monoclonal or polyclonal antibody as described above; (ii) where appropriate, the reagents for constituting the medium suitable for the immunological reaction; (iii) the reagents for detecting the antigen-antibody complexes produced by the immunological reaction. This kit is in particular useful for carrying out Western blotting experiments and for immunoprecipitation experiments.

The hOBPIIs according to the present invention may be used in many applications.

The first applications of the OBPs lie in the control of odorants; these applications essentially concern personal hygiene (perfumery, cosmetology, pharmacy) or general hygiene.

They may be used in a process for controlling the volatilization of an odorant; such a process is characterized in that it comprises a step of binding of said odorant with a polypeptide according to the invention or with the LCN1, RBP or apolipoprotein D proteins.

It is also possible to bind the odorant to a solid support using an OBPII according to the invention

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mechanisms for detecting the fatty acid load of the food intake, in particular in the buccal region. The invention therefore also relates to the use of the polypeptides above, in combination with fatty acids, for decreasing the consumption of fatty acids, in particular in hyperlipidemias or in obesity. The polypeptides above may therefore be used for treating hyperlipidemias and obesity. Specifically, these proteins contribute to the detection of the fatty acid content in the food taken in (Gilbertson, 1998); an excess of these proteins should lead to the physiological system which detects the fatty load of a food intake being deceived. Thus, a food portion which is low in fat but supplemented with OBP, ApoD, RBP or LCN1 pre-loaded with fatty acid will be falsely identified as being rich in fat.

Another application is to supplement a nonmaternal milk with one or more of the polypeptides described above.

The OBPs according to the present invention may be used in preventive and curative therapy.

Thus, a lack of detection of this type of protein in a biological specimen using an antibody, a primer and/or a probe according to the invention may be an element for the diagnosis of anosmia.

Similarly, it is possible to envision the detection of anti-OBPII antibodies in a biological sample of human origin; the presence or the titer of these antibodies may be related to certain types of allergy, in particular in asthmatics. In this case, it may be possible to treat this type of allergy by administering polypeptide fragments as described, which would decrease the immune reactions to the external allergies.

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The invention therefore relates to a process for detecting antibodies against human OBPIIs (hOBPIIs) in human serum from an allergic and/or asthmatic patient using an hOBPII polypeptide according to the invention.

5

According to another embodiment, the invention relates to a process for detecting antibodies against human OBPIIs (hOBPIIs) in a biological fluid from a patient suffering from a cancer, and in particular from prostate cancer and/or from breast cancer and/or from 10 uterine cancer and/or from ovarian cancer and/or from lung cancer. Specifically, the proteins according to the invention are overexpressed in tumors, and in particular in prostate cancer (US 5804368, WO 97 10503, CS 84 02898, CS 83 02012, CS 82 08506, CS 82 08215), 15 breast cancer (Stoecz and Gould, 1995; Simard et al., 1992), uterine cancer, ovarian cancer and lung cancer.

The OBPs according to the present invention may also be 20 used in pharmaceutical compositions, this being in particular in order to deliver certain drugs.

In fact, lipocalins are used by mammals to transport hydrophobic molecules in biological fluids. It even 25 appears that they are the natural transporters of xenobiotics *in vivo*. For example, an experimental overload of xenobiotics creates, in male rats, tumors in the proximal convoluted tube (PCT) of the nephron (Borghoff et al., 1990). Specifically, the MUP proteins, 30 only produced in male rats, are reabsorbed by the PCT cells; after lysosomal degradation, the MUP proteins release the xenobiotics which they were transporting. These xenobiotics accumulate in these cells and lead them toward a tumor pathway by mutagenesis. Lipocalins 35 therefore appear to be the ideal structure to trap, in their calyx, a molecule used as a medicinal product, thus avoiding a general distribution thereof. Thus, G. Beste et al. (1999) and application WO 99 16873 describe a strategy of mutagenesis and screening

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According to another embodiment, the polypeptide according to the invention, as an agent for targeting a pharmaceutical compound, is combined with a molecule which allows specific cellular addressing. Among the
5 molecules which allow specific cellular addressing, mention should be made of the group of steroids, of interleukins, of cytokines, of lymphokines, of interferons, of growth factors, of hormones and of antibodies. The molecule is preferably a steroid. The
10 combination of the polypeptide according to the invention and said molecule is produced either via a noncovalent attachment, using for example the avidin-biotin system, or via a covalent attachment, using for example chemical bridging agents.

15 Among the pharmaceutical compounds which the polypeptide according to the invention may transport and target, mention should be made of medicinal products, and in particular anticancer agents. The anticancer
20 agents are selected from the group of antiproliferative, antineoplastic or cytotoxic agents, and are used to arrest the development of cancers and to induce regression and/or elimination of the tumor mass. These anticancer agents are preferably radioisotopes, and
25 even more preferably gamma-ray-emitting radioisotopes, such as iodine¹³¹, yttrium⁹⁰, gold¹⁹⁹, palladium¹⁰⁰, copper⁶⁷, bismuth²¹⁷ and antimony²¹¹. Beta- and alpha-ray-emitting radioisotopes may also be used for the therapy. The non-isotopic anticancer agents bound to
30 the polypeptide according to the invention are many and varied; mention may be made of: (i) antimetabolites, such as antifolate agents, methotrexate, (ii) purine and pyrimidine analogs (mercaptopurine, fluorouracil, 5-azacytidine), (iii) antibiotics, (iv) lectins (ricin, abrin) and (iv) bacterial toxins (diphtheria toxin);
35 the toxins are preferably chosen from *Pseudomonas* exotoxin A, diphtheria toxin, cholera toxin, *Bacillus anthrox* toxin, Pertussis toxin, *Shigella* Shiga toxin,

Shiga-like toxin, *Escherichia coli* toxins, colicin A, d-endotoxin and *Haemophilus A* hemagglutinin.

The invention also relates to a pharmaceutical composition comprising a pharmaceutical compound bound to at least one polypeptide according to the invention, and a pharmaceutically acceptable vehicle. It is also within the scope of the invention to increase the binding capacity by mutagenesis of this same protein. The invention also relates to a pharmaceutical composition as defined above, for treating cancer, and in particular prostate cancer, breast cancer, uterine cancer, ovarian cancer, liver cancer and pulmonary epithelial cell carcinoma. Given that the OBP named hOBPIIb, apolipoprotein D, RBP and α -1-acid glycoprotein are expressed in the prostate, the pharmaceutical composition according to the invention is intended for the treatment of prostate cancer. Similarly, given that it has been demonstrated that the mRNA of the ApoD, RBP and LCN1 proteins is produced in the mammary gland, in addition to the production of hOBPIIb already described in application WO 99 07740, the pharmaceutical composition according to the invention is intended for the treatment of breast cancer.

The invention also relates to a polypeptide according to the invention, as a pharmaceutical compound transporter. The term "transporter" is intended to denote polypeptides according to the invention capable of vehicling a pharmaceutical compound, in the body, without said compound being released at a preferred site in the body. Such a polypeptide constitutes a means for delivering said pharmaceutical compound in the body.

The present invention also relates to a pharmaceutical composition according to the invention, characterized

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in that said polypeptide constitutes a delayed form of delivery of said pharmaceutical compound in the body.

Specifically, the pharmaceutical compound of interest
 5 bound to the OBPII polypeptide according to the invention gradually diffuses in the body as said transporter polypeptide is catabolized in the body. The use of recombinant DNA techniques allows those skilled in the art to modify the half-life duration of the
 10 OBPII polypeptide in the body by introducing modifications into said polypeptide. Thus, it may be advantageous to develop a homologous polypeptide according to the invention, for which the protease cleavage sites have been mutated in order to increase
 15 the half-life of said polypeptide in the body. It may also be advantageous to develop a multimeric polypeptide, expressed for example in the form of a fusion protein, in order to avoid a "glomerular escape" (renal clearance) and thus to increase the half-life of said
 20 polypeptide in the body.

According to another aspect, the invention relates to a compound characterized in that it is chosen from an antibody, a polypeptide, a ligand, a polynucleotide, an
 25 oligonucleotide and a vector according to the invention, as a medicinal product, and in particular as active principles of a medicinal product; these compounds will preferentially be in soluble form, and combined with a pharmaceutically acceptable vehicle.
 30 The term "pharmaceutically acceptable vehicle" is intended to denote any type of vehicle conventionally used in the preparation of injectable compositions, i.e. a diluent and/or a suspending agent, such as an isotonic or buffered saline solution. These compounds
 35 will preferably be administered systemically, in particular intravenously, intramuscularly, intradermally or orally. Their optimal methods of administration, doses and pharmaceutical forms may be determined according to the criteria generally taken into account

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fetal blood and to the general blood circulation of the fetus.

In addition, the polypeptide according to the invention
5 may be used to transport a compound from the fetus to the gestating mother. Specifically, the fetus secretes, via the villi of the nasal structures, polypeptides according to the invention which participate in molecule exchanges between the fetus and the mother at
10 the amniotic membrane. An object of the present invention is therefore also to provide the fetus with polypeptides according to the invention so as to allow the transport of compounds present in the fetus, for example xenobiotics, to the placenta and/or to the
15 amniotic membrane, in order to allow the detoxification of the fetus.

According to another aspect of the invention, the polypeptide according to the invention may be used as a
20 pregnancy marker. Specifically, the polypeptide according to the invention is present in various biological liquids of the mother, such as the urine or blood; measuring the concentration thereof and/or the presence thereof constitutes a pregnancy marker. In
25 this capacity, this marker may be used with, or in place of, or in addition to the pregnancy marker constituted by human choriogonadotrophin, hCG; the polypeptide according to the invention and hCG are, in fact, molecules which are secreted by the trophoblastic
30 placental cells.

Finally, according to another aspect of the invention, the polypeptide may be used as a marker for feto-placental pathological conditions. It may constitute a
35 marker for the rupture of the amniotic sack membrane.

Other characteristics and advantages of the invention appear in the remainder of the description with the

examples and the figures, the legends of which are represented below.

FIGURE 1: Genomic organization of the LCN1/hOBPII locus

5

The top line represents the region of chromosome 9q34. The double-headed arrow indicates the gap between the location of the polymorphic markers D9S1811 and D9S67; the relative position of the markers D9S67 and D9S1826 is uncertain.

10

The middle level indicates the partial organization of the cosmids at 3 different loci: LCN1c, LCN1b-hOBPIIb, LCN1-hOBPIIa. The cosmids (approximately 40 kb) are represented by horizontal lines with their name and the arms of the T3 and T7 vectors noted below the line. The arrows represent the various genes or pseudogenes with their respective orientation. In the cosmids, the vertical dotted lines represent the EcoRI sites. The symbol ⊗ indicates an uncertain orientation of the locus.

15

20

The bottom level shows the intron/exon structures of the LCN1 and hOBPII genes: the black boxes represent the exons; the arrows represent the transcription initiation site; ol1 to ol5 represent the oligonucleotide probes used to screen the cosmids; Alu represents the presence of repeat sequences.

25

FIGURE 2: Dot Plot Analysis of:

30

- (A) sequence of the LCN1-hOBPIIa locus (AC000396+ACXXXXX) versus the LCN1b-hOBPIIb locus (AC002098);
- (B) sequence of the LCN1-hOBPIIa locus (AC000396+ACXXXXX) versus the sequence of the LCN1c locus;
- (C) sequence of the LCN1b-hOBPIIb locus (AC002098) versus the sequence of the LCN1c locus (AC002106).

35

Genomic sequences were filtered for the repetitive sequences using "Repeatmasker" (REF), and then compared

and dotplotted with the program gcg using a window size of 25, and a criterion of 20 with an 80% homology.

FIGURE 3: Nucleotide sequence of hOBPII genes

5 The upper lines represent the hOBPIIa sequence and the lower lines represent the hOBPIIb sequence for which only the different nucleotides are represented; a dash indicates the absence of corresponding sequences. The shaded capitals are the exon sequences and the lower case letters are the intron sequences. The sizes indicated on the left are indicated in bp. The TATA box is in bold characters and the polyadenylation signal is underlined. The boxes indicate the splice acceptor sites for exons 5, 5b and 5c.

FIGURE 4: Diagrammatic representation of the two hOBPII genes and of their corresponding mRNAs

20 The horizontal lines represent the exon/intron organization with sizes indicated in bp. The shaded boxes numbered from 1 to 7 are the coding exon sequences of the main transcripts; the letters b and c refer to supernumerary exons. The various transcripts are represented using assembled boxes: the first, hOBPIIa α and hOBPIIb α , correspond to the main transcripts, the others correspond to the forms derived from alternate splicing. ↓ indicates a shift in the reading frame resulting from the insertion or from the deletion of an exon and * represents a stop codon. The letter "a" represents an α -helix and "b" β -sheets predicted by the DSC program. The letters in italics are predictions obtained with the "Predator" program.

35 **FIGURE 5: Alignment of the protein sequences derived from the two human genes hOBPIIa and hOBPIIb (hOBPIIa α = OBP2aaHOMS, hOBPIIb α = OBP2baHOMSA, hOBPIIb β = OPB2bbHOMSA, hOBPIIa γ = OBP2agHOMSA, hOBPIIa β = OBP2abHOMSA), of human tear lipocalins (LCN1_HOMSA), of**

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rat OBPII (OBP2_RATNO), of bovine lactoglobulin BLG (LACB_BOSTA), of mouse MUP (MUP6_MUSMU), of human RBP (RBP_HOMSA), of bovine OBP (OBP_BOSTA), of rat MUP (MUP_RATNO) and of porcine OBP (OBP_SUSSC).

5

The residues in the dark gray boxes are identical and those in the light gray boxes are similar. The elements of secondary structure predicted with the DSC program are underlined and the amino acid residues are in italics. The β -sheets and the α -helices are numbered for hOBPIIa and b. The anticipated signal peptide cleavage site is indicated with an arrow (AAA↓LS) at position 15. Nonaligned sequences of very divergent forms of spliced genes, hOBPa δ (OPB2adHOMSA) and hOBPby (OBP2bgHOMSA) have been added at the bottom of the alignment after the analysis.

10

15

FIGURE 6: RT-PCR analysis

(A) Detection of the LCN1, LCN1b and LCN1c RT-PCR products with their specific probes, and (B) detection of the hOBPIIa and hOBPIIb RT-PCR products with their specific probes. (C) Control G3PDH RNA. The sizes are indicated in bp.

25

FIGURE 7: Tissue location of the hOBP mRNAs

Sections of the middle meatus (A, B), of the conchae (C, D), of the prostate (E, F), of the vas deferens (G, H), and of the mammary glands (I, J) were hybridized with digoxigenin-11-UTP-labeled hOBP riboprobes. The hybridization with a sense hOBP riboprobe did not reveal any signal (B, D, F, H, J). A specific hybridization signal is obtained with an antisense riboprobe (A, C, E, G, I). The arrows indicate different structures: AC: acinar cells, EC: epithelial cells, GC: glandular cells, SD: secretory duct and L: lumen (X200).

30

35

FIGURE 11: Study of the polyclonal antibody against the hOBPII proteins

The fusion protein is loaded in wells 1 and 3. A sample
5 of human nasal mucus is loaded in wells 2 and 4. The
proteins contained in wells 1 and 2 were revealed with
Coomassie blue staining. The antibody hybridized on the
membrane obtained by Western blotting specifically
10 reveals the GST-hOBPIIb protein between 30 and 42 kDa
(well 3) and the 18 kDa hOBPIIa and hOBPIIb (well 4) in
the human nasal mucus.

FIGURE 12: Immunohistochemistry on the tissues of the olfactory apparatus

15 Strong immunoreactivity can be observed on the section
of septum (green staining, A) on the two sections of
concha (C and D) and on the section of middle meatus
(F). The negative controls prepared for the septum (B),
20 for the concha (E) and for the middle meatus (G) show
no reactivity. The cell nuclei are located by DAPI
staining (blue staining). The magnifications used are
X 100 for the sections A, B, C, E and G, and X 200 for
the sections D and F.

25 **FIGURE 13: Immunohistochemistry on tissues of the oral sphere**

These two tissues, lacrymal glands (A, B) and von
30 Ebner's glands (E, F) show strong immunoreactivity
compared to the results obtained with the pre-immune
serum (C for the lacrymal glands and D for the von
Ebner's glands). Magnification X 200.

35 **FIGURE 14: Immunohistochemistry on the mammary glands and lung**

Strong immunoreactivity is visible on the sections of
mammary glands (A and B) compared to the negative

FIGURE 18: Demonstration of the hOBPII protein in various structures of the oral sphere: lacrymal gland (A: 200x, B: 600x), von Ebner's glands (C: 100x, D: 400x), lung (E: 200x, 800x)

5

Detection of the hOBPII protein with a rabbit polyclonal serum revealed by peroxidase activity detected by DAB. AC: secretory acinar cell, P: DAB precipitate (hOBPII detection).

10

FIGURE 19: Demonstration of the hOBPII proteins in various placental structures: chorionic villi (A: 300x, B: 800x), amniotic membrane (C: 800x), cord (D: 600x)

15

Detection of the hOBPII proteins with a rabbit polyclonal serum revealed by peroxidase activity detected by DAB. L: lacuna (contains maternal blood), C: fetal capillary, Am: amniotic membrane, E: epithelial cells derived from the extra-embryonic ectoblast, MB: basement membrane, M: mesenchymal layer, P: DAB precipitate (hOBPII detection).

20

FIGURE 20: Demonstration of the LCN1 protein in various placental structures: chorionic villi (A: 200x, B: 800x), amniotic membrane (C: 600x), cord (D: 500x)

25

Detection of the LCN1 protein with a rabbit polyclonal serum revealed by peroxidase activity detected by DAB. L: lacuna (contains maternal blood), C: fetal capillary, Am: amniotic membrane, E: epithelial cells derived from the extra-embryonic ectoblast, MB: basement membrane, M: mesenchymal layer, P: DAB precipitate (LCN1 detection).

30

FIGURE 21: Demonstration of the ApoD protein in various placental structures: chorionic villi (A: 100x, B: 500x), amniotic membrane (C: 400x), cord (D: 400x)

5 Detection of the ApoD protein with a mouse monoclonal antibody revealed by peroxidase activity detected by DAB. L: lacuna (contains maternal blood), C: fetal capillary, Am: amniotic membrane, E: epithelial cells derived from the extra-embryonic ectoblast, MB: basement
10 membrane, M: mesenchymal layer, P: DAB precipitate (ApoD detection).

EXAMPLES

15 **EXAMPLE 1: MATERIALS AND METHODS**

1.A. Human chromosome 9 cosmid library

A copy of the library of cosmids specific for human
20 chromosome 9, LL09NC01P, constructed by Dr. J. Allmeman (Biochemical Sciences Division, Lawrence Livermore National Library, Livermore, CA USA) under the aegis of the National Gene Library Project, supported financially by the American Department of Energy, was used. The
25 library was screened and the clones analyzed as previously described (Lacazette et al., 1997).

1.B. Cloning and sequence analysis

30 A Lambda gt11 human testes cDNA library (Clontech) (10^7 p.f.u.) was amplified by 30 polymerase chain reaction (PCR) cycles (94°C 45 sec, 54°C 45 sec, 72°C 1 min 30 sec) with the primer oliEST58 CCTGCAGGTACATGAGCTTCC and 5' or 3' screening amplimers for inserts located on
35 the arms of the lambda gt11 vectors. A nested PCR was then carried out with oliEST26 CGCTGTATTTGCCAGGCTCC and oligonucleotides specific for the arm of the vector. The PCR products were subcloned into the pGEM-T(r)

vector, which made it possible to obtain the 5' end of the hOBPII gene cDNAs.

The sequences obtained using the standard oligonucleotide pGEM-T(r) and a dye terminator-based sequencing ready reaction mix (Applied Biosystems) were separated by electrophoresis using an ABI PRISM 377 automatic sequencer (Perkin Elmer) and were then analyzed with the sequence navigator 1.0.1. software (Perkin Elmer). Full length cDNA clones of hOBPIIa (hOBPIIa α , hOBPIIa β , hOBPIIa δ , hOBPIIa γ) and hOBPIIb (hOBPIIb α , hOBPIIb β , hOBPIIb γ) were obtained from the RT-PCR by purifying the bands of interest according to the manufacturer's instructions (Qiagen gel extraction kit) or, for the weakly expressed alternative forms, by subcloning the nested PCR products into a pGEM-T(r) vector.

1.C. RT-PCR analysis

Tissue samples were collected from 45 to 55 year old Caucasian individuals, in agreement with the French regulations in force. The total RNA is extracted according to a single-step method using the RNA NOW[®] reagent, according to the manufacturer's (Biogentex) instructions. 5 μ g of total RNA were reverse-transcribed in a final volume of 20 μ l using 0.5 ng of oligonucleotide GACTCGAGTCGACATCGATTTTTTTTTTTTTTTTTT with the Superscript[®] preamplification system (Gibco BRL). Three μ l of these reactions were then used for the following PCRs. The expression of the specific mRNAs was determined by PCR using: the primers TL: CCTCTCCCAGCCCCAGCAAG and AP: GACTCGAGTCGACATCG for the LCN1-type genes (LCN1, LCN1b, LCN1c) and, for the hOBPII-type genes, the primers DE: CGCCCAGTGACCTGCCGAGGTC, and FI: CTTTATTTGGAGTCAGGTGGGTG. As a quality control for the RNAs the primers G3PDH1: CTCTGCCCCCTCTGCTGATG and G3PDH2: CCTGCTTCACCACTTCTTG for the G3PDH gene were used; the G3PDH gene is considered to be constitutively

expressed in all cell types. 32 PCR cycles (94°C 45 sec, 54°C 45 sec, 72°C 2 min 30 sec) were performed and the amplification products were separated on 1% agarose gel. The DNA was transferred onto a Hybond N+®
5 membrane.

For the detection of the expression of the various genes, several oligonucleotides specific for the respective genes were synthesized:

10

- olLCN1: GACTCAGACTCCGGAGATGA,
- olLCN1b: AACTCAGACACCAGAGATGA,
- olLCN1c: GACTCAGATCCCGGAGATGA,
- ol5: CCAGGAGGGACCACTACA specific for the hOBPIIb gene,
- 15 - ol4: CCGGGACGGACGACTACG specific for the hOBPIIa gene,
- G3PDH3: CTCATGACCACAGTCCATGC.

The oligonucleotides are labeled with $\gamma^{32}\text{P}$ -ATP using T4 kinase (Applied Biosystem); the labeled oligonucleotide
20 hybridizations are performed at 42°C. The final wash is performed in a solution of 0.1 X SSC, 0.1% SDS at 48°C for 20 min. The specificity of the oligonucleotide hybridization reactions is controlled using samples of digested cosmid DNA (p233G2 for LCN1 and hOBPIIa, P19E7
25 for LCN1b and hOBPIIb, and P181A9 for LCN1c) loaded onto the gel with RT-PCR products.

1.D. Genotyping and linkage analysis

30 The genotyping is carried out with PCR reactions using 100 ng of genomic DNA originating from 8 CEPH reference families, and using the oligonucleotides oli9 TGTTCCGGGAACGCAGCTT and oli10a TGCCGCTGTCCCCACGTCGG. The thermocycler parameters consist of an initial cycle at
35 94°C for 10 min followed by 30 cycles at 94°C for 30 sec, 55°C for 30 sec and 70°C for 45 sec, and then a final elongation step of 10 min at 70°C. The PCR products are then analyzed on a 3% agarose gel. The information regarding the markers for chromosome 9 may

be obtained at the following internet address
(<http://galton.ucl.ac.uk>); the analyses were carried
out using the linkage study tools previously described
in Lacazette *et al.* (1997). The haplotypes are
5 reconstructed manually according to the recombination
events previously described in family 1362 (Attwood *et al.*, 1994).

1.E. Secondary structure predictions

10

A multiple alignment of the lipocalin proteins for
which the crystallographic structures have already been
described (Monaco *et al.*, 1992; Spinelli *et al.*, 1998),
with the hOBPIIa and hOBPIIb proteins, was obtained
15 using the Clustalx software (<ftp://ftp.infobiogen.fr>). The
putative secondary structures were determined with the
DSC (Discrimination of protein Secondary structure
Class) program developed by R.D. King and M.J.E.
Sternberg ([http://bioweb.pasteur.fr/seqanal/interfaces/
20 dsc-simple.html](http://bioweb.pasteur.fr/seqanal/interfaces/dsc-simple.html)). The secondary structures of the
proteins corresponding to the alternatively spliced
forms are presumed to be identical to the conventional
forms before the reading frame shift; after the shift,
the structural prediction is made with a single
25 sequence and is performed using the Predator software
(<http://pbil.ibcp.fr/cgi-bin>).

1.F. Phylogenetic analysis

30 The lipocalin proteins, the sequences of which are
entirely known, were aligned three times consecutively
using the Clustalx software (<ftp://ftp.infobiogen.fr>).
Distances in the phylogenetic tree were calculated with
Clustalx and plotted with Njplot.

35

1.G. In situ hybridization

Serial sections cut on a cryostat (8 μ m thick) are
collected on SuperFrost® Plus slides (Menzel Glazer)

and stored at -80°C . Antisense and sense RNA probes are obtained according to standard techniques using T7 or SP6 polymerase, from a matrix obtained by digestion of the phOBPIIaP2 cDNA clone with NcoI or PstI restriction enzymes, and using DIG-11-UTP (Boehringer Mannheim) (the probe is approximately 150 nucleotides long). The matrices digested with PstI and transcribed with T7 RNA polymerase correspond to the antisense probe and the matrices digested with NcoI and transcribed with the SP6 RNA polymerase correspond to the sense probe. Tissue sections are fixed in 4% paraformaldehyde for 15 min and then rinsed for 5 min in 2 X PBS. After acetylation (2 x 5 min in a triethanolamine (TEA) buffer, pH 8, containing 0.25%, v/v, of acetic anhydride), the tissue sections are prehybridized at 60°C for 15 min in 50% formamide/1 X SSC. The labeled probes are applied to each of the sections in 50 μl of hybridization buffer (50% formamide, 1 X Denhardt's, 500 $\mu\text{g}/\text{ml}$ total tRNA, 10% Dextran sulfate, 10 mM dithiothreitol). The sections are covered and then incubated in humid chambers at 50°C overnight. After hybridization, the slides are immersed at 55°C in a washing buffer (50% formamide, 1 X SSC) for 2 hours. Each slide is then rinsed twice 5 min in 2 X SSC at ambient temperature, and then treated for 30 min with 10 mg/ml of RNase at 37°C and, finally, immersed in a washing solution (50% formamide, 2 X SSC) for 2 hours at 55°C . The slides are then placed in 0.1 X SSC for 15 min at 55°C . The immunodetection is performed using an alkaline phosphatase-conjugated anti-DIG antibody (Fab fragments) according to the Boehringer Mannheim protocol. The sections are examined at different magnifications using an Axiophot microscope (Zeiss).

35 **1.H. Production of a monoclonal antibody against the hOPB_{II} proteins**

Two hundred and fifty μl of fusion proteins were given to the company Agro-Bio for the production of

antibodies. Rabbits (New Zealand White SPF) are injected on days 0, 14, 28 and 42. The sera are taken on days 0, 35, 49 and 63. In order to test the antibody, the fusion protein is loaded in lanes 1 and 3 (figure 11), while human nasal mucus is loaded in lanes 2 and 4. Lanes 1 and 2, and also the lane corresponding to the molecular weight ladder, correspond to revelation with Coomassie blue. Lanes 3 and 4 correspond to detection of the proteins recognized by the antibody using the Western blot technique. Lanes 1 and 3 show the presence of a series of truncated recombinant proteins between 30 and 42 kDa derived from a relatively inefficient synthesis due to the presence of many codons which are rare in the bacterium, within the hOBPIIb sequence. However, this production of recombinant protein was sufficient to produce a polyclonal antibody of good quality, as indicated by the revelation, in lane 4 corresponding to the nasal mucus, of an 18 kDa band specific for the hOBPII proteins.

1.I. Immunohistochemistry

1.I.1. 8 μ m serial sections of various tissues are fixed with paraformaldehyde (5 min) and rinsed three times with 1 X PBS (15 min) then incubated for 30 min in a 3% solution of BSA in 1 X PBS, before being incubated overnight in the presence of the anti-fusion protein antibody in a 1 X PBS solution. After three rinses in 1 X PBS (15 min), a rabbit anti-IgG antibody coupled to FITC (green staining by fluorescence) in a 1 X PBS solution is placed in contact with the slides for 3 hours. After rinsing in 1 X PBS, a solution of DAPI (100 ng/ μ l in 1 X PBS) is applied for 10 min (counterstaining of the cell nuclei in blue). After three rinses in 1 X PBS (15 min) the slides are mounted in water-glycerol (50/50). The analysis is carried out in the presence of DAPI and FITC filters with the aid of a CDD camera using an integration time of between 4 and 32 ms.

1.I.2. The tissues are frozen at -80°C in the presence of OCT. 7 micrometer serial sections are cut using a microtome. The tissues are then fixed with 4% paraformaldehyde in PBS. The slides are incubated for 10 min in a solution of methanol in the presence of 4.5% of H_2O_2 , and then rinsed three times in PBS for 5 min. After incubation for 30 min in a 3% bovine serum albumin solution, the first antibody (diluted to 1/350) is incubated overnight at ambient temperature (rabbit polyclonal antibodies for hOBPII and LCN1, and mouse monoclonal antibody for ApoD). Then, after three 15 min rinses in PBS, the second antibody (diluted to 1/300) (anti-rabbit for hOBPII and LCN1, and anti-mouse for ApoD) is incubated for 3 h at ambient temperature. After three rinses in PBS for 15 min, peroxidase revelation with DAB is carried out for 2 to 10 min, using the "Vector" kit. The slides are then rinsed with water and counterstained with Mayer's hematoxylin for 2 min. After two rinses with water, the slides are dehydrated by passing them successively through baths containing 70° , 95° , 100° and 100° ethanol solutions and, finally, two baths of toluene. The slides are then mounted in Canada balsam and observed using a Zeiss microscope.

**EXAMPLE 2: IDENTIFICATION OF A GENE HOMOLOGOUS TO LCN1
LOCATED ON HUMAN CHROMOSOME 9**

The identification of the LCN1 complementary DNA encoding the lipocalin from human tears (Lassagne and Gachon, 1993) has previously been reported, as has the location of the gene on chromosome 9q34 (Lassagne et al, 1993; Lacazette et al, 1997). The two genes encoding the von Ebner's gland proteins 1 and 2 (Kock et al, 1994) correspond to the proteins homologous to LCN1 in rats; this poses the question of whether other genes encoding an LCN1 protein exist in the human genome. *In situ* hybridization experiments (Lassagne et

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al, 1993) and also analyses with somatic hybrids (Lassagne *et al*, 1995) indicate that, if they exist, these additional human genes must be located in chromosomal region 9q34.

5

A library of cosmids specific for human chromosome 9 (LL09NP01) was screened with the human LCN1 cDNA probe; 26 cosmids were identified; they were digested with EcoRI or PvuII, and then hybridized successively with the LCN1 DNA and various oligonucleotides (figure 1).

10

The cosmids are divided into 3 groups. The first group (clones P32H3, P41B5, P63B6, P92H20, P109C6, P145H6, P195B4, P233G2, P233F2, P265D4 and P276H8) correspond to the cosmids containing a sequence of the previously isolated LCN1 gene (accession number: L14927) made up of 7 exons (Holzfeind and Redl, 1994). The second group (clone P19E4, P19E7, P42H9, P98H5 and P142H8) corresponds to the sequence LCN1b homologous to LCN1 (clone P19E4) (accession number: Y10826) from the promoter up to the 6th exon, which then diverges. A third group of cosmids (clone P110C1, P174E4, P174E5, P181A9, P181B6, P211A7, P238C6 and P291E1) contains a region LCN1c, established from the partial sequencing of the clone P181A9 (accession number Y10827), which is highly homologous to LCN1 only from the promoter up to exon 2. Thus, LCN1 is the only gene which contains the 7th and last exon. In addition, the TATA boxes are degenerate in the promoters of the LCN1b and LCN1c genes.

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EXAMPLE 3: IDENTIFICATION OF GENOMIC DUPLICATIONS CONTAINING THE LIPOCALIN GENES AND MAPPING OF CHROMOSOME REGION 9Q34

During the period in time over which the LCN1 gene family was identified, a vast project of physical mapping led to the identification of cosmid contigs partially covering chromosome region 9q34 (Nahmias *et al*, 1994, Van Slegtenhorst *et al.*, 1995, Hornigold *et*

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sequence contig between ABO and the Surfeit locus, confirming the location of LCN1c. P161A1 and P203H12 have less precise locations using the data from Hornigold and from his collaborators because of the
5 limit of the restriction mapping strategy for the duplicated regions. P203H12 (AC000396) appears to correspond to the LCN1 gene which was previously located close to the D9S1826 marker (Lacazette et al., 1997), with the exception of a 60 base pair insertion.

10

In order to test whether P203H12 may correspond to a 4th duplicated region, the region was tested by PCR on genomic DNA from 150 unrelated individuals using primers specific for AC000396 and L14927. A single band
15 corresponding to the length of the AC000396 sequence was detected (data not shown), thus proving that P203H12 contains the previously described LCN1 gene. The LCN1b locus was not positioned precisely in the region, as demonstrated by the fact that the AC002098
20 ends and also the ends of the cosmids containing LCN1b detect no homologous sequence in the databanks.

20

The sequence analysis made it possible to identify a putative minisatellite at position 3177-3724 of the
25 cosmid corresponding to AC002098 (figure 1); a rare polymorphism was thus demonstrated in a population of 20 unrelated individuals (PIC = 0.05). This new polymorphic marker is informative only in family 1362 of the 8 CEPH reference families tested.

30

The linkage analysis revealed two-point lod scores greater than 3 at $\theta = 0$ for the D9S275 and D9S1818 markers. The reconstruction of the haplotype confirmed the location of the LCN1b gene between the D9S1811 and
35 D9S67 markers in chromosome 9q34 (figure 1).

EXAMPLE 4: IDENTIFICATION OF TWO NEW LIPOCALIN GENES

The sequence analyses revealed new data.

The comparison of AC002098 to the databank revealed similarities between this sequence and the lipocalin genes. In addition to the region 21000 to 27000, which
5 contains the LCN1b gene, the region around position 2150 contains sequences which exhibit similarities with the sequences encoding the rat type II odorant-binding proteins, lipocalins from tears and also the EST AA460385.

10

In parallel, the comparison of the AC000396 sequence to the databank revealed, in addition to the region 11100 to 17100 containing the LCN1 gene, similarities for the same group of sequences in the region 36600 to 37800.

15

The EST AA460385 expressed in human testes corresponds to 4 exons of the new lipocalin gene present in the clone P161A1 (AC002098). A putative 50 base pair exon similar to the EST sequence is also present at the end
20 of the clone P203H12 (AC000396).

The inventors have therefore concluded that a new lipocalin gene exists at a position distal to LCN1b. The inventors have put forward the hypothesis that,
25 subsequent to a genomic duplication, a second gene orthologous to the gene encoding the EST is present in the 3' region of LCN1.

The sequencing of the cosmid P233G2 containing this
30 region (figure 1) with oligonucleotides defined from the EST in fact revealed the presence of another lipocalin gene at a locus 20 kb distal to the LCN1 gene. In order to identify the first exons of the two new genes, nested PCRs on cDNA clones originating from a testes
35 library were carried out between the oligonucleotides located in the 5' region of the EST and the arms of the vector. The PCR products were cloned and their sequence revealed three additional exons going by the genomic sequence (figure 1). A TATA box is present upstream of

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the first exon in both cases (figure 3). The two mRNAs, hOBPIIa corresponding to the gene located downstream of LCN1 and hOBPIIb located downstream of LCN1b (figures 3 and 4), are 97.5% and 63% identical to those corresponding to the LCN1 gene. The intron/exon organizations of these two genes are consistent with the lipocalin family.

In addition, the deduced protein sequences (170 amino acids) confirm that they belong to the lipocalin family. The two putative mature proteins hOBPIIa (MW = 17.8 kDa) and hOBPIIb (MW = 18.0 kDa) are 89% identical. These two novel proteins contain a putative signal peptide of 15 amino acids (figure 5), the conserved residues GXWY at position 27 to 30, and two cysteines which may be involved in a disulfide bridge (position 74 to 166).

However, the paired sequence comparisons with the other lipocalins indicate weak conservation of the amino acid residues (15 to 25%), with the exception of human TL/VEG encoded by the LCN1 gene (mean value 43% over 155 amino acids) and rat OBPII (mean value 45.5%). Moreover, the calculated isoelectric points of hOBPIIa and of hOBPIIb are 7.85 and 8.72, respectively, whereas those of the lipocalins are acid (in the region of 4.5), with the exception of rat OBPII (PI = 9.01). The prediction for the secondary structures of the hOBPIIa and hOBPIIb proteins with the DSC program, by carrying out multiple alignments with the sequences of lipocalins of known structures, indicates the presence of 8 strands of antiparallel β -sheets which may allow the formation of a calyx, followed by an α -helix and a final β -sheet, in agreement with the known data for the structures of the lipocalins (figure 5).

**EXAMPLE 5: STUDY OF THE EXPRESSION OF THE LIPOCALIN
GENES IDENTIFIED**

In order to specify whether LCN1b, LCN1c, hOBPIIa and
5 hOBPIIb are expressed genes, RT-PCR analyses were
carried out on 18 different human tissues known to
produce lipocalins (figure 6).

Two pairs of primers were synthesized; one recognizes
10 the LCN1-type mRNAs and the other the hOBPII-type
mRNAs. The hybridization of the oligonucleotides
specific for the genes, on membranes on which the RT-
PCR products are immobilized after transfer via the
Southern technique, and then the subcloning of the
15 hybridized products revealed that LCN1b and LCN1c are
not expressed in any of the products tested, whereas
the LCN1 mRNAs are detected in the lacrymal gland, the
sweat gland, the von Ebner's glands, the nasal septum,
the epithelium of the nasal concha, as well as the
20 placenta and mammary glands (figure 6A). These data and
the sequence information, which indicates that the TATA
box and the last exon have been lost, make it possible
to affirm that LCN1b and LCN1c are pseudogenes which do
not contribute to the formation of the human LCN1
25 proteins.

On the other hand, the hOBPIIa and hOBPIIb genes are
expressed, which confirms the previous detection
thereof in the cDNA libraries. Surprisingly, although
30 both proteins hOBPIIa and hOBPIIb are very similar over
their entire sequence, including in the 1.5 kb promoter
region, their expression profiles are different
(figure 6B). The hOBPIIa protein is strongly expressed
in the nasal septum, the middle meatus, the nasal
35 concha, the testes and the placenta, and more weakly in
the mammary glands, the lacrymal glands, the sweat
glands, the von Ebner's glands and the lung. On the
other hand, the hOBPIIb protein is expressed
predominantly in the prostate, the testes and the

mammary glands, and more weakly in the submaxillary glands, the nasal septum and the middle meatus.

In addition, the RT-PCR analyses revealed the existence of alternative splicing of the product of transcription of the hOBPIIa gene and of the hOBPIIb gene, which generates, respectively, four and three mRNAs (figures 3, 4, 5 and 6).

The transcription of the hOBPIIa gene generates at least four mRNAs which encode four different proteins; the first mRNA encodes the hOBPIIa α protein which corresponds to the hOBPIIa protein described above. In the hOBPIIa gene, three different splice acceptor sites have been identified for exon 5 (figures 3 and 4), thus forming two other splice variants. A first splice variant has an acceptor site for exon 5 located 49 bp before the previous one (exon 5b); this generates a 725 nucleotide mRNA which encodes the 146 amino acid protein hOBPIIa β . This protein is identical up to the 8th putative β -sheet and then different with only 16 additional amino acids. A second splice variant has an acceptor site for exon 5 located 65 bp before the previous one (exon 5c); this generates a 741 nucleotide mRNA which encodes a 228 amino acid protein hOBPIIa γ . This protein has the first eight putative β -sheets identical to those of hOBPIIa α , and is then different in the C-terminal region (figure 5) due to a reading frame shift generated by this alternative splicing event; the structure of this C-terminal region predicted by the Predator program is a long bent region containing a 9th β -sheet.

In the case of the hOBPIIb gene, in addition to the hOBPIIb α mRNA previously described, a 106 bp supernumerary exon (exon 3b) between the previous exons 3 and 4 was identified (figure 3). This longer mRNA (782 nucleotides) encodes a 165 amino acid protein hOBPIIb β . From a protein structure point of view,

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hOBPIIb β is identical to hOBPIIb α up to the 5th putative β -sheet and then differs due to a reading frame shift. The predictions from the computer programs indicate that the motif ALWEALAIIDTRLK is an α -helix which is just behind the fifth β -sheet. Two additional β -sheets may be present in the long C-terminal portion.

None of the alternative splicing variants for one gene is detected symmetrically for the other gene, although the putative splice acceptor and donor sites may be present (figure 3). In addition to these protein motifs which conserve the conventional structure of a lipocalin, we have demonstrated, for the two genes hOBPIIa and hOBPIIb, the existence of a small amount of mRNA which has undergone alternative splicing and encodes proteins, the sequence of which is not directly related to the lipocalins. mRNAs encoding hOBPIIa δ and hOBPIIb γ , which lack the sequence encoding exon 2 and which have, respectively, exon 5b and exon 5, encode putative secreted proteins of 147 and 85 amino acids, respectively (figures 4 and 5); these proteins diverge from the previous proteins from the 24th amino acid.

In order to identify the cells expressing the hOBPII genes, the inventors hybridized digoxigenin-labeled sense and antisense ribonucleic acid probes on tissue sections (figure 7). The hOBPII mRNAs are detected in the acinar cells of the middle meatus and of the nasal conchae, and also epithelial cells of the conchae; this supports the idea that the hOBPII proteins are involved in olfactory function. In addition to the production in the oral sphere, mRNAs encoding hOBPII were detected in the genital sphere, in particular in the glandular cells of the prostate, and in the secretory epithelial cells of the vas deferens. No signal was detected in the male gonads, which suggests that the expression of the hOBPII genes detected in the RT-PCR experiments corresponded to the presence of additional ducts in the tissue preparation (rete testis and lobuli

epididymidis). In combining these results from the detection of all of the mRNAs encoding hOBPII with those of the RT-PCR approach, it appeared that five hOBPII proteins (hOBPII α , hOBPII β , hOBPII γ , hOBPII δ , hOBPII ϵ) are secreted by the epithelial cells of the ducts of the male gonads, and also acinar cells of the middle meatus and of the nasal conchae; in these cells, the mRNAs encoding hOBPII α are highly predominant. Only the two hOBPII δ proteins (hOBPII δ and hOBPII ϵ) are secreted by the epithelio-glandular cells of the prostate and of the mammary glands.

EXAMPLE 6: PHYLOGENETIC ANALYSIS AND CLASSIFICATION OF THE LIPOCALINS

The dotplot analysis (figure 2) and also the genomic duplications which we revealed indicate a common origin for the LCN1 and hOBPII genes. To clarify the relationships between the members of the lipocalin family and to verify that we identified the human gene orthologous to rat OBPII (Dear *et al.*, 1991), we constructed a phylogenetic distance tree with the vertebrate lipocalins (figure 8).

The inventors demonstrated nine main groups of lipocalins derived from a common precursor:

- the apolipoprotein and retinol binding protein (RBP) family (group 1);
- the prostaglandin D synthase and neutrophil gelatinase-associated lipocalin precursor group (group 2);
- the alpha-1-microglobulin/bikunin (protein HC) subfamily (group 3);
- the orosomucoid subfamily (A1AG, A1AH, A1AI) (group 4);
- the oral sphere subfamily 1 (OBPII-type-LCN1/VEGP, VNSP I and II, LALP, CanF1) (group 5);
- the lactoglobulin subfamily (group 6);

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- the oral sphere subfamily 2 (mouse major urinary proteins (MUP)) (group 8);
- 5 - the oral sphere subfamily 3 (OBP1, mouse OBPII, aphrodisin, probasin, BD20) (group 9).

The present invention relates more particularly to groups 5, 8 and 9.

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Group 5 contains the hOBPII proteins which are closely related from an evolutionary point of view to rat OBPII. Taken together, the present results indicate that the human hOBPII genes are orthologs of the rat OBPII gene. This group also contains the LCN1-VEGP proteins of various species. Taking into consideration the genomic organization of the hOBPII-LCN1 genes, the data from the tree illustrate the duplication event (arrow) which gives rise to the ancestral hOBPII and LCN1 genes from their common lipocalin precursor (figure 9). More recently, the original duplications of the 50 kb region containing hOBPII-LCN1 (arrow) generated, in humans, the two hOBPII genes and the LCN1 gene and its pseudogene LCN1b. The additional duplication which gave rise to the LCN1c pseudogene is partial in the human genome and does not produce a functional protein, and is therefore absent from the present tree. In addition, the two rat VEG proteins are more closely related to one another in the phylogenetic tree than to the human LCN1 protein. The situation is identical for the two human OBPII proteins compared to the rat OBPII protein. These results are in favor of a process of gene conversion for at least some lipocalin genes. This may also be correlated to the fact that these genes are located on the same chromosomal arm.

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Group 9 corresponds to the oral sphere family 3 and contains OBPs which have already been described. The aphrodisin protein has already been described as a

pheromone transporter (Henzel *et al.*, 1988) and appears to be orthologous to the rat and mouse OBP1 protein, with two paralogous OBP1 genes. Finally, the olfactory lipocalins produced by the Bowman's glands of *Rana pipiens* (OLFA RANPI), which are considered to be potential odorant transporters in frog mucus, are not related to any putative OBP group (groups 5, 8 and 9), thus suggesting the existence of other OBP categories.

Some lipocalins have been described as being allergens. The major allergen of dogs, the CanF1 protein, mainly expressed in the von Ebner's glands (Konieczny *et al.*, 1997), appears in the tree to be the dog VEG protein. This result, which shows that LCN1 may be allergenic, prompts the inventors to propose that the OBPII proteins are involved in allergic processes. Similarly, the major allergen of horses, EquC1, expressed in the liver and the salivary glands (Grégoire *et al.*, 1996), appears to be orthologous to an MUP protein member (group 3). The major allergen of cows (BD20 BOSTA), which is present in the oral sphere family 2 (group 4) is more closely related to probasin. CanF2, which is mainly expressed in the parotid gland (Konieczny *et al.*, 1997), does not appear to be orthologous to a previously described lipocalin.

The present invention has made it possible to reveal the existence of a genomic duplication at the q34 locus of human chromosome 9, which harbors a gene family of the LCN1 type; this family comprises, besides the LCN1 gene, which has been previously described, two pseudogenes and also two hOBPII genes which are paralogous to LCN1. The inventors have revealed that the hOBPII-LCN1 family results from consecutive genomic duplication events. The sequences and also the genomic organization revealed that the LCN1 and hOBPII genes derive from a common ancestor and were generated by means of tandem duplication. The sequence comparisons demonstrated that the hOBPIIa α and hOBPIIb α proteins are the hOBPII

protein forms closest to LCN1; this is corroborated by the fact that the size of exon 5 of LCN1 is 121 bp, which corresponds to the size of exon 5a of the hOBPII protein, and by the fact that no exon 3b was found in the LCN1 protein. Similar results are obtained when this comparison is carried out with the other seven exons of the lipocalins. These data are in favor of the diversity of the hOBPII proteins being acquired through the integration of surrounding additional genomic DNA at the upstream splice acceptor site for exon 5 of hOBPIIa or through the recruitment of a supernumerary exon (exon 3b) for the hOBPIIb protein. The hypothesis of a recruitment of exons rather than a decrease in the size of the exons or than a loss of exons is more probable.

The inventors showed that the hOBPII and LCN1 genes encode proteins involved in various functions, as shown by the expression of these genes in both the oral sphere and the genital sphere. In addition, the inventors showed, by the phylogenetic analysis (example 6), that several different proteins could contribute to the same odorant-binding protein function; thus, three lipocalin subfamilies (groups 5, 8 and 9) corresponding to several proteins are found expressed in the oral sphere, and in particular in the nasal and buccal glands.

This analysis, showing that several lipocalins contribute to the same physiological function and that the same protein could contribute to various functions, led the inventors to analyze the expression of the other human lipocalins in all of the tissues studied (figure 10). This led to the demonstration that the gene encoding apolipoprotein D is expressed in the glands of the nasal concha and of the middle meatus, thus making it a potential odorant-binding protein. Similarly, retinol binding protein (RBP) is expressed

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by the nasal epithelium and may also contribute to this function.

5 The inventors therefore propose the inclusion of the ApoD and RBP proteins in the human OBP family and include them in all the claims relating to these human OBPs.

10 The inventors have demonstrated that the transcription of the two hOBPII genes generates many alternative transcripts which encode different proteins, the structure of which is compatible with that of a hydrophobic ligand transporter.

15 The inventors have demonstrated the expression of the two hOBPII genes in the oral sphere (nasal glands, von Ebner's glands, submaxillary glands, lacrymal glands, lung). The inventors have also demonstrated that the hOBPII proteins according to the invention are produced
20 by the cells of the genital sphere; the hOBPII gene is mainly expressed in the prostate, the vas deferens and the mammary glands, while the expression of the hOBPIIa gene is restricted to the vas deferens. The in situ analyses have revealed that the mRNA is produced by the
25 glandular cells of the prostate and epithelial secretory cells of the vas deferens, supporting the idea that the corresponding proteins are secreted into the seminal fluid. The proteins according to the invention may therefore be involved in the reproductive
30 function, but also in all the other functions usually attributed to lipocalins.

EXAMPLE 7: PRODUCTION OF A RECOMBINANT HOBPIIb α PROTEIN IN A PROKARYOTIC SYSTEM

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A PCR on the plasmid DNA of an hOBPIIb α clone using the primers BIIa/b (5' GTC GGA TCC CTG TCC TTC ACC CTG GAG G 3'), a sense oligonucleotide beginning 45 bases after the protein-initiating ATG, and XIIf (5' GTC CTC GAG

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GTG TTC GGG AAC GCA GCT TC 3'), an antisense oligo-nucleotide preceding the stop codon of the hOBPIIb α protein, made it possible to amplify all of the DNA encoding the secreted hOBPIIb α protein. The BamH I and

5 Xho I enzymatic restriction sites located at the ends of the two oligonucleotides (bases underlined) were used for directional cloning into a plasmid expression vector pGEX-6P1, followed by transformation by electro-

10 poration (1800 V, 200 Ω , 25 μ F) into a bacterial strain BL21. Synthesis of the recombinant protein is obtained by adding IPTG, at 5 mM final for 3 h, to 250 ml of culture of the strain in LB medium containing 100 μ g/ml of ampicillin pre-incubated at 37°C for 2 h. The

15 centrifuged cultures are taken up in 25 ml of TENG buffer. The lysate is sonicated and recentrifuged. The fusion protein is then purified using 4 ml of beads bearing covalently attached insoluble glutathione (Sigma) per 25 ml of supernatant. After incubation for 4 h, they are washed with 3 volumes of 1M NaCl and then

20 with 10 volumes of 1 X PBS. The elution is obtained by bringing the beads into contact with a glutathione solution (10 mM reduced glutathione, 50 mM Tris-HCl, pH 8.0) for 10 min.

25 The amount of recombinant protein produced is estimated by migration on polyacrylamide gel and staining with Coomassie blue (figure 11). The specificity of induction of a recombinant protein is tested by Western blot using a goat anti-GST antibody (figure 11), revealed

30 with a peroxidase-coupled goat anti-IgG antibody, using an ECL+plus kit (Amersham). The separation of the fusion protein into two proteins is obtained by proteolysis of 100 μ g of pre-dialyzed recombinant protein with 2U of preScissionTM protease (Pharmacia

35 Biotech) in 10 μ l final containing 1 X enzyme buffer, for 4 h at 5°C.

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**EXAMPLE 8: DETECTION OF THE hOBPIIs IN VARIOUS TISSUES
BY IMMUNOHISTOCHEMISTRY**

The location of the hOBPII proteins in the nasal
5 structures (figures 12 and 15), the buccal structures
and the lacrymal glands (figures 13 and 18), the
mammary glands and the lungs (figures 14 and 18) is
revealed by immunohistochemistry.

10 **EXAMPLE 9: DETECTION OF THE hOBPIIs, LCN1 and ApoD IN
THE NASAL STRUCTURES BY RT-PCR AND BY
IMMUNOHISTOCHEMISTRY**

The RT-PCRs carried out using the sets of primers for
15 the hOBPIIs and for LCN1 reveal the presence of the
corresponding mRNAs in at least one of the nasal
structures. The techniques of immunohistochemical
detection both by fluorescence detection and by
revelation using peroxidase and DAB reveal the presence
20 of the hOBPII (figure 15) and LCN1 (figure 16) proteins
in the epithelial cells of the acini of the nasal
glands, and also in the lumina of the secretory ducts.
Some can also be detected on mucus attached to the
tissues.

25 The RT-PCRs carried out using the sets of primers for
apolipoprotein D reveal the presence of the corres-
ponding mRNAs in at least one of the nasal structures.
The techniques of immunohistochemical detection by
30 revelation using peroxidase and DAB reveal the presence
of the ApoD protein (figure 17) in the epithelial cells
of the acini of the nasal glands, and also in the
lumina of the secretory ducts, although the expression
appears to be weaker and in fewer acini than for the
35 hOBPIIs.

**EXAMPLE 10: DETECTION OF THE hOBPIIs, LCN1 and ApoD IN
THE PLACENTA BY RT-PCR AND BY IMMUNOHISTO-
CHEMISTRY**

5 The RT-PCRs carried out using the sets of primers for
the hOBPIIs and for LCN1 reveal the presence of the
corresponding mRNAs in the placenta. The techniques of
immunohistochemical detection by revelation using
10 peroxidase and DAB reveal the presence of the hOBPII
(figure 19) and LCN1 (figure 20) proteins very strongly
in the lacunae of the placental villi and much more
weakly, and very diffusely, in the epithelial cells of
the amniotic membrane and in subjacent cells, both for
the amniotic membrane and for the cord. The expression
15 of the mRNAs detected by RT-PCR in the placental villi
indicates that these hOBPII proteins are produced by
the placenta in order to transfer lipophilic molecules
(hormones, fatty acids, vitamins, etc.) to the fetus
via the fetal blood circulation. The proteins detected
20 in the amniotic membrane may correspond to the
absorption of these proteins from the amniotic fluid,
since it is known that the nasal structures of the
fetus are bathed in this amniotic fluid. The hOBPII and
LCN1 proteins would thus contribute to the transport of
25 lipophilic substances which are small in size from the
mother to the fetus in the chorionic villi via the
fetal blood, and from the fetus to the mother via the
amniotic fluid.

30 The RT-PCRs carried out using the sets of primers for
the ApoD protein reveal the presence of the
corresponding mRNAs in the placenta. The techniques of
immunohistochemical detection by revelation using
peroxidase and DAB reveal the presence of the ApoD
35 protein very strongly in the lacunae of the placental
villi (figure 21). The ApoD protein is not detected in
the cells of the amniotic membrane nor in subjacent
cells, both for the amniotic membrane and for the
cordon. The expression of the mRNAs detected by RT-PCR

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in the placenta indicates that the ApoD protein is produced by the placenta in order to transfer lipophilic molecules (hormones, fatty acids, vitamins, etc.) to the fetus via the fetal blood circulation. The ApoD protein may also be produced by the fetal liver. The ApoD protein would thus contribute to the transport of lipophilic substances which are small in size from the mother to the fetus in the chorionic villi via the fetal blood.

10

More generally, the expression of lipocalins by the placenta, such as alpha-1-acid glycoprotein (A1AG), prostaglandin D synthase (PGDS), PP14 and RBP, as revealed by the RT-PCR techniques, indicates that these proteins, which have hydrophobic molecule-transporting properties, are recruited in massive amounts for mother-child connections, for transporting these molecules which may be steroid hormones, fatty acids, vitamins or xenobiotics, and more generally metabolism-derived detoxification products.

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CLAIMS

1. An isolated polypeptide comprising an amino acid sequence having at least 90% identity with the amino acid sequences SEQ ID No. 2, SEQ ID No. 4, SEQ ID No. 6, SEQ ID No. 8, SEQ ID No. 12 or SEQ ID No. 14.
2. An isolated polypeptide, characterized in that it comprises a polypeptide chosen from:
 - a) a polypeptide of sequence SEQ ID No. 2, SEQ ID No. 4, SEQ ID No. 6, SEQ ID No. 8, SEQ ID No. 12 or SEQ ID No. 14;
 - b) a polypeptide which is a variant of a polypeptide of amino acid sequences defined in a);
 - c) a polypeptide homologous to the polypeptide defined in a) or b) and comprising at least 90% identity with said polypeptide of a);
 - d) a fragment of at least 15 consecutive amino acids of a polypeptide defined in a), b) or c);
 - e) a biologically active fragment of a polypeptide defined in a), b) or c).
3. An isolated polypeptide selected from a polypeptide corresponding to the sequence SEQ ID No. 2 and named OBPII_{αα}, to the sequence SEQ ID No. 4 and named OBPII_{αβ}, to the sequence SEQ ID No. 6 and named OBPII_{αγ}, and to the sequence SEQ ID No. 12 and named OBPII_{ββ}.
4. The polypeptide as claimed in any one of claims 1 to 3, characterized in that it comprises at least the Gly-Thr-Trp-Tyr domain.

5. An isolated polynucleotide, characterized in that it encodes a polypeptide as claimed in any one of claims 1 to 4.

5 6. The polynucleotide as claimed in claim 5, of sequence SEQ ID No. 1, SEQ ID No. 3, SEQ ID No. 5, SEQ ID No. 7, SEQ ID No. 11 or SEQ ID No. 13.

7. An isolated polynucleotide, characterized in that
10 it comprises a polynucleotide chosen from:

a) a polynucleotide of sequence SEQ ID No. 1, SEQ ID No. 3, SEQ ID No. 5, SEQ ID No. 7, SEQ ID No. 11 or SEQ ID No. 13, or the sequence of which is that of
15 the RNA corresponding to the sequence SEQ ID No. 1, SEQ ID No. 3, SEQ ID No. 5, SEQ ID No. 7, SEQ ID No. 11 or SEQ ID No. 13;

b) a polynucleotide, the sequence of which is complementary to the sequence of a polynucleotide defined in a),
20 in a),

c) a polynucleotide, the sequence of which comprises at least 80% identity with a polynucleotide defined in a) or b),
25 a) or b),

d) a fragment of at least 15 consecutive nucleotides, in preference 21 consecutive nucleotides, and preferably 30 consecutive nucleotides, of a polynucleotide defined in a), b), c) or d).
30

8. The polynucleotide as claimed in claim 7, characterized in that it is labeled directly or indirectly with a radioactive compound or a nonradioactive
35 compound.

9. The use of a polynucleotide as claimed in claim 8, as a probe for detecting nucleic acid sequences.

10. The use of a polynucleotide as claimed in any one of claims 5 to 8, as a primer for amplifying or polymerizing nucleic acid sequences.

5

11. The use of a polynucleotide as claimed in claim 8, as a sense or antisense oligonucleotide for controlling the expression of the corresponding protein product.

10

12. A recombinant vector for cloning a polynucleotide as claimed in one of claims 5 to 8 and/or for expressing a polypeptide as claimed in one of claims 1 to 4, characterized in that it contains a polynucleotide as claimed in any one of claims 5 to 8.

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13. The vector as claimed in claim 12, characterized in that it comprises the elements which allow the expression, optionally the secretion, of said polypeptide in a host cell.

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14. The vector as claimed in any one of claims 12 to 13, characterized in that the elements which allow the expression of said polypeptide are chosen from:

25

a) the isolated polynucleotide of sequence SEQ ID No. 15 and SEQ ID No. 16;

30

b) a polynucleotide, the sequence of which is complementary to the sequence of the polynucleotide defined in a);

35

c) a polynucleotide, the sequence of which comprises at least 80% identity with a polynucleotide defined in a) or in b);

d) a polynucleotide which hybridizes, under conditions of high stringency, with a sequence of the polynucleotide defined in a), b) or c).

15. The vector as claimed in claims 13 and 14, for expression in eukaryotic cells, selected from viral DNA and naked DNA.
- 5 16. A host cell, characterized in that it is transformed with a vector as claimed in one of claims 12 to 15.
- 10 17. A process for preparing a recombinant polypeptide, characterized in that a host cell as claimed in claim 16 is cultured under conditions which allow the expression and, optionally, the secretion of said recombinant polypeptide, and in that said recombinant
15 polypeptide is recovered.
18. A recombinant polypeptide which can be obtained using a process as claimed in claim 17.
- 20 19. The use of a polypeptide chosen from a polypeptide as claimed in any one of claims 1 to 4 and 18, or a fragment thereof, as a protein which binds to a hydrophobic ligand, preferably an odorous molecule.
- 25 20. The use of a polypeptide as claimed in any one of claims 1 to 4 and 18, or of the polypeptide hOBPII_{ba} of sequence SEQ ID No. 10, a polypeptide which is a variant of said polypeptide hOBPII_{ba}, a polypeptide homologous to said polypeptide hOBPII_{ba} comprising at
30 least 90% identity with said polypeptide hOBPII_{ba} or a fragment thereof, as a competitive inhibitor, as an agonist or as an antagonist of the cellular receptors for lipocalins.
- 35 21. A monoclonal or polyclonal antibody, and fragments thereof, characterized in that it is specifically against an isolated polypeptide as claimed in one of claims 1 to 4 and 18.

22. The use of an antibody as claimed in claim 21, for demonstrating the presence of a polypeptide as claimed in one of claims 1 to 4 and 18 in a biological sample.

23. A process for detecting an antibody against hOBPII in human serum from an allergic and/or asthmatic patient, using an hOBPII polypeptide.

24. A process for controlling the volatilization of an odorant, characterized in that it comprises a step of binding of said odorant with a polypeptide as claimed in any one of claims 1 to 4 and 18, or with the polypeptide hOBPII_{ba} of sequence SEQ ID No. 10, a polypeptide which is a variant of said polypeptide hOBPII_{ba} or a polypeptide homologous to said polypeptide hOBPII_{ba} comprising at least 90% identity with said polypeptide hOBPII_{ba}.

25. The process as claimed in claim 24, characterized in that the polypeptide is bound to a solid support.

26. The process as claimed in claim 24, characterized in that the polypeptide is in a liquid composition.

27. The process as claimed in claim 26, characterized in that said composition is a fragranced composition for the skin.

28. A process for screening a molecule, preferably odorants or flavors, which comprises passing the molecule over a substrate which comprises a polypeptide as claimed in any one of claims 1 to 4 and 18, or the polypeptide hOBPII_{ba} of sequence SEQ ID No. 10, a polypeptide which is a variant of said polypeptide hOBPII_{ba} or a polypeptide homologous to said polypeptide hOBPII_{ba} comprising at least 90% identity with said polypeptide hOBPII_{ba}, bound to said substrate, said polypeptide

binding said odorant or flavor, and recovering said odorant or flavor from the polypeptide if necessary.

29. The process as claimed in claim 28, characterized
5 in that the molecules are human pheromones.

30. A process for solubilizing lipophilic molecules,
characterized in that it comprises binding said
molecule to a polypeptide as claimed in any one of
10 claims 1 to 4 and 18, or to the polypeptide hOBPII_{ba} of
sequence SEQ ID No. 10, a polypeptide which is a
variant of said polypeptide hOBPII_{ba} or a polypeptide
homologous to said polypeptide hOBPII_{ba} comprising at
least 90% identity with said polypeptide hOBPII_{ba}.

15 31. The application of the polypeptides as claimed in
one of claims 1 to 4 and 18, or of the polypeptide
hOBPII_{ba} of sequence SEQ ID No. 10, of a polypeptide
which is a variant of said polypeptide hOBPII_{ba} or of a
20 polypeptide homologous to said polypeptide hOBPII_{ba}
comprising at least 90% identity with said polypeptide
hOBPII_{ba}, in combination with dietary fatty acids, as a
food supplement.

25 32. The application as claimed in claim 31, for
preparing a medicinal product intended for the
treatment of hyperlipidemias and obesity.

30 33. The application as claimed in claim 31, for
supplementing nonmaternal milks.

34. The polypeptide as claimed in any one of claims 1
to 4 and 18, or the polypeptide hOBPII_{ba} of sequence SEQ
ID No. 10, a polypeptide which is variant of said poly-
35 peptide hOBPII_{ba} or a polypeptide homologous to said
polypeptide hOBPII_{ba} comprising at least 90% identity
with said polypeptide hOBPII_{ba}, as an agent for
targeting a pharmaceutical compound.

35. The polypeptide as claimed in claim 34, characterized in that said polypeptide is expressed in the form of a protein from fusion with a protein which
5 allows specific cellular addressing.

36. The polypeptide as claimed in claim 35, characterized in that said protein which allows specific cellular addressing is chosen from the group composed
10 of interleukins, of cytokines, of lymphokines, of interferons, of growth factors, of hormones and of antibodies.

37. The polypeptide as claimed in claim 34, characterized in that said polypeptide is combined with a
15 molecule which allows specific cellular addressing.

38. The polypeptide as claimed in claim 37, characterized in that said molecule which allows specific
20 cellular addressing is chosen from the group composed of steroids, of interleukins, of cytokines, of lymphokines, of interferons, of growth factors, of hormones and of antibodies.

39. The polypeptide as claimed in any one of claims 1 to 4 and 18, or the polypeptide hOBPII_{ba} of sequence SEQ ID No. 10, a polypeptide which is a variant of said polypeptide hOBPII_{ba} or a polypeptide homologous to said polypeptide hOBPII_{ba} comprising at least 90% identity
30 with said polypeptide hOBPII_{ba}, as a pharmaceutical compound transporter.

40. A pharmaceutical composition comprising a pharmaceutical compound bound at least to a polypeptide as
35 claimed in one of claims 34 to 39, and a pharmaceutically acceptable vehicle.

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41. The pharmaceutical composition as claimed in claim 40, characterized in that the pharmaceutical compound is chosen from the group of anticancer agents.

5 42. The pharmaceutical composition as claimed in claim 41, characterized in that said anticancer agent is a radioactive isotope chosen from the group: iodine¹³¹, yttrium⁹⁰, gold¹⁹⁹, palladium¹⁰⁰, copper⁶⁷, bismuth²¹⁷ and antimony²¹¹.

10

43. The pharmaceutical composition as claimed in any one of claims 40 to 42, characterized in that said polypeptide as claimed in any one of claims 34 to 39 constitutes a delayed form of delivery of said pharmaceutical compound in the body.

15

44. A pharmaceutical composition comprising an expression vector as claimed in claim 12 or 13, and a pharmaceutically acceptable vehicle.

20

45. The pharmaceutical composition as claimed in any one of claims 40 to 44, for treating cancer preferably chosen from breast cancer, uterine cancer, prostate cancer, liver cancer and pulmonary epithelial cell carcinoma.

25

46. The use of a polypeptide chosen from the polypeptide hOBPII_{ba} of sequence SEQ ID No. 10, a polypeptide which is a variant of said polypeptide hOBPII_{ba} and a polypeptide homologous to said polypeptide hOBPII_{ba} comprising at least 90% identity with said polypeptide hOBPII_{ba}, for preparing a medicinal product intended for the treatment of uterine cancer, prostate cancer, liver cancer and pulmonary epithelial cell cancer.

30

35

47. The use of the polypeptide as claimed in any one of claims 1 to 4 and 18, or the polypeptide hOBPII_{ba} of

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sequence SEQ ID No. 10, or a polypeptide which is a variant of said polypeptide hOBPII_{ba} or a polypeptide homologous to said polypeptide hOBPII_{ba} comprising at least 90% identity with said polypeptide hOBPII_{ba}, for
5 preparing a medicinal product intended for the transport of a compound across the placental barrier.

48. The use as claimed in claim 47, characterized in that said transport is carried out from the gestating
10 mother to the fetus, and in that said compound is chosen from hormones, essential fatty acids, lipophilic medicinal products and vitamins.

49. The use as claimed in claim 47, characterized in that said transport is carried out from the fetus to
15 the mother and is intended for the detoxification of the fetus.

50. The polypeptide as claimed in claims 1 to 4 and
20 18, or the polypeptide hOBPII_{ba} of sequence SEQ ID No. 10, a polypeptide which is a variant of said polypeptide hOBPII_{ba} or a polypeptide homologous to said polypeptide hOBPII_{ba} comprising at least 90% identity with said polypeptide hOBPII_{ba}, as a pregnancy marker.

25
51. The polypeptide as claimed in claims 1 to 4 and 18, or the polypeptide hOBPII_{ba} of sequence SEQ ID No. 10, a polypeptide which is a variant of said polypeptide hOBPII_{ba} or a polypeptide homologous to said
30 polypeptide hOBPII_{ba} comprising at least 90% identity with said polypeptide hOBPII_{ba}, as a marker for a fetoplacental pathological condition.

(12) DEMANDE INTERNATIONALE PUBLIÉE EN VERTU DU TRAITÉ DE COOPÉRATION
EN MATIÈRE DE BREVETS (PCT)(19) Organisation Mondiale de la Propriété
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(10) Numéro de publication internationale
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// A61P 3/04, 3/06, 35/00GACHON, Françoise [FR/FR]; 11, rue des Paillards,
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Regimbeau, 26, avenue Kléber, F-75116 Paris (FR).(81) États désignés (national): AF, AG, AL, AM, AT, AU, AZ,
BA, BB, BG, BR, BY, BZ, CA, CH, CN, CR, CU, CZ, DE,
DK, DM, DZ, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU,
ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS,
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NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR,
TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZW.(84) États désignés (régional): brevet ARIPO (GI, GM, KE,
LS, MW, MZ, SD, SI, SZ, TZ, UG, ZW), brevet eurasien
(AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), brevet européen
(AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU,
MC, NL, PT, SE), brevet OAPI (BF, BJ, CF, CG, CI, CM,
GA, GN, GW, ML, MR, NE, SN, TD, TG).

Publiée:

— Sans rapport de recherche internationale, sera republiée
dès réception de ce rapportEn ce qui concerne les codes à deux lettres et autres abrégia-
tions, se référer aux "Notes explicatives relatives aux codes et
abréviations" figurant au début de chaque numéro ordinaire de
la Gazette du PCT(54) Title: ODORANT BINDING HUMAN PROTEINS FIXING HYDROPHOBIC LIGANDS: POLYPEPTIDES AND
POLYNUCLEOTIDES CODING FOR SAID POLYPEPTIDES AND USES THEREOF(54) Titre: "ODORANT BINDING" PROTEINES HUMAINES FIXANT DES LIGANDS HYDROPHOBES: POLYPEPTIDES
ET POLYNUCLEOTIDES CODANT LESDITS POLYPEPTIDES ET LEURS APPLICATIONS(57) Abstract: The invention concerns novel polypeptides fixing hydrophobic ligands and in particular odours, called OBPII (Olor-
ant Binding Protein), the polynucleotides coding for said polypeptides and specific antibodies directed against said polypeptides.
The invention also concerns the uses of said molecules for personal hygiene, uses in agri-food system and nutritional and therapeutic
uses(57) Abrégé: L'invention concerne de nouveaux polypeptides fixant des ligands hydrophobes et notamment des odeurs, dénommés
OBPII (Odorant Binding Protein), les polynucléotides codant pour lesdits polypeptides et des anticorps spécifiques dirigés contre
lesdits polypeptides. L'invention concerne également les applications de ces molécules notamment pour l'hygiène corporelle, les
applications agro-alimentaires, nutritionnelles et thérapeutiques.

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1 / 21

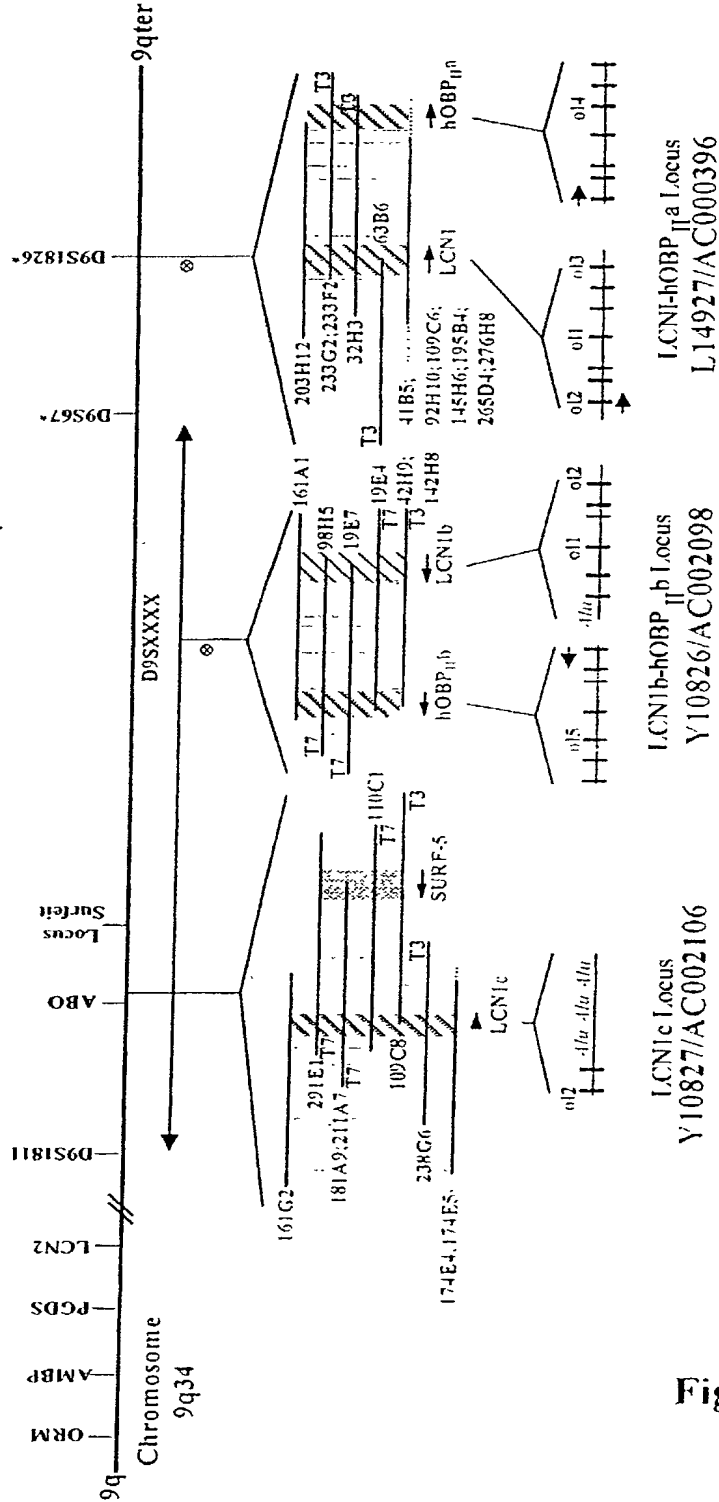


Fig. 1

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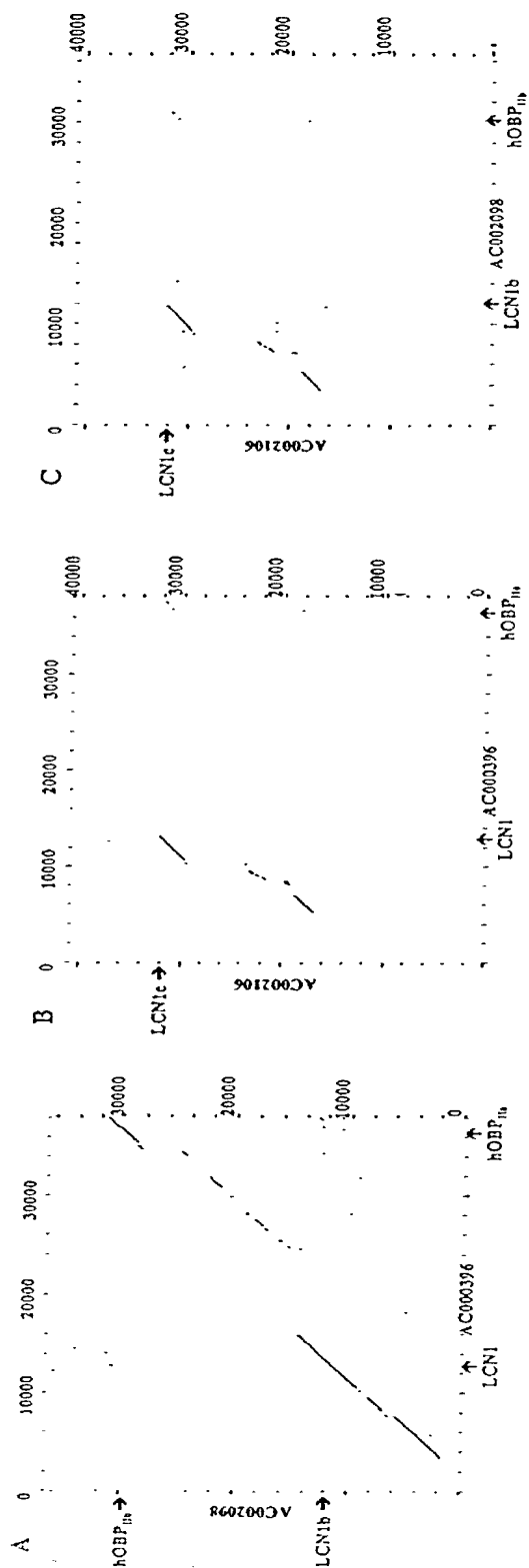


FIG. 2

[illegible]

FIG. 3

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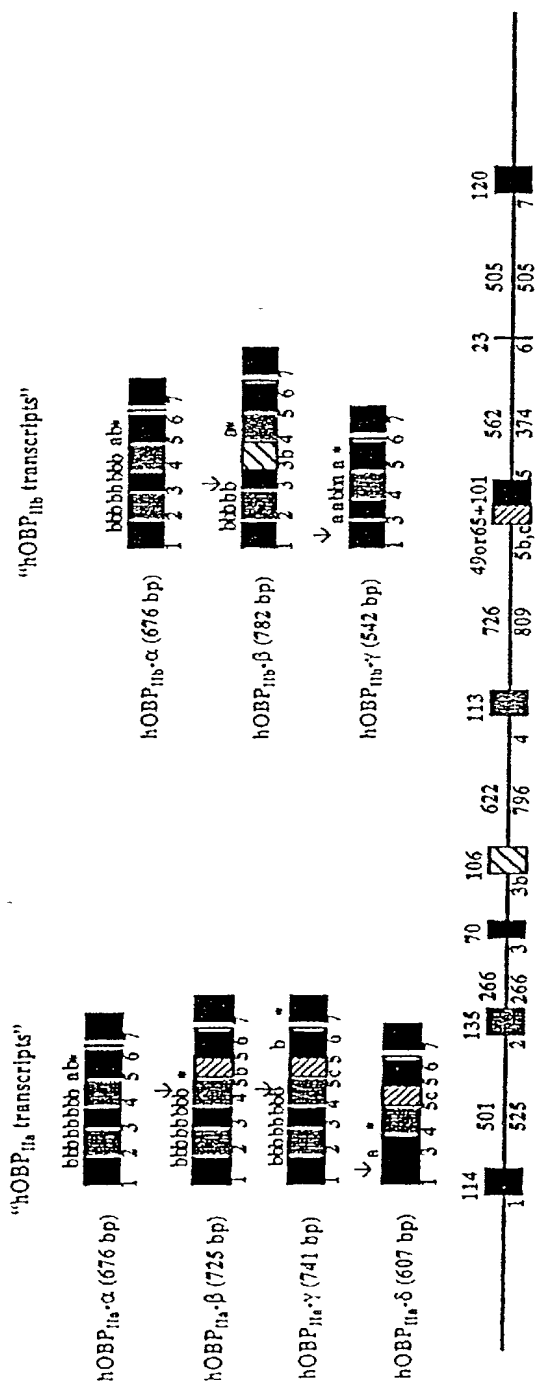


FIG.4

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FIG. 5

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 OBP2baHOMSA -----KTLFETLGAALFTLE-----β1
 OBP2abHOMSA -----KTLFETLGAALFTLE-----β1
 OBP2aqHOMSA -----KTLFETLGAALFTLE-----β1
 OBP2bbHOMSA -----KTLFETLGAALFTLE-----β1
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 OBP2_RATNO -----MKSRLTVMILEMAVHGOEAPPDQ--ELLSKVTHTVTCENHTGKG--EAKGFMTVLTIER--DQVRIIFKKGHHLRSTH--DQVRIIFKKGHHLRSTH--KKTFTYKELIPVK
 LACB_BOSTA -----MKCLILATTCGAALIVTQMGGLDLCQV--EENSLIAASDISLSAGSALRVVPEELKPTPE--DEAILKOWENGELAAKQV--DAEIKLAVAKDALNENKVLVLDIDYK
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 RBP_HOMSA -----MKHWWALLLAAWAAERDCRVSSFRVKNFNGRTF--EAMAKKDDPEGLFLODN--IVAEEFDEMSATAGRVRLNNVA--EADAVGTFDDE--EAKKQYVAVASFLKGHDDH
 OBP_BOSTA -----MKMLLILCLGLTILGCVHAEESSTGRNLDVAJNDP--EISIVASHREKIDENGSGV--FPHQHDVLEN--SFGFRIKENGGERELYAVAY--SPDSEPFVEYDGNFTFLKTDYD
 MUP_RATNO -----MKMLLILCLGLTILGCVHAEESSTGRNLDVAJNDP--EISIVASHREKIDENGSGV--FPHQHDVLEN--SFGFRIKENGGERELYAVAY--SPDSEPFVEYDGNFTFLKTDYD
 OBP_SUSSC -----MKMLLILCLGLTILGCVHAEESSTGRNLDVAJNDP--EISIVASHREKIDENGSGV--FPHQHDVLEN--SFGFRIKENGGERELYAVAY--SPDSEPFVEYDGNFTFLKTDYD
 OBP2adHOMSA -----KMTLFLGVTLGLAALSTLLEEEDEGESVHPENPDADCGAWQIQRLWGQEAHIFAGAADGRLLRLLORPAPWGPALHGKACGICSLQGRAAVPTLHLATSPAGRNPTNLEALEEFK
 OBP2bgHOMSA -----KMTLFLGVTLGLAALSTLLEEEDEGESVHPENPDADCGAWQIQRLWGQEAHIFAGAAQEGPLHLLORPAPWGPAPHGKACG

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 OBP2baHOMSA -----β7
 OBP2abHOMSA -----β7
 OBP2aqHOMSA -----β7
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 LCN1_HOMSA -----β7
 OBP2_RATNO -----β7
 LACB_BOSTA -----β7
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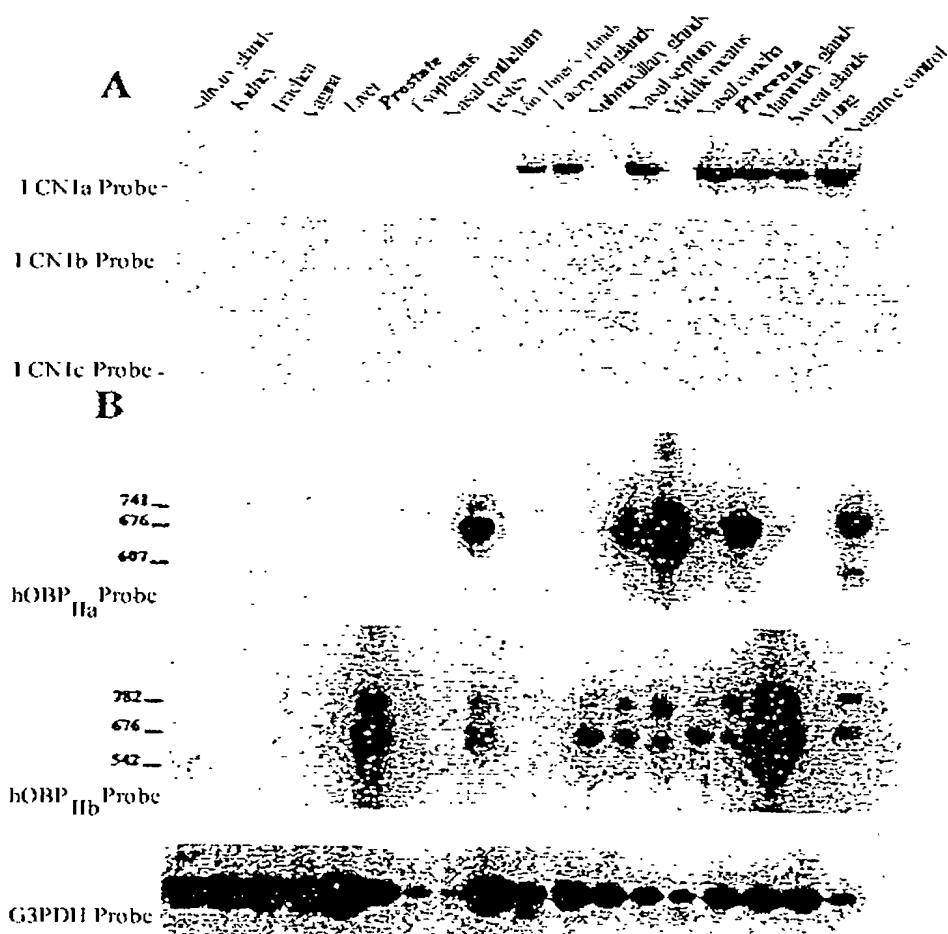
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 OBP2_RATNO -----β8
 LACB_BOSTA -----β8
 MUP6_MUSMU -----β8
 RBP_HOMSA -----β8
 OBP_BOSTA -----β8
 MUP_RATNO -----β8
 OBP_SUSSC -----β8

OBP2aaHOMSA -----β9
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 OBP_BOSTA -----β9
 MUP_RATNO -----β9
 OBP_SUSSC -----β9

OBP2adHOMSA -----KLVQRKGLSEEDIFMPLQSGCVLEH
 OBP2bgHOMSA -----KLVQRKGLSEEDIFMPLQSGCVLEH

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FIG. 6



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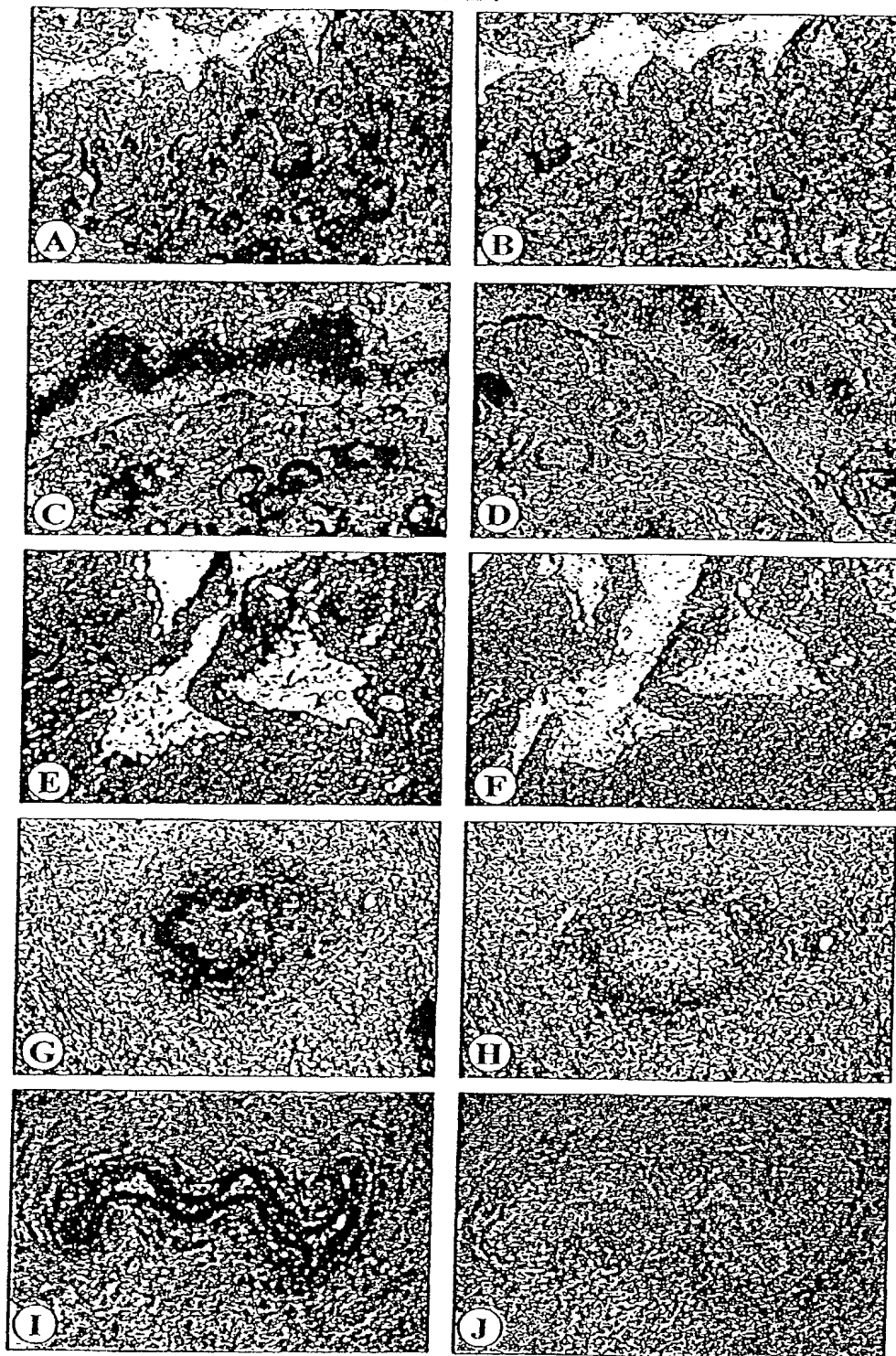


FIG. 7



FIG. 8

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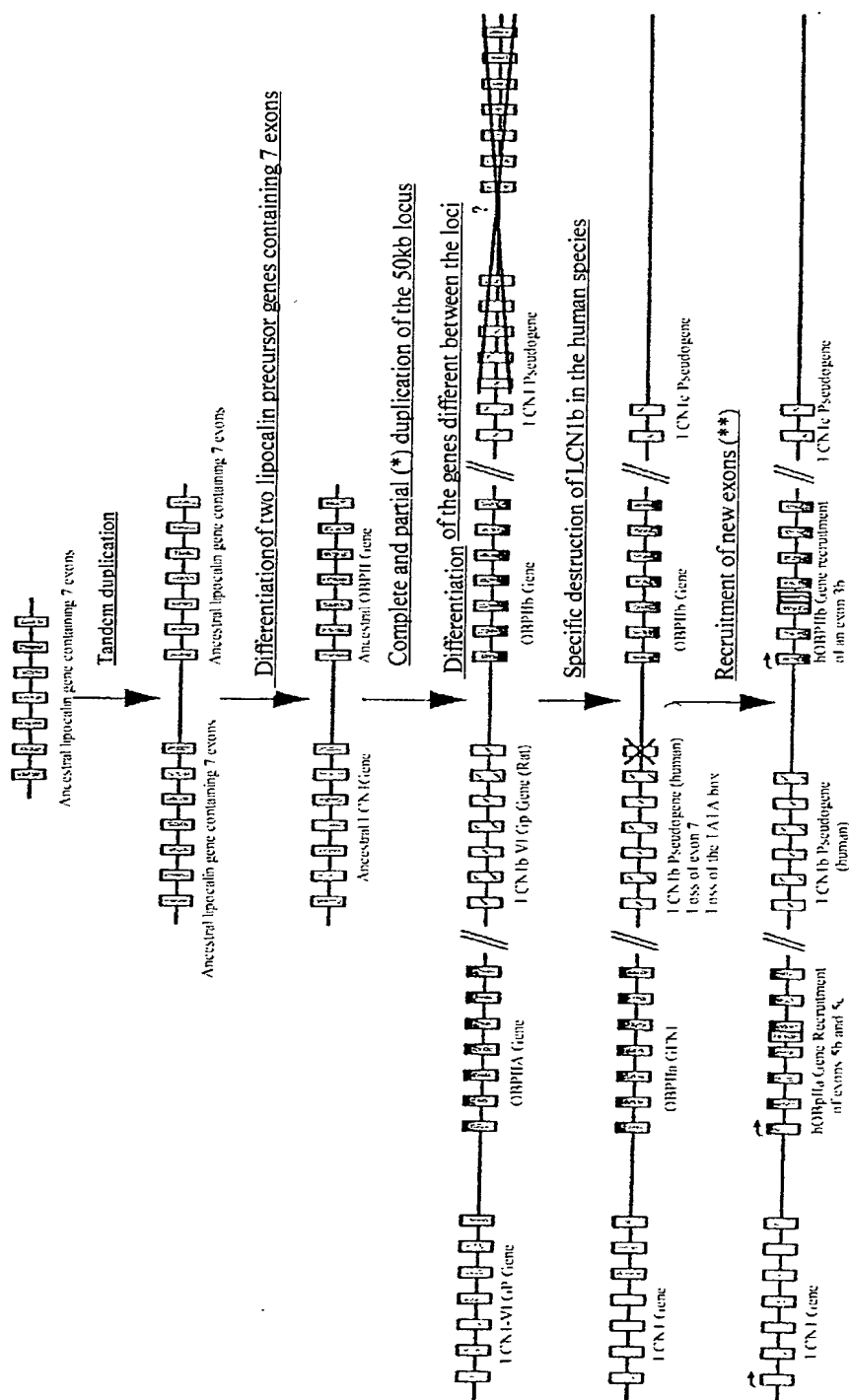


FIG.9

FIG. 10

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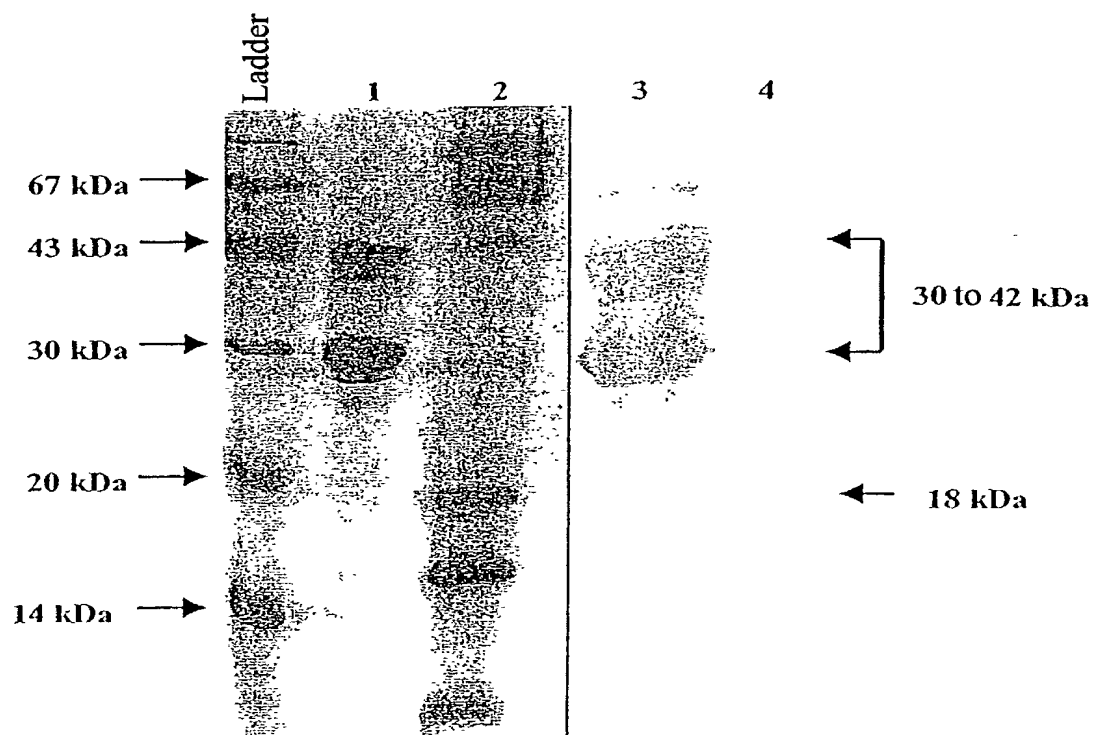


FIG. 11

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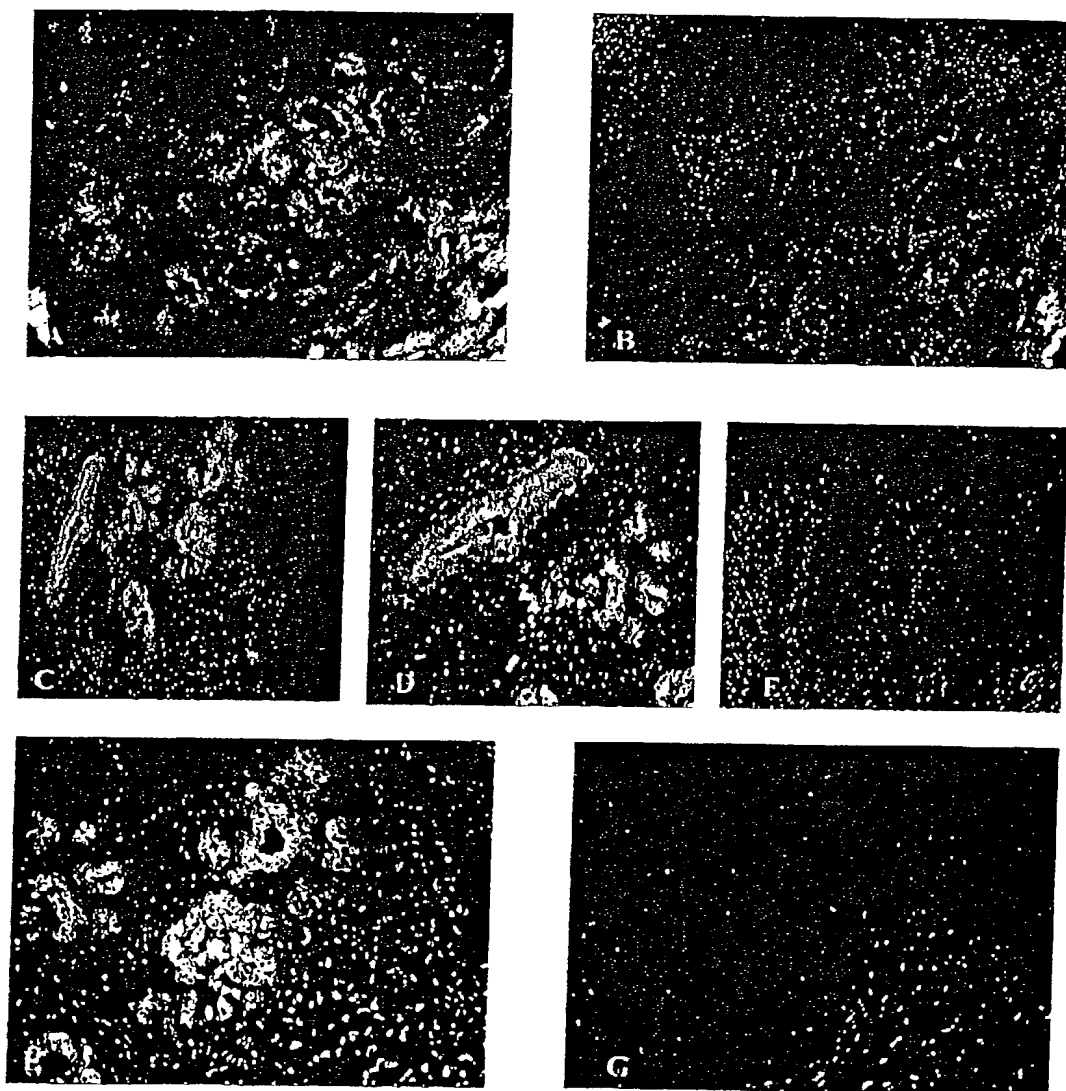


FIGURE 12

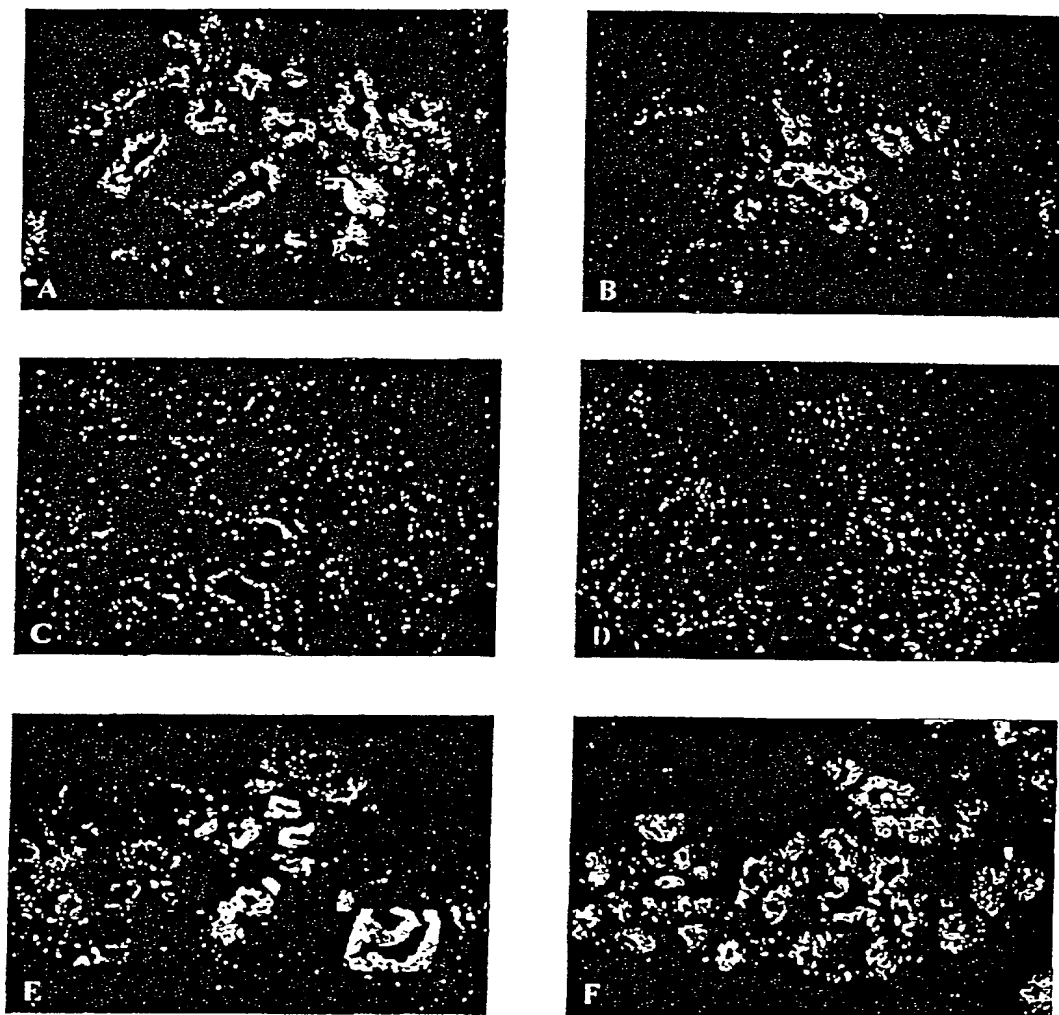


FIGURE 13

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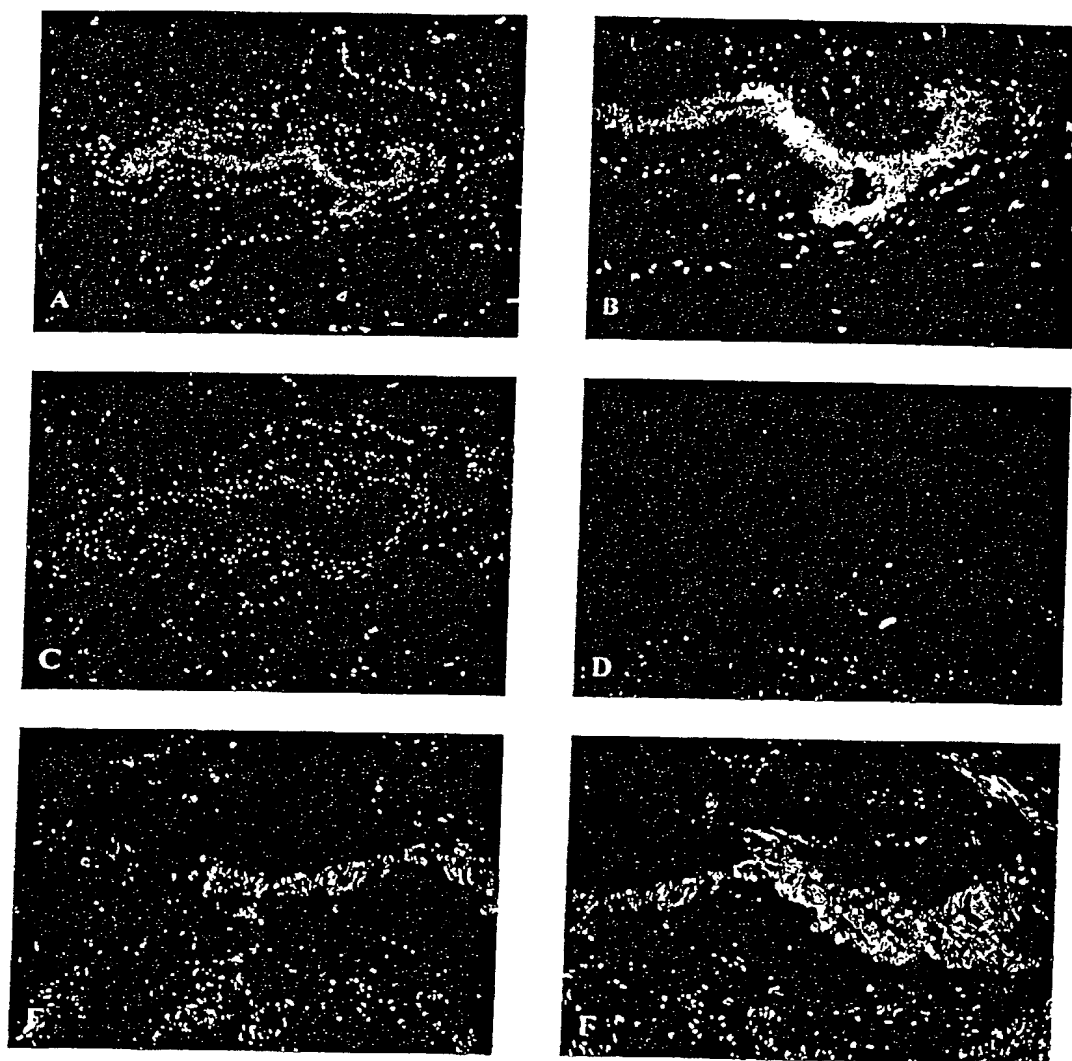


FIGURE 14

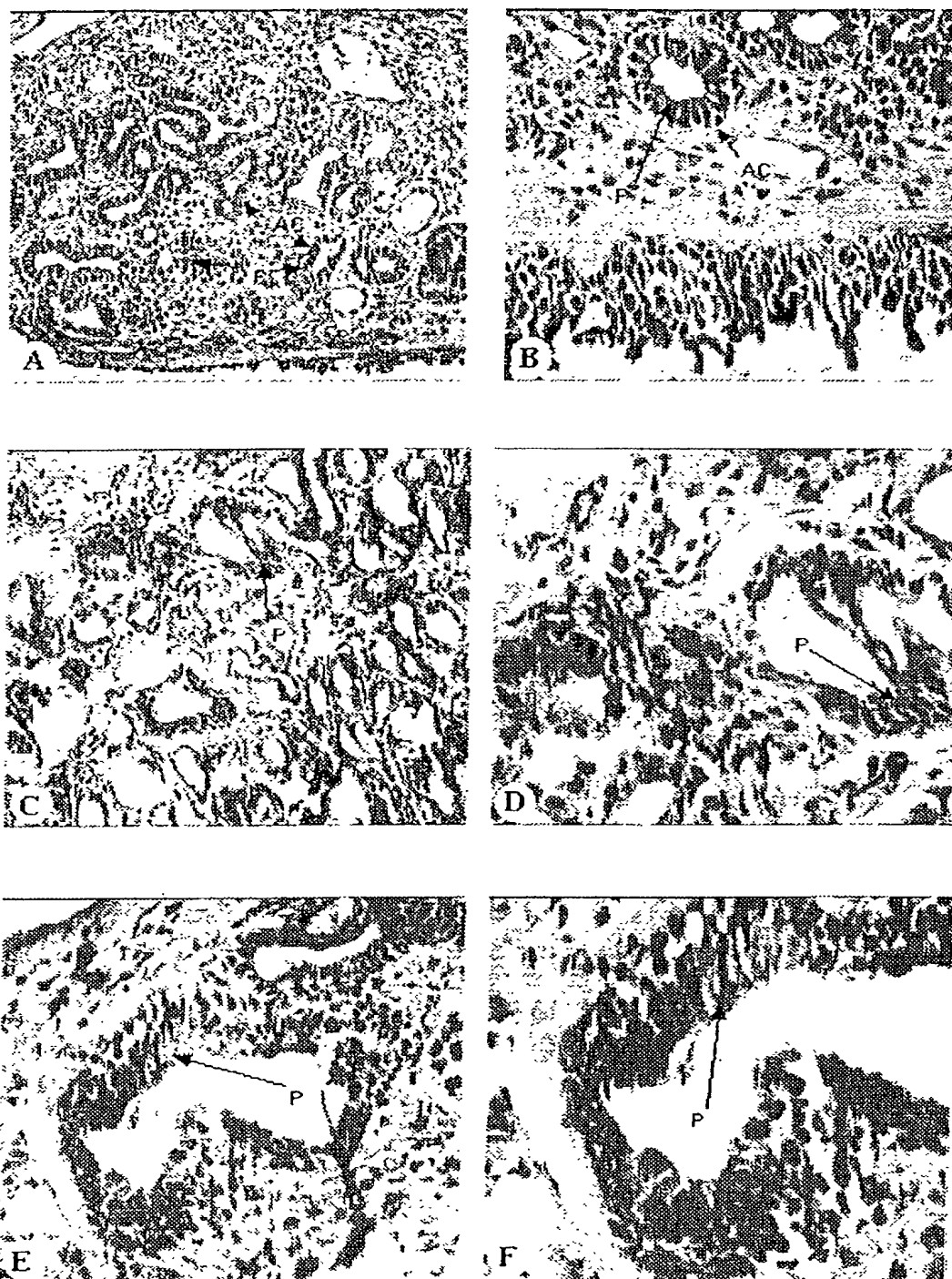


FIG. 15

FIG. 16

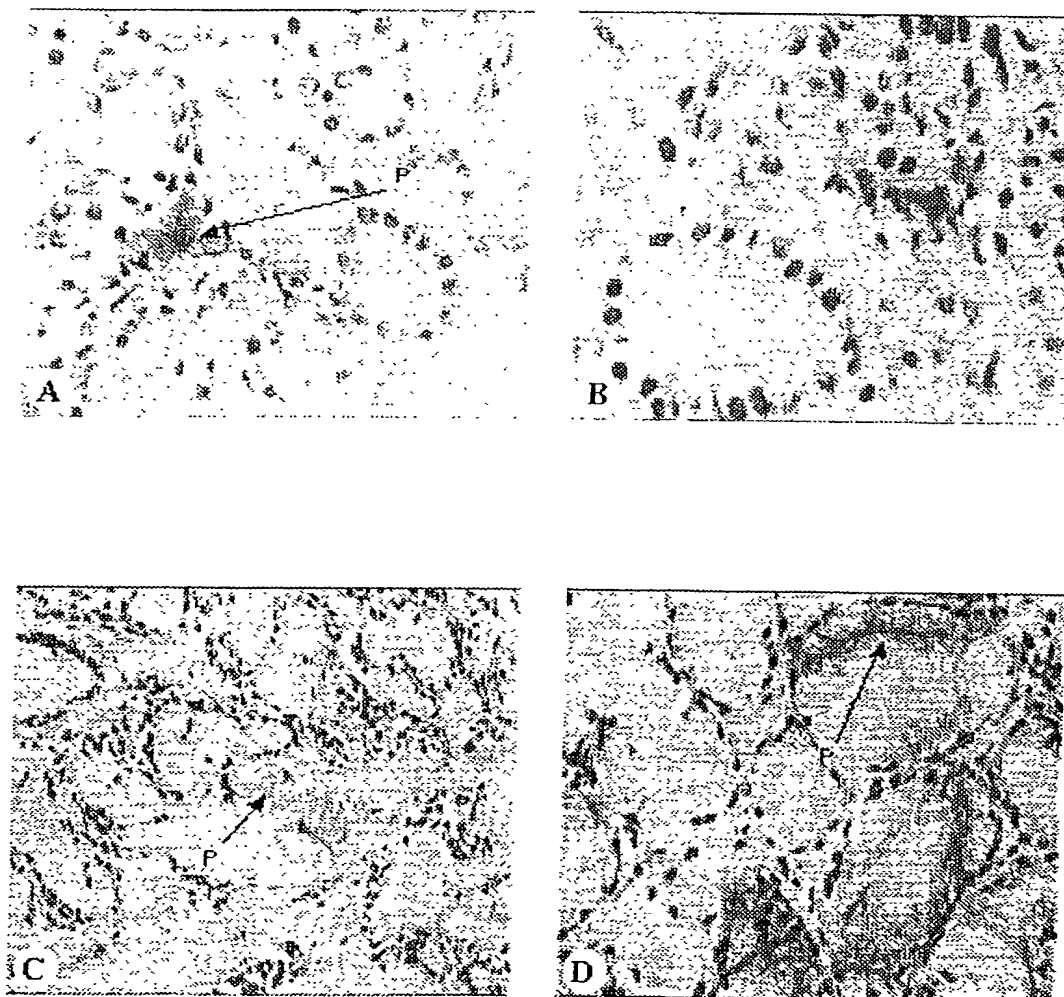


FIG. 17

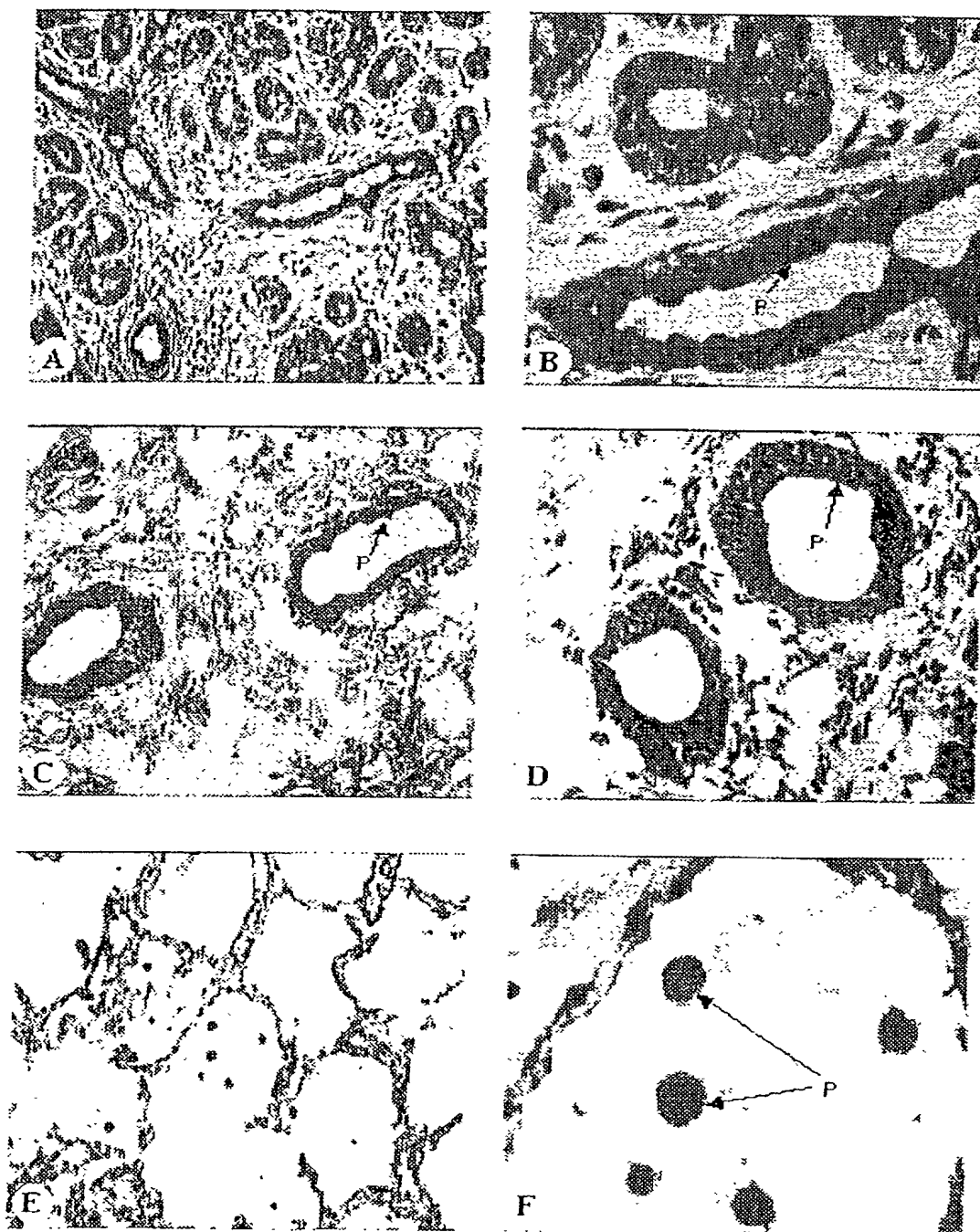


FIG. 18

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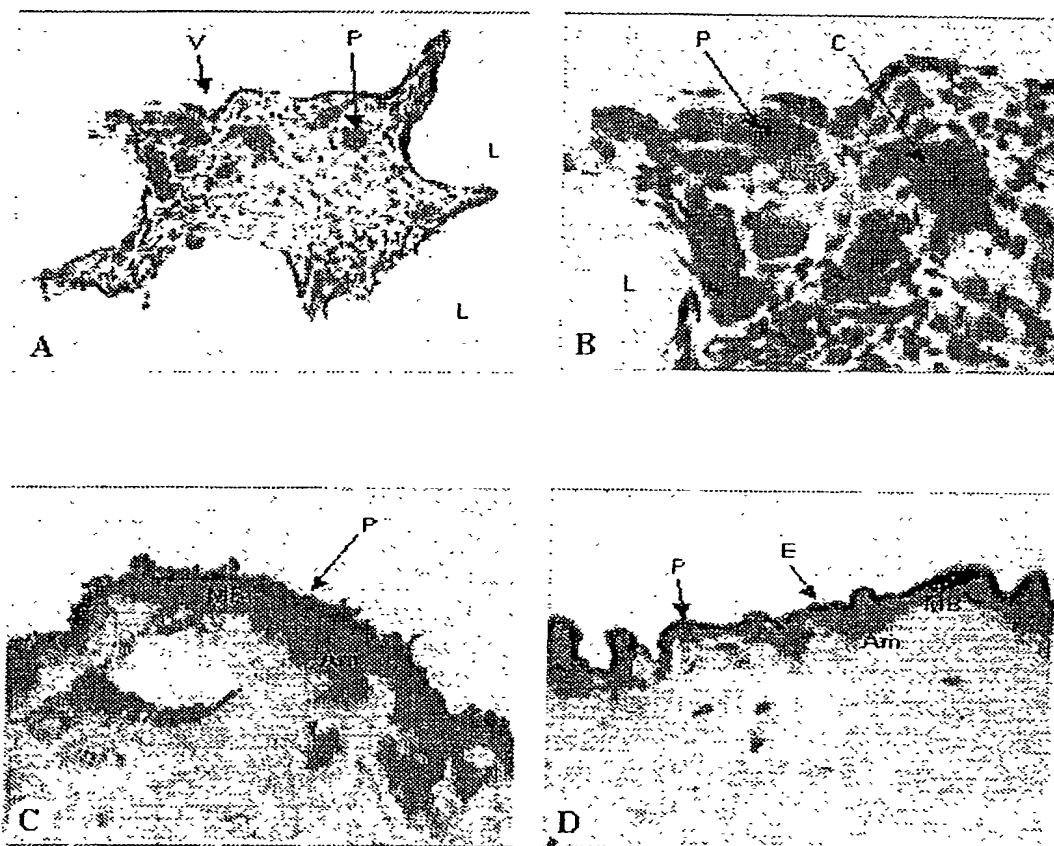


FIG. 19

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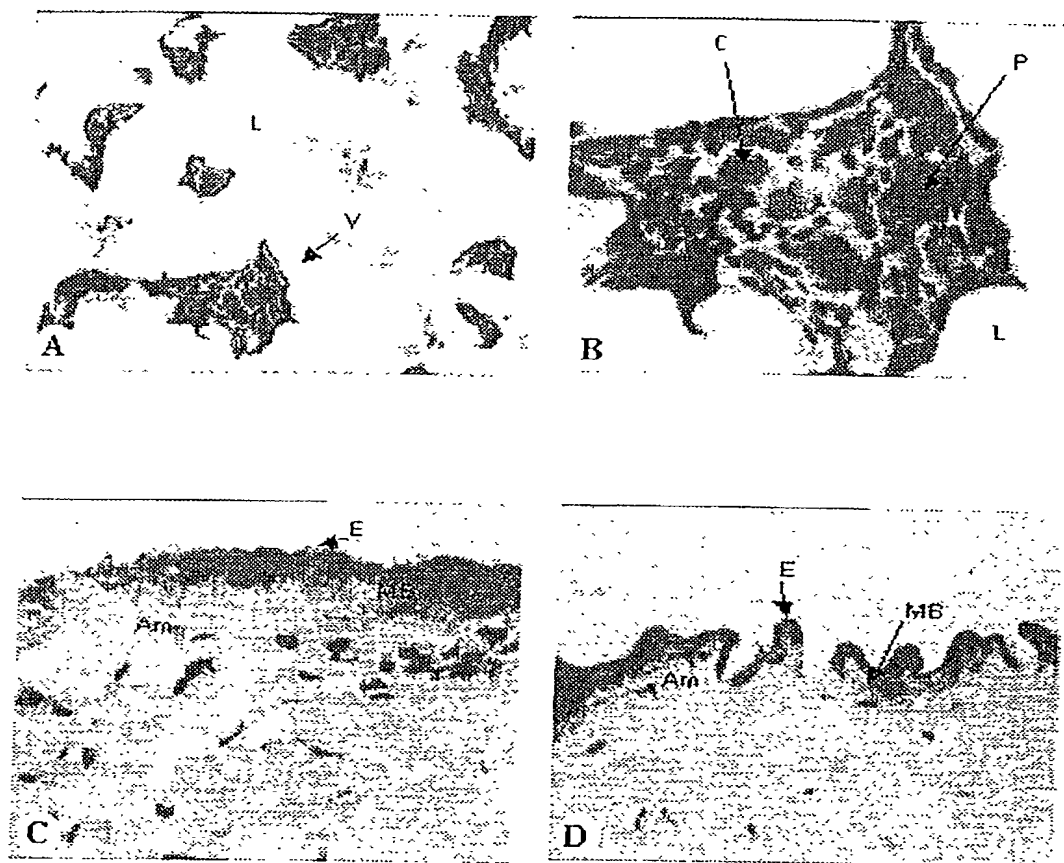


FIG. 20

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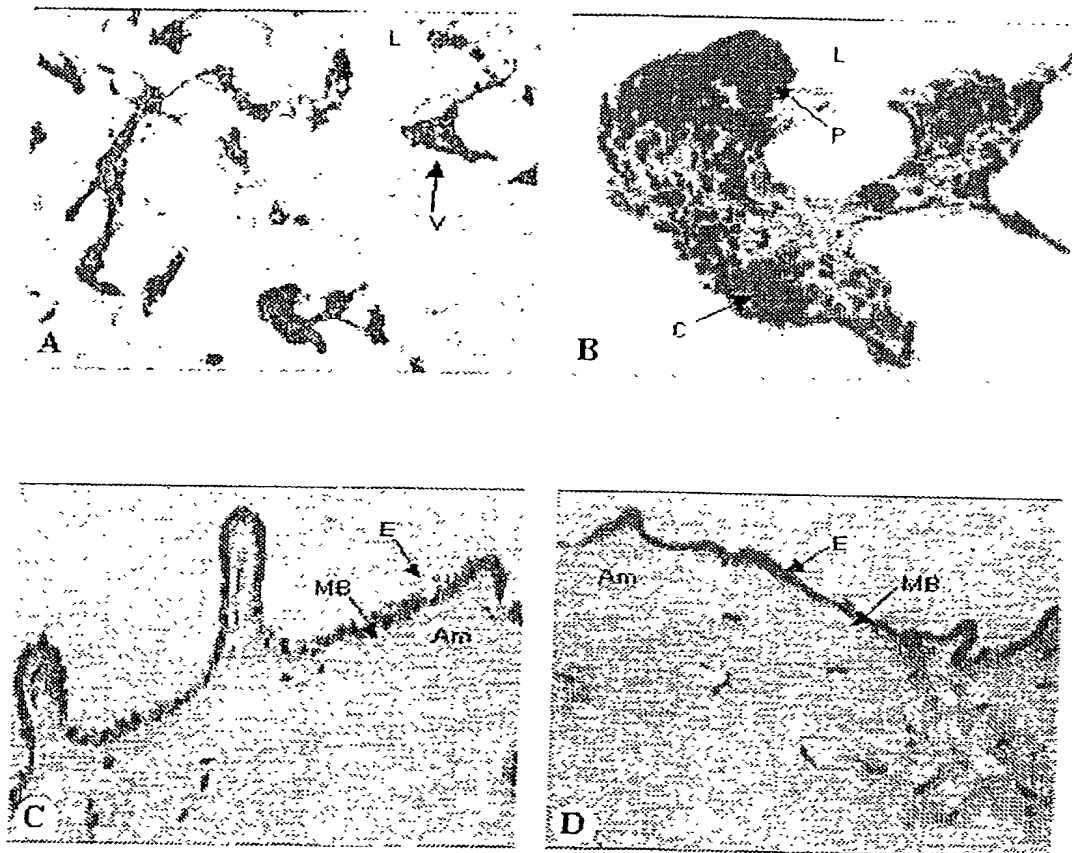


FIG. 21

DECLARATION AND POWER OF ATTORNEY

As a below named inventor, I HEREBY DECLARE:

THAT my residence, post office address, and citizenship are as stated below next to my name;

THAT I believe I am the original, first, and sole inventor (if only one inventor is named below) or an original, first, and joint inventor (if plural inventors are named below or in an attached Declaration) of the subject matter which is claimed and for which a patent is sought on the invention entitled **ODORANT-BINDING HUMAN PROTEINS HYDROPHOBIC LIGANDS: POLYPEPTIDES AND POLYNUCLEOTIDES CODING FOR SAID POLYPEPTIDES AND USES THEREOF**

(Attorney Docket No. 065691-0260)

the specification of which (check one)

_____ is attached hereto.

XX was filed on Aug. 11, 2000 as ~~United States Application Number~~ PCT International Application Number PCT/FR00/02319 and was amended on _____ (if applicable).

THAT I do not know and do not believe that the same invention was ever known or used by others in the United States of America, or was patented or described in any printed publication in any country, before I (we) invented it;

THAT I do not know and do not believe that the same invention was patented or described in any printed publication in any country, or in public use or on sale in the United States of America, for more than one year prior to the filing date of this United States application;

THAT I do not know and do not believe that the same invention was first patented or made the subject of an inventor's certificate that issued in any country foreign to the United States of America before the filing date of this United States application if the foreign application was filed by me (us), or by my (our) legal representatives or assigns, more than twelve months (six months for design patents) prior to the filing date of this United States application;

THAT I have reviewed and understand the contents of the above-identified specification, including the claim(s), as amended by any amendment specifically referred to above;

THAT I believe that the above-identified specification contains a written description of the invention, and of the manner and process of making and using it, in such full, clear, concise, and exact terms as to enable any person skilled in the art to which it pertains, or with which it is most nearly connected, to make and use the invention, and sets forth the best mode contemplated by me of carrying out the invention; and

THAT I acknowledge the duty to disclose to the U.S. Patent and Trademark Office all information known to me to be material to patentability as defined in Title 37, Code of Federal Regulations, §1.56.

I HEREBY CLAIM foreign priority benefits under Title 35, United States Code § 119(a)-(d) or § 365(b) of any foreign application(s) for patent or inventor's certificate, or § 365(a) of any PCT international application which designated at least one country other than the United States of America, listed below and have also identified below any foreign application for patent or inventor's certificate or of any PCT international application having a filing date before that of the application on which priority is claimed.

Prior Foreign Application Number	Country	Foreign Filing Date	Priority Claimed?	Certified Copy Attached?
99 10439	France	Aug. 12, 1999	yes	

I HEREBY CLAIM the benefit under Title 35, United States Code § 119(e) of any United States provisional application(s) listed below.

U.S. Provisional Application Number	Filing Date

I HEREBY CLAIM the benefit under Title 35, United States Code, § 120 of any United States application(s), or § 365(c) of any PCT international application designating the United States of America, listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States or PCT International application in the manner provided by the first paragraph of Title 35, United States Code, § 112, I acknowledge the duty to disclose information which is material to patentability as defined in Title 37, Code of Federal Regulations, § 1.56 which became available between the filing date of the prior application and the national or PCT international filing date of this application.

U.S. Parent Application Number	PCT Parent Application Number	Parent Filing Date	Parent Patent Number
	PCT/FR00/02319	August 11, 2000	

I HEREBY APPOINT the registered attorneys and agents at Customer Number 22428



22428

PATENT TRADEMARK OFFICE

to have full power to prosecute this application and any continuations, divisions, reissues, and reexaminations thereof, to receive the patent, and to transact all business in the United States Patent and Trademark Office connected therewith.

I request that all correspondence be directed to:

Stephen B. Maebius
FOLEY & LARDNER
Customer Number: 22428



22428

PATENT TRADEMARK OFFICE

Telephone: (202) 672-5569

Facsimile: (202) 672-5399

I UNDERSTAND AND AGREE THAT the foregoing attorneys and agents appointed by me to prosecute this application do not personally represent me or my legal interests, but instead represent the interests of the legal owner(s) of the invention described in this application.

I FURTHER DECLARE THAT all statements made herein of my own knowledge are true, and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code, and that such willful false statements may jeopardize the validity of the application or any patent issuing thereon.

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1-0

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Residence

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FRX

Citizenship

French

Post Office Address

The same as residence

Inventor's signature

Date

June 13, 2002

Name of second inventor

2-0

LACAZETTE Eric

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Citizenship

French

Ferrand / France

FRX

Post Office Address

The same as residence

Inventor's signature

Date

June 13, 2002

Name of third inventor

300 GACHON Francoise

Residence

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Citizenship

French

Post Office Address

The same as residence

Inventor's signature

Date

June 13, 2002

[Signature]

Name of fourth inventor

Residence

Citizenship

Post Office Address

Inventor's signature

Date

Attny's Docket No. _____

SMALL ENTITY DECLARATION
[37 CFR 1.9(c-f)]

Each undersigned declares that:

- (1) ☐ the application attached hereto
(2) ☐ U.S. Application Serial No. _____ filed _____
(3) ☐ U.S. Patent No. _____ Issued _____

is entitled to the benefits of "small entity" status for paying reduced fees under 35 USC 41(a) and (b) to the Patent and Trademark Office by virtue of the following:

(4) ☒ Each undersigned declares that he/she qualifies as an independent inventor, or would qualify had he/she made the invention, as defined in 37 CFR 1.9(c).

(5) ☐ The undersigned declares that he/she is an official empowered to act on behalf of the concern identified below; that this concern qualifies as a small business concern as defined in 37 CFR 1.9(d).

(6) ☐ The undersigned declares the he/she is an official empowered to act on behalf of the organization identified below; that this organization qualifies as a nonprofit organization as defined in

- (a) ☐ 37 CFR 1.9(e)(1)
(b) ☐ 37 CFR 1.9(e)(2)
(c) ☐ 37 CFR 1.9(e)(3)
(d) ☐ 37 CFR 1.9(e)(4)

State law of France

(7) Each person, concern or organization to which I/we have assigned, granted, conveyed or licensed, or am under an obligation under contract or law to assign, grant, convey, or license any rights in the invention is listed below:

- (a) ☐ no such person, concern or organization
(b) ☐ persons, concerns or organizations listed below

[a separate declaration is required from each named person, concern or organization having rights to this invention averring to their status as "small entities."]

Full Name PITOT Gilles

Address 151 rue du Chevaleret, 75013 Paris / France

☒ Individual ☐ Small Business Concern ☐ Nonprofit Organization

I/we acknowledge the duty to file, in this application or patent, notification of any change in status resulting in loss of entitlement of small entity status prior to paying, or at the time of paying, the earliest of the issue fee or any maintenance fee due after the date on which status as a small entity is no longer appropriate. (37 CFR 1.28(b))

I/we hereby declare all statements made herein of his/her own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application, any patent issued thereon or any patent to which this declaration is directed.

(8) PITOT Gilles June 13, 2002
Typed Name of Inventor Signature Date

Typed Name of Inventor Signature Date

(9) _____
Name of Small Business Concern or Nonprofit Organization

By _____
Typed Name Signature Date

Title of Signatory

SEQUENCE LISTING

<110> UNIVERSITE D'AUVERGNE CLERMONT I

<120> HUMAN ODORANT-BINDING PROTEINS WHICH BIND HYDROPHOBIC LIGANDS:
POLYPEPTIDES AND POLYNUCLEOTIDES ENCODING SAID POLYPEPTIDES, AND
USES THEREOF

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PCT/FR00/02319

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Asp	Lys	Asp	Phe	Pro	Glu	Asp	Arg	Arg	Pro	Arg	Lys	Val	Ser	Pro	Val	
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aag	gtg	aca	gcc	ctg	ggc	ggg	ggg	aac	ttg	gaa	gcc	acg	ttc	acc	ttc	246
Lys	Val	Thr	Ala	Leu	Gly	Gly	Gly	Asn	Leu	Glu	Ala	Thr	Phe	Thr	Phe	
			55				60					65				
atg	agg	gag	gat	cgg	tgc	atc	cag	aag	aaa	atc	ctg	atg	cgg	aag	acg	294
Met	Arg	Glu	Asp	Arg	Cys	Ile	Gln	Lys	Lys	Ile	Leu	Met	Arg	Lys	Thr	
	70					75					80					
gag	gag	cct	ggc	aaa	ttc	agc	gcc	tat	ggg	ggc	agg	aag	ctc	ata	tac	342
Glu	Glu	Pro	Gly	Lys	Phe	Ser	Ala	Tyr	Gly	Gly	Arg	Lys	Leu	Ile	Tyr	
85				90						95					100	
ctg	cag	gag	ctg	ccc	ggg	acg	gac	gac	tac	gtc	ttt	tac	tgc	aaa	gac	390
Leu	Gln	Glu	Leu	Pro	Gly	Thr	Asp	Asp	Tyr	Val	Phe	Tyr	Cys	Lys	Asp	
				105					110					115		
cag	cgc	cgt	ggg	ggc	ctg	cgc	tac	atg	gga	aag	ctt	gtg	gca	tct	gct	438
Gln	Arg	Arg	Gly	Gly	Leu	Arg	Tyr	Met	Gly	Lys	Leu	Val	Ala	Ser	Ala	
			120					125					130			
ccc	tgc	agg	gcc	gtg	ccg	ctg	tcc	cca	cgt	cgg	ctc	acc	tgg	cca	cct	486
Pro	Cys	Arg	Ala	Val	Pro	Leu	Ser	Pro	Arg	Arg	Leu	Thr	Trp	Pro	Pro	
		135					140					145				
cac	ctg	cag	gta	gga	atc	cta	ata	cca	acc	tgg	agg	ccc	tgg	aag	aat	534
His	Leu	Gln	Val	Gly	Ile	Leu	Ile	Pro	Thr	Trp	Arg	Pro	Trp	Lys	Asn	
			150			155					160					
tta	aga	aat	tgg	tgc	agc	aca	agg	gac	tct	cgg	agg	agg	aca	ttt	tca	582
Leu	Arg	Asn	Trp	Cys	Ser	Thr	Arg	Asp	Ser	Arg	Arg	Arg	Thr	Phe	Ser	
165				170						175					180	
tgc	ccc	tgc	aga	cgg	gaa	gct	gcg	ttc	tcg	aac	act	agg	cag	ccc	ccg	630
Cys	Pro	Cys	Arg	Arg	Glu	Ala	Ala	Phe	Ser	Asn	Thr	Arg	Gln	Pro	Pro	
				185					190					195		
ggg	ctg	cac	ctc	cag	agc	cca	ccc	tac	cac	cag	aca	cag	agc	ccg	gac	678
Gly	Leu	His	Leu	Gln	Ser	Pro	Pro	Tyr	His	Gln	Thr	Gln	Ser	Pro	Asp	
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8

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Glu Asn Pro Asp Ala Glu Asp Gly Gly Ala Trp Gln Ile Gln Arg Leu
35 40 45
Trp Gly Gln Glu Ala His Ile Pro Ala Gly Ala Ala Arg Asp Gly Arg
50 55 60
Leu Arg Leu Leu Leu Gln Arg Pro Ala Pro Trp Gly Pro Ala Leu His
65 70 75 80
Gly Lys Ala Cys Gly Ile Cys Ser Leu Gln Gly Arg Ala Ala Val Pro
85 90 95
Thr Leu Ala His Leu Ala Thr Ser Pro Ala Gly Arg Asn Pro Asn Thr
100 105 110
Asn Leu Glu Ala Leu Glu Glu Phe Lys Lys Leu Val Gln Arg Lys Gly
115 120 125
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130 135 140
Leu Glu His
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<212> DNA

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<220>

<221> CDS

<222> (43)..(552)

<223> cDNA2098 (676) - conventional form (hOBPIIb-alpha)

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Phe Leu Gly Val Thr Leu Gly Leu Ala Ala Ala Leu Ser Phe Thr Leu
5 10 15 20
gag gag gag gat atc aca ggg acc tgg tac gtg aag gcc atg gtg gtc 150
Glu Glu Glu Asp Ile Thr Gly Thr Trp Tyr Val Lys Ala Met Val Val
25 30 35
gat aag gac ttt ccg gag gac agg agg ccc agg aag gtg tcc cca gtg 198
Asp Lys Asp Phe Pro Glu Asp Arg Arg Pro Arg Lys Val Ser Pro Val

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aag gtg aca gcc ctg ggc ggt ggg aag ttg gaa gcc acg ttc acc ttc				246
Lys Val Thr Ala Leu Gly Gly Gly Lys Leu Glu Ala Thr Phe Thr Phe				
	55	60	65	
atg agg gag gat cgg tgc atc cag aag aaa atc ctg atg cgg aag acg				294
Met Arg Glu Asp Arg Cys Ile Gln Lys Lys Ile Leu Met Arg Lys Thr				
	70	75	80	
gag gag cct ggc aaa tac agc gcc tat ggg ggc agg aag ctc atg tac				342
Glu Glu Pro Gly Lys Tyr Ser Ala Tyr Gly Gly Arg Lys Leu Met Tyr				
	85	90	95	100
ctg cag gag ctg ccc agg agg gac cac tac atc ttt tac tgc aaa gac				390
Leu Gln Glu Leu Pro Arg Arg Asp His Tyr Ile Phe Tyr Cys Lys Asp				
	105	110	115	
cag cac cat ggg ggc ctg ctc cac atg gga aag ctt gtg ggt agg aat				438
Gln His His Gly Gly Leu Leu His Met Gly Lys Leu Val Gly Arg Asn				
	120	125	130	
tct gat acc aac cgg gag gcc ctg gaa gaa ttt aag aaa ttg gtg cag				486
Ser Asp Thr Asn Arg Glu Ala Leu Glu Glu Phe Lys Lys Leu Val Gln				
	135	140	145	
cgc aag gga ctc tgc gag gag gac att ttc acg ccc ctg cag acg gga				534
Arg Lys Gly Leu Ser Glu Glu Asp Ile Phe Thr Pro Leu Gln Thr Gly				
	150	155	160	
agc tgc gtt ccc gaa cac taggcagccc ccgggtctgc acctccagag				582
Ser Cys Val Pro Glu His				
	165	170		
cccaccctac caccagacac agagcccgga ccacctggac ctaccctcca gccatgaccc				642
ttccctgctc ccaccacact gactccaaat aaag				676
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Ala Met Val Val Asp Lys Asp Phe Pro Glu Asp Arg Arg Pro Arg Lys				
35 40 45				
Val Ser Pro Val Lys Val Thr Ala Leu Gly Gly Gly Lys Leu Glu Ala				
50 55 60				
Thr Phe Thr Phe Met Arg Glu Asp Arg Cys Ile Gln Lys Lys Ile Leu				

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65	70	75	80
Met Arg Lys Thr Glu Pro Gly Lys Tyr Ser Ala Tyr Gly Gly Arg			
	85	90	95
Lys Leu Met Tyr Leu Gln Glu Leu Pro Arg Arg Asp His Tyr Ile Phe			
	100	105	110
Tyr Cys Lys Asp Gln His His Gly Gly Leu Leu His Met Gly Lys Leu			
	115	120	125
Val Gly Arg Asn Ser Asp Thr Asn Arg Glu Ala Leu Glu Glu Phe Lys			
	130	135	140
Lys Leu Val Gln Arg Lys Gly Leu Ser Glu Glu Asp Ile Phe Thr Pro			
	145	150	155
			160
Leu Gln Thr Gly Ser Cys Val Pro Glu His			
	165	170	

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 <212> DNA
 <213> Homo sapiens

 <220>
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 <222> (1)..(537)
 <223> cDNA2098 (782) - long form (hOBPIIb-beta)

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 Arg Pro Val Thr Cys Arg Gly Arg Gln His Arg Ala Leu Glu Met Lys
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 acc ctg ttc ctg ggt gtc acg ctc ggc ctg gcc gct gcc ctg tcc ttc 96
 Thr Leu Phe Leu Gly Val Thr Leu Gly Leu Ala Ala Ala Leu Ser Phe
 20 25 30

 acc ctg gag gag gag gat atc aca ggg acc tgg tac gtg aag gcc atg 144
 Thr Leu Glu Glu Glu Asp Ile Thr Gly Thr Trp Tyr Val Lys Ala Met
 35 40 45

 gtg gtc gat aag gac ttt ccg gag gac agg agg ccc agg aag gtg tcc 192
 Val Val Asp Lys Asp Phe Pro Glu Asp Arg Arg Pro Arg Lys Val Ser
 50 55 60

 cca gtg aag gtg aca gcc ctg ggc ggt ggg aag ttg gaa gcc acg ttc 240
 Pro Val Lys Val Thr Ala Leu Gly Gly Gly Lys Leu Glu Ala Thr Phe
 65 70 75 80

 acc ttc atg agg gag gat cgg tgc atc cag aag aaa atc ctg atg cgg 288
 Thr Phe Met Arg Glu Asp Arg Cys Ile Gln Lys Lys Ile Leu Met Arg
 85 90 95

 aag acg gag gag cct ggc aaa tac agc gcc tgc ttg tcc gca gtc gag 336
 Lys Thr Glu Glu Pro Gly Lys Tyr Ser Ala Cys Leu Ser Ala Val Glu

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11

100 105 110

atg gac cag atc acg cct gcc ctg tgg gag gcc cta gcc att gac aca 384
Met Asp Gln Ile Thr Pro Ala Leu Trp Glu Ala Leu Ala Ile Asp Thr
115 120 125

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Leu Arg Lys Leu Arg Ile Gly Thr Arg Arg Pro Arg Ile Arg Trp Gly
130 135 140

cag gaa gct cat gta cct gca gga gct gcc cag gag gga cca cta cat 480
Gln Glu Ala His Val Pro Ala Gly Ala Ala Gln Glu Gly Pro Leu His
145 150 155 160

ctt tta ctg caa aga cca gca cca tgg ggg cct gct cca cat ggg aaa 528
Leu Leu Leu Gln Arg Pro Ala Pro Trp Gly Pro Ala Pro His Gly Lys
165 170 175

gct tgt ggg taggaattct gataccaacc gggaggccct ggaagaattt 577
Ala Cys Gly

aagaaattgg tgcagcgcaa gggactctcg gaggaggaca ttttcacgcc cctgcagacg 637

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cccacccacc tgactccaaa taaag 782

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20 25 30

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35 40 45

Val Val Asp Lys Asp Phe Pro Glu Asp Arg Arg Pro Arg Lys Val Ser
50 55 60

Pro Val Lys Val Thr Ala Leu Gly Gly Gly Lys Leu Glu Ala Thr Phe
65 70 75 80

Thr Phe Met Arg Glu Asp Arg Cys Ile Gln Lys Lys Ile Leu Met Arg
85 90 95

Lys Thr Glu Glu Pro Gly Lys Tyr Ser Ala Cys Leu Ser Ala Val Glu
100 105 110

Met Asp Gln Ile Thr Pro Ala Leu Trp Glu Ala Leu Ala Ile Asp Thr

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 35 40 45
 Trp Gly Gln Glu Ala His Ile Pro Ala Gly Ala Ala Gln Glu Gly Pro
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 Gly Lys Ala Cys Gly
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<220>
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15

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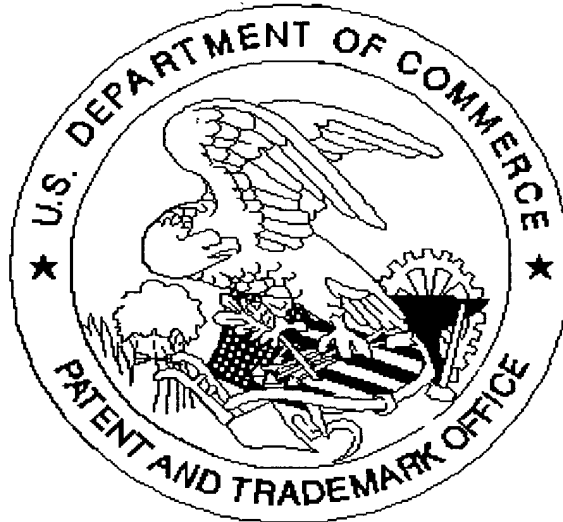
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